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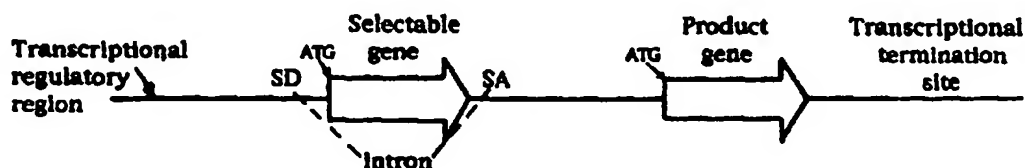
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(54) Title: METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLS



(57) Abstract

A method for selecting recombinant host cells expressing high levels of a desired protein is described. This method utilizes eukaryotic host cells harboring a DNA construct comprising a selectable gene (preferably an amplifiable gene) and a product gene provided 3' to the selectable gene. The selectable gene is positioned within an intron defined by a splice donor site and a splice acceptor site and the selectable gene and product gene are under the transcriptional control of a single transcriptional regulatory region. The splice donor site is generally an efficient splice donor site and thereby regulates expression of the product gene using the transcriptional regulatory region. The transfected cells are cultured so as to express the gene encoding the product in a selective medium comprising an amplifying agent for sufficient time to allow amplification to occur, whereupon either the desired product is recovered or cells having multiple copies of the product gene are identified.

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METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLSBACKGROUND OF THE INVENTIONField of the Invention

This invention relates to a method of selecting for high-expressing
5 host cells, a method of producing a protein of interest in high yields and
a method of producing eukaryotic cells having multiple copies of a sequence
encoding a protein of interest.

Description of Background and Related Art

The discovery of methods for introducing DNA into living host cells
10 in a functional form has provided the key to understanding many fundamental
biological processes, and has made possible the production of important
proteins and other molecules in commercially useful quantities.

Despite the general success of such gene transfer methods, several
common problems exist that may limit the efficiency with which a gene
15 encoding a desired protein can be introduced into and expressed in a host
cell. One problem is knowing when the gene has been successfully
transferred into recipient cells. A second problem is distinguishing
between those cells that contain the gene and those that have survived the
transfer procedures but do not contain the gene. A third problem is
20 identifying and isolating those cells that contain the gene and that are
expressing high levels of the protein encoded by the gene.

In general, the known methods for introducing genes into eukaryotic
cells tend to be highly inefficient. Of the cells in a given culture, only
a small proportion take up and express exogenously added DNA, and an even
25 smaller proportion stably maintain that DNA.

Identification of those cells that have incorporated a product gene
encoding a desired protein typically is achieved by introducing into the
same cells another gene, commonly referred to as a selectable gene, that
encodes a selectable marker. A selectable marker is a protein that is
30 necessary for the growth or survival of a host cell under the particular
culture conditions chosen, such as an enzyme that confers resistance to an
antibiotic or other drug, or an enzyme that compensates for a metabolic or
catabolic defect in the host cell. For example, selectable genes commonly
used with eukaryotic cells include the genes for aminoglycoside
35 phosphotransferase (APH), hygromycin phosphotransferase (hyg),
dihydrofolate reductase (DHFR), thymidine kinase (tk), neomycin, puromycin,
glutamine synthetase, and asparagine synthetase.

The method of identifying a host cell that has incorporated one gene
on the basis of expression by the host cell of a second incorporated gene
40 encoding a selectable marker is referred to as cotransfection (or
cotransfection). In that method, a gene encoding a desired polypeptide and
a selection gene typically are introduced into the host cell
simultaneously, although they may be introduced sequentially. In the case
of simultaneous cotransfection, the gene encoding the desired polypeptide

and the selectable gene may be present on a single DNA molecule or on separate DNA molecules prior to being introduced into the host cells. Wigler et al., Cell, 16:777 (1979). Cells that have incorporated the gene encoding the desired polypeptide then are identified or isolated by
5 culturing the cells under conditions that preferentially allow for the growth or survival of those cells that synthesize the selectable marker encoded by the selectable gene.

The level of expression of a gene introduced into a eukaryotic host cell depends on multiple factors, including gene copy number, efficiency
10 of transcription, messenger RNA (mRNA) processing, stability, and translation efficiency. Accordingly, high level expression of a desired polypeptide typically will involve optimizing one or more of those factors.

For example, the level of protein production may be increased by covalently joining the coding sequence of the gene to a "strong" promoter
15 or enhancer that will give high levels of transcription. Promoters and enhancers are nucleotide sequences that interact specifically with proteins in a host cell that are involved in transcription. Kriegler, Meth. Enzymol., 185:512 (1990); Maniatis et al., Science, 236:1237 (1987). Promoters are located upstream of the coding sequence of a gene and
20 facilitate transcription of the gene by RNA polymerase. Among the eukaryotic promoters that have been identified as strong promoters for high-level expression are the SV40 early promoter, adenovirus major late promoter, mouse metallothionein-I promoter, Rous sarcoma virus long terminal repeat, and human cytomegalovirus immediate early promoter (CMV).

Enhancers stimulate transcription from a linked promoter. Unlike
25 promoters, enhancers are active when placed downstream from the transcription initiation site or at considerable distances from the promoter, although in practice enhancers may overlap physically and functionally with promoters. For example, all of the strong promoters
30 listed above also contain strong enhancers. Bendig, Genetic Engineering, 7:91 (Academic Press, 1988).

The level of protein production also may be increased by increasing the gene copy number in the host cell. One method for obtaining high gene copy number is to directly introduce into the host cell multiple copies of
35 the gene, for example, by using a large molar excess of the product gene relative to the selectable gene during cotransfection. Kaufman, Meth. Enzymol., 185:537 (1990). With this method, however, only a small proportion of the cotransfected cells will contain the product gene at high copy number. Furthermore, because no generally applicable, convenient
40 method exists for distinguishing such cells from the majority of cells that contain fewer copies of the product gene, laborious and time-consuming screening methods typically are required to identify the desired high-copy number transfectants.

Another method for obtaining high gene copy number involves cloning
45 the gene in a vector that is capable of replicating autonomously in the host cell. Examples of such vectors include mammalian expression vectors

derived from Epstein-Barr virus or bovine papilloma virus, and yeast 2-micron plasmid vectors. Stephens & Hentschel, Biochem. J., 248:1 (1987); Yates et al., Nature, 313:812 (1985); Beggs, Genetic Engineering, 2:175 (Academic Press, 1981).

5 Yet another method for obtaining high gene copy number involves gene amplification in the host cell. Gene amplification occurs naturally in eukaryotic cells at a relatively low frequency. Schimke, J. Biol. Chem., 263:5989 (1988). However, gene amplification also may be induced, or at least selected for, by exposing host cells to appropriate selective
10 pressure. For example, in many cases it is possible to introduce a product gene together with an amplifiable gene into a host cell and subsequently select for amplification of the marker gene by exposing the cotransfected cells to sequentially increasing concentrations of a selective agent. Typically the product gene will be coamplified with the marker gene under
15 such conditions.

The most widely used amplifiable gene for that purpose is a DHFR gene, which encodes a dihydrofolate reductase enzyme. The selection agent used in conjunction with a DHFR gene is methotrexate (Mtx). A host cell is cotransfected with a product gene encoding a desired protein and a DHFR
20 gene, and transfectants are identified by first culturing the cells in culture medium that contains Mtx. A suitable host cell when a wild-type DHFR gene is used is the Chinese Hamster Ovary (CHO) cell line deficient in DHFR activity, prepared and propagated as described by Urlaub & Chasin, Proc. Nat. Acad. Sci. USA, 77:4216 (1980). The transfected cells then are
25 exposed to successively higher amounts of Mtx. This leads to the synthesis of multiple copies of the DHFR gene, and concomitantly, multiple copies of the product gene. Schimke, J. Biol. Chem., 263:5989 (1988); Axel et al., U.S. Patent No. 4,399,216; Axel et al., U.S. Patent No. 4,634,665. Other references directed to co-transfection of a gene together with a genetic
30 marker that allows for selection and subsequent amplification include Kaufman in Genetic Engineering, ed. J. Setlow (Plenum Press, New York), Vol. 9 (1987); Kaufman and Sharp, J. Mol. Biol., 159:601 (1982); Ringold et al., J. Mol. Appl. Genet., 1:165-175 (1981); Kaufman et al., Mol. Cell Biol., 5:1750-1759 (1985); Kaetzel and Nilson, J. Biol. Chem., 263:6244-
35 6251 (1988); Hung et al., Proc. Natl. Acad. Sci. USA, 83:261-264 (1986); Kaufman et al., EMBO J., 6:87-93 (1987); Johnston and Kucey, Science, 242:1551-1554 (1988); Urlaub et al., Cell, 33:405-412 (1983).

To extend the DHFR amplification method to other cell types, a mutant DHFR gene that encodes a protein with reduced sensitivity to methotrexate
40 may be used in conjunction with host cells that contain normal numbers of an endogenous wild-type DHFR gene. Simonsen and Levinson, Proc. Natl. Acad. Sci. USA, 80:2495 (1983); Wigler et al., Proc. Natl. Acad. Sci. USA, 77:3567-3570 (1980); Haber and Schimke, Somatic Cell Genetics, 8:499-508 (1982).

45 Alternatively, host cells may be co-transfected with the product gene, a DHFR gene, and a dominant selectable gene, such as a neo^r gene. Kim

and Wold, Cell, 42:129 (1985); Capon et al., U.S. Pat. No. 4,965,199. Transfectants are identified by first culturing the cells in culture medium containing neomycin (or the related drug G418), and the transfectants so identified then are selected for amplification of the DHFR gene and the product gene by exposure to successively increasing amounts of Mtx.

As will be appreciated from this discussion, the selection of recombinant host cells that express high levels of a desired protein generally is a multi-step process. In the first step, initial transfectants are selected that have incorporated the product gene and the selectable gene. In subsequent steps, the initial transfectants are subject to further selection for high-level expression of the selectable gene and then random screening for high-level expression of the product gene. To identify cells expressing high levels of the desired protein, typically one must screen large numbers of transfectants. The majority of transfectants produce less than maximal levels of the desired protein. Further, Mtx resistance in DHFR transformants is at least partially conferred by varying degrees of gene amplification. Schimke, Cell, 37:705-713 (1984). The inadequacies of co-expression of the non-selected gene have been reported by Wold et al., Proc. Natl. Acad. Sci. USA, 76:5684-5688 (1979). Instability of the amplified DNA is reported by Kaufman and Schimke, Mol. Cell Biol., 1:1069-1076 (1981); Haber and Schimke, Cell, 26:355-362 (1981); and Fedespiel et al., J. Biol. Chem., 259:9127-9140 (1984).

Several methods have been described for directly selecting such recombinant host cells in a single step. One strategy involves co-transfecting host cells with a product gene and a DHFR gene, and selecting those cells that express high levels of DHFR by directly culturing in medium containing a high concentration of Mtx. Many of the cells selected in that manner also express the co-transfected product gene at high levels. Page and Sydenham, Bio/Technology, 9:64 (1991). This method for single-step selection suffers from certain drawbacks that limit its usefulness. High-expressing cells obtained by direct culturing in medium containing a high level of a selection agent may have poor growth and stability characteristics, thus limiting their usefulness for long-term production processes. Page and Snyderman, Bio/Technology, 9:64 (1991). Single-step selection for high-level resistance to Mtx may produce cells with an altered, Mtx-resistant DHFR enzyme, or cells that have altered Mtx transport properties, rather than cells containing amplified genes. Haber et al., J. Biol. Chem., 256:9501 (1981); Assaraf and Schimke, Proc. Natl. Acad. Sci. USA, 84:7154 (1987).

Another method involves the use of polycistronic mRNA expression vectors containing a product gene at the 5' end of the transcribed region and a selectable gene at the 3' end. Because translation of the selectable gene at the 3' end of the polycistronic mRNA is inefficient, such vectors exhibit preferential translation of the product gene and require high levels of polycistronic mRNA to survive selection. Kaufman, Meth.

Enzymol., 185:487 (1990); Kaufman, Meth. Enzymol., 185:537 (1990); Kaufman et al., EMBO J., 6:187 (1987). Accordingly, cells expressing high levels of the desired protein product may be obtained in a single step by culturing the initial transfectants in medium containing a selection agent appropriate for use with the particular selectable gene. However, the utility of these vectors is variable because of the unpredictable influence of the upstream product reading frame on selectable marker translation and because the upstream reading frame sometimes becomes deleted during methotrexate amplification (Kaufman et al., J. Mol. Biol., 159:601-621 [1982]; Levinson, Methods in Enzymology, San Diego: Academic Press, Inc. [1990]). Later vectors incorporated an internal translation initiation site derived from members of the picornavirus family which is positioned between the product gene and the selectable gene (Pelletier et al., Nature, 334:320 [1988]; Jang et al., J. Virol., 63:1651 [1989]).

A third method for single-step selection involves use of a DNA construct with a selectable gene containing an intron within which is located a gene encoding the protein of interest. See U.S. Patent No. 5,043,270 and Abrams et al., J. Biol. Chem., 264(24): 14016-14021 (1989). In yet another single-step selection method, host cells are co-transfected with an intron-modified selectable gene and a gene encoding the protein of interest. See WO 92/17566, published October 15, 1992. The intron-modified gene is prepared by inserting into the transcribed region of a selectable gene an intron of such length that the intron is correctly spliced from the corresponding mRNA precursor at low efficiency, so that the amount of selectable marker produced from the intron-modified selectable gene is substantially less than that produced from the starting selectable gene. These vectors help to insure the integrity of the integrated DNA construct, but transcriptional linkage is not achieved as selectable gene and the protein gene are driven by separate promoters.

Other mammalian expression vectors that have single transcription units have been described. Retroviral vectors have been constructed (Cepko et al., Cell, 37:1053-1062 [1984]) in which a cDNA is inserted between the endogenous Moloney murine leukemia virus (M-MuLV) splice donor and splice acceptor sites which are followed by a neomycin resistance gene. This vector has been used to express a variety of gene products following retroviral infection of several cell types.

With the above drawbacks in mind, it is one object of the present invention to increase the level of homogeneity with regard to expression levels of stable clones transfected with a product gene of interest, by expressing a selectable marker (DHFR) and the protein of interest from a single promoter.

It is another object to provide a method for selecting stable, recombinant host cells that express high levels of a desired protein product, which method is rapid and convenient to perform, and reduces the numbers of transfected cells which need to be screened. Furthermore, it is

an object to allow high levels of single and two unit polypeptides to be rapidly generated from clones or pools of stable host cell transfectants.

It is an additional object to provide expression vectors which bias for active integration events (i.e. have an increased tendency to generate transformants wherein the DNA construct is inserted into a region of the genome of the host cell which results in high level expression of the product gene) and can accommodate a variety of product genes without the need for modification.

10 SUMMARY OF THE INVENTION

Accordingly, the present invention is directed to a DNA construct (DNA molecule) alternative terminology comprising a 5' transcriptional initiation site and a 3' transcriptional termination site, a selectable gene (preferably an amplifiable gene) and a product gene provided 3' to the selectable gene, a transcriptional regulatory region regulating transcription of both the selectable gene and the product gene, the selectable gene positioned within an intron defined by a splice donor site and a splice acceptor site. The splice donor site preferably comprises an effective splice donor sequence as herein defined and thereby regulates expression of the product gene using the transcriptional regulatory region.

20 In another embodiment, the invention provides a method for producing a product of interest comprising culturing a eukaryotic cell which has been transfected with the DNA construct described above, so as to express the product gene and recovering the product.

25 In a further embodiment, the invention provides a method for producing eukaryotic cells having multiple copies of the product gene comprising transfecting eukaryotic cells with the DNA construct described above (where the selectable gene is an amplifiable gene), growing the cells in a selective medium comprising an amplifying agent for a sufficient time for amplification to occur, and selecting cells having multiple copies of the product gene. Preferably transfection of the cells is achieved using electroporation.

35 After transfection of the host cells, most of the transfectants fail to exhibit the selectable phenotype characteristic of the protein encoded by the selectable gene, but surprisingly a small proportion of the transfectants do exhibit the selectable phenotype, and among those transfectants, the majority are found to express high levels of the desired product encoded by the product gene. Thus, the invention provides an improved method for the selection of recombinant host cells expressing high levels of a desired product, which method is useful with a wide variety of eukaryotic host cells and avoids the problems inherent in existing cell selection technology.

BRIEF DESCRIPTION OF THE DRAWINGS

Figures 1A-1D illustrate schematically various DNA constructs encompassed by the instant invention. The large arrows represent the selectable gene and the product gene, the V formed by the dashed lines shows the region of the precursor RNA internal to the 5' splice donor site (SD) and 3' splice acceptor site (SA) that is excised from vectors that contain a functional SD. The transcriptional regulatory region, selectable gene, product gene and transcriptional termination site are depicted in Figure 1A. Figure 1B depicts the DNA constructs of Example 1. The various splice donor sequences are depicted, i.e., wild type ras splice donor sequence (WT ras), mutant ras splice donor sequence (MUTANT ras) and non-functional splice donor sequence (Δ GT). The probes used for Northern blot analysis in Example 1 are shown in Figure 1B. Figure 1C depicts the DNA constructs of Example 2 and Figure 1D depicts the DNA construct of Example 3 used for expression of anti-IgE V_H.

Figure 2 depicts schematically the control DNA construct used in Example 1.

Figures 3A-Q depict the nucleotide sequence (SEQ ID NO: 1) of the DHFR/intron-(WT ras SD)-tPA expression vector of Example 1.

Figure 4 is a bar graph which shows the number of colonies that form in selective medium after electroporation of linearized duplicate miniprep DNA's prepared in parallel from the three vectors shown in Figure 1B (i.e. with wild type ras splice donor sequence [WT ras], mutant ras splice donor sequence [MUTANT ras] and non-functional splice donor sequence [Δ GT]) and from the control vector that has DHFR under control of SV40 promoter and tPA under control of CMV promoter (see Figure 2). Cells were selected in nucleoside free medium and counted with an automated colony counter.

Figures 5A-C are bar graphs depicting expression of tPA from stable pools and clones generated from the vectors shown in Figure 1B. In Figure 5A greater than 100 clones from each vector transfection were mixed, plated in 24 well plates, and assayed by tPA ELISA at "saturation". In Figure 5B, twenty clones chosen at random derived from each of the vectors were assayed by tPA ELISA at "saturation". In Figure 5C, the pools mentioned in Figure 5A (except the Δ GT pool) were exposed to 200nM Mtx to select for DHFR amplification and then pooled and assayed for tPA expression.

Figures 6A-P depict the nucleotide sequence (SEQ ID NO: 2) of the DHFR/intron-(WT ras SD)-TNFr-IgG expression vector of Example 2.

Figures 7A-B are bar graphs depicting expression of TNFr-IgG using dicistronic or control vectors (see Example 2). Vectors containing TNFr-IgG (but otherwise identical to those described for tPA expression in Example 1) were constructed (see Figure 1C), introduced into dp12.CHO cells by electroporation, pooled, and assayed for product expression before (Figure 7A) and after (Figure 7B) being subjected to amplification in 200nM Mtx.

Figure 8 depicts schematically the DNA construct used for expression of the V_L of anti-IgE in Example 3.

Figures 9A-O depict the nucleotide sequence (SEQ ID NO: 3) of the anti-IgE V_H expression vector of Example 3.

Figures 10A-Q depict the nucleotide sequence (SEQ ID NO: 4) of the anti-IgE V_L expression vector of Example 3.

5 Figure 11 is a bar graph depicting anti-IgE expression in Example 3. Heavy (V_H) and light (V_L) chain expression vectors were constructed, co-electroporated into CHO cells, clones were selected and assayed for antibody expression. Additionally, pools were established and assessed with regard to expression before and after Mtx selection at 200nM and 1μM.

10 DESCRIPTION OF THE PREFERRED EMBODIMENTS

Definitions:

The "DNA construct" disclosed herein comprises a non-naturally occurring DNA molecule which can either be provided as an isolate or integrated in another DNA molecule e.g. in an expression vector or the
15 chromosome of an eukaryotic host cell.

The term "selectable gene" as used herein refers to a DNA that encodes a selectable marker necessary for the growth or survival of a host cell under the particular cell culture conditions chosen. Accordingly, a host cell that is transformed with a selectable gene will be capable of
20 growth or survival under certain cell culture conditions wherein a non-transfected host cell is not capable of growth or survival. Typically, a selectable gene will confer resistance to a drug or compensate for a metabolic or catabolic defect in the host cell. Examples of selectable genes are provided in the following table. See also Kaufman, Methods in
25 Enzymology, 185: 537-566 (1990), for a review of these.

TABLE 1
Selectable Genes and their Selection Agents

Selection Agent	Selectable Gene
Methotrexate	Dihydrofolate reductase
Cadmium	Metallothionein
PALA	CAD
Xyl-A-or adenosine and 2'- deoxycoformycin	Adenosine deaminase
Adenine, azaserine, and coformycin	Adenylate deaminase
6-Azaauridine, pyrazofuran	UMP Synthetase
Mycophenolic acid	IMP 5'-dehydrogenase

	Mycophenolic acid with limiting xanthine	Xanthine-guanine phosphoribosyltransferase
	Hypoxanthine, aminopterin, and thymidine (HAT)	Mutant HGPRTase or mutant thymidine kinase
5	5-Fluorodeoxyuridine	Thymidylate synthetase
	Multiple drugs e.g. adriamycin, vincristine or colchicine	P-glycoprotein 170
	Aphidicolin	Ribonucleotide reductase
10	Methionine sulfoximine	Glutamine synthetase
	β -Aspartyl hydroxamate or Albizziin	Asparagine synthetase
	Canavanine	Arginosuccinate synthetase
	α -Difluoromethylornithine	Ornithine decarboxylase
15	Compactin	HMG-CoA reductase
	Tunicamycin	N-Acetylglucosaminyl transferase
	Borrelidin	Threonyl-tRNA synthetase
	Ouabain	Na ⁺ K ⁺ -ATPase

The preferred selectable gene is an amplifiable gene. As used herein, the term "amplifiable gene" refers to a gene which is amplified (i.e. additional copies of the gene are generated which survive in intrachromosomal or extrachromosomal form) under certain conditions. The amplifiable gene usually encodes an enzyme (i.e. an amplifiable marker) which is required for growth of eukaryotic cells under those conditions. For example, the gene may encode DHFR which is amplified when a host cell transformed therewith is grown in Mtx. According to Kaufman, the selectable genes in Table 1 above can also be considered amplifiable genes. An example of a selectable gene which is generally not considered to be an amplifiable gene is the neomycin resistance gene (Cepko et al., *supra*).

As used herein, "selective medium" refers to nutrient solution used for growing eukaryotic cells which have the selectable gene and therefore includes a "selection agent". Commercially available media such as Ham's F10 (Sigma), Minimal Essential Medium ([MEM], Sigma), RPMI-1640 (Sigma), and Dulbecco's Modified Eagle's Medium ([DMEM], Sigma) are exemplary nutrient solutions. In addition, any of the media described in Ham and Wallace, *Meth. Enz.*, 58:44 (1979), Barnes and Sato, *Anal. Biochem.*, 102:255

(1980), U.S. Patent Nos. 4,767,704; 4,657,866; 4,927,762; or 4,560,655; WO 90/03430; WO 87/00195; U.S. Patent Re. 30,985; or U.S. Patent No. 5,122,469, the disclosures of all of which are incorporated herein by reference, may be used as culture media. Any of these media may be
5 supplemented as necessary with hormones and/or other growth factors (such as insulin, transferrin, or epidermal growth factor), salts (such as sodium chloride, calcium, magnesium, and phosphate), buffers (such as HEPES), nucleosides (such as adenosine and thymidine), antibiotics (such as Gentamycin™ drug), trace elements (defined as inorganic compounds usually
10 present at final concentrations in the micromolar range), and glucose or an equivalent energy source. Any other necessary supplements may also be included at appropriate concentrations that would be known to those skilled in the art. The preferred nutrient solution comprises fetal bovine serum.

The term "selection agent" refers to a substance that interferes with
15 the growth or survival of a host cell that is deficient in a particular selectable gene. Examples of selection agents are presented in Table 1 above. The selection agent preferably comprises an "amplifying agent" which is defined for purposes herein as an agent for amplifying copies of the amplifiable gene, such as Mtx if the amplifiable gene is DHFR. See Table
20 1 for examples of amplifying agents.

As used herein, the term "transcriptional initiation site" refers to the nucleic acid in the DNA construct corresponding to the first nucleic acid incorporated into the primary transcript, i.e., the mRNA precursor, which site is generally provided at, or adjacent to, the 5' end of the DNA
25 construct.

The term "transcriptional termination site" refers to a sequence of DNA, normally represented at the 3' end of the DNA construct, that causes RNA polymerase to terminate transcription.

As used herein, "transcriptional regulatory region" refers to a
30 region of the DNA construct that regulates transcription of the selectable gene and the product gene. The transcriptional regulatory region normally refers to a promoter sequence (i.e. a region of DNA involved in binding of RNA polymerase to initiate transcription) which can be constitutive or inducible and, optionally, an enhancer (i.e. a *cis*-acting DNA element,
35 usually from about 10-300 bp, that acts on a promoter to increase its transcription).

As used herein, "product gene" refers to DNA that encodes a desired protein or polypeptide product. Any product gene that is capable of expression in a host cell may be used, although the methods of the
40 invention are particularly suited for obtaining high-level expression of a product gene that is not also a selectable or amplifiable gene. Accordingly, the protein or polypeptide encoded by a product gene typically will be one that is not necessary for the growth or survival of a host cell under the particular cell culture conditions chosen. For example, product
45 genes suitably encode a peptide, or may encode a polypeptide sequence of

amino acids for which the chain length is sufficient to produce higher levels of tertiary and/or quaternary structure.

Examples of bacterial polypeptides or proteins include, e.g., alkaline phosphatase and β -lactamase. Examples of mammalian polypeptides or proteins include molecules such as renin; a growth hormone, including human growth hormone, and bovine growth hormone; growth hormone releasing factor; parathyroid hormone; thyroid stimulating hormone; lipoproteins; alpha-1-antitrypsin; insulin A-chain; insulin B-chain; proinsulin; follicle stimulating hormone; calcitonin; luteinizing hormone; glucagon; clotting factors such as factor VIIIC, factor IX, tissue factor, and von Willebrands factor; anti-clotting factors such as Protein C; atrial natriuretic factor; lung surfactant; a plasminogen activator, such as urokinase or human urine or tissue-type plasminogen activator (t-PA); bombesin; thrombin; hemopoietic growth factor; tumor necrosis factor-alpha and -beta; enkephalinase; RANTES (regulated on activation normally T-cell expressed and secreted); human macrophage inflammatory protein (MIP-1-alpha); a serum albumin such as human serum albumin; mullerian-inhibiting substance; relaxin A-chain; relaxin B-chain; prorelaxin; mouse gonadotropin-associated peptide; a microbial protein, such as beta-lactamase; DNase; inhibin; activin; vascular endothelial growth factor (VEGF); receptors for hormones or growth factors; integrin; protein A or D; rheumatoid factors; a neurotrophic factor such as bone-derived neurotrophic factor (BDNF), neurotrophin-3, -4, -5, or -6 (NT-3, NT-4, NT-5, or NT-6), or a nerve growth factor such as NGF- β ; platelet-derived growth factor (PDGF); fibroblast growth factor such as aFGF and bFGF; epidermal growth factor (EGF); transforming growth factor (TGF) such as TGF-alpha and TGF-beta, including TGF- β 1, TGF- β 2, TGF- β 3, TGF- β 4, or TGF- β 5; insulin-like growth factor-I and -II (IGF-I and IGF-II); des(1-3)-IGF-I (brain IGF-I), insulin-like growth factor binding proteins; CD proteins such as CD-3, CD-4, CD-8, and CD-19; erythropoietin; osteoinductive factors; immunotoxins; a bone morphogenetic protein (BMP); an interferon such as interferon-alpha, -beta, and -gamma; colony stimulating factors (CSFs), e.g., M-CSF, GM-CSF, and G-CSF; interleukins (ILs), e.g., IL-1 to IL-10; superoxide dismutase; T-cell receptors; surface membrane proteins; decay accelerating factor; viral antigen such as, for example, a portion of the AIDS envelope; transport proteins; homing receptors; addressins; regulatory proteins; antibodies; chimeric proteins such as immunoadhesins and fragments of any of the above-listed polypeptides.

The product gene preferably does not consist of an anti-sense sequence for inhibiting the expression of a gene present in the host. Preferred proteins herein are therapeutic proteins such as TGF- β , TGF- α , PDGF, EGF, FGF, IGF-I, DNase, plasminogen activators such as t-PA, clotting factors such as tissue factor and factor VIII, hormones such as relaxin and insulin, cytokines such as IFN- γ , chimeric proteins such as TNF receptor IgG immunoadhesin (TNFr-IgG) or antibodies such as anti-IgE.

The term "intron" as used herein refers to a nucleotide sequence present within the transcribed region of a gene or within a messenger RNA precursor, which nucleotide sequence is capable of being excised, or spliced, from the messenger RNA precursor by a host cell prior to translation. Introns suitable for use in the present invention are suitably prepared by any of several methods that are well known in the art, such as purification from a naturally occurring nucleic acid or de novo synthesis. The introns present in many naturally occurring eukaryotic genes have been identified and characterized. Mount, Nuc. Acids Res., 10:459 (1982). Artificial introns comprising functional splice sites also have been described. Winey et al., Mol. Cell Biol., 9:329 (1989); Gattermann et al., Mol. Cell Biol., 9:1526 (1989). Introns may be obtained from naturally occurring nucleic acids, for example, by digestion of a naturally occurring nucleic acid with a suitable restriction endonuclease, or by PCR cloning using primers complementary to sequences at the 5' and 3' ends of the intron. Alternatively, introns of defined sequence and length may be prepared synthetically using various methods in organic chemistry. Narang et al., Meth. Enzymol., 68:90 (1979); Caruthers et al., Meth. Enzymol., 154:287 (1985); Froehler et al., Nuc. Acids Res., 14:5399 (1986).

As used herein "splice donor site" or "SD" refers to the DNA sequence immediately surrounding the exon-intron boundary at the 5' end of the intron, where the "exon" comprises the nucleic acid 5' to the intron. Many splice donor sites have been characterized and Ohshima et al., J. Mol. Biol., 195:247-259 (1987) provides a review of these. An "efficient splice donor sequence" refers to a nucleic acid sequence encoding a splice donor site wherein the efficiency of splicing of messenger RNA precursors having the splice donor sequence is between about 80 to 99% and preferably 90 to 95% as determined by quantitative PCR. Examples of efficient splice donor sequences include the wild type (WT) ras splice donor sequence and the GAC:GTAAGT sequence of Example 3. Other efficient splice donor sequences can be readily selected using the techniques for measuring the efficiency of splicing disclosed herein.

The terms "PCR" and "polymerase chain reaction" as used herein refer to the in vitro amplification method described in US Patent No. 4,683,195 (issued July 28, 1987). In general, the PCR method involves repeated cycles of primer extension synthesis, using two DNA primers capable of hybridizing preferentially to a template nucleic acid comprising the nucleotide sequence to be amplified. The PCR method can be used to clone specific DNA sequences from total genomic DNA, cDNA transcribed from cellular RNA, viral or plasmid DNAs. Wang & Mark, in PCR Protocols, pp. 70-75 (Academic Press, 1990); Scharf, in PCR Protocols, pp. 84-98; Kawasaki & Wang, in PCR Technology, pp. 89-97 (Stockton Press, 1989). Reverse transcription-polymerase chain reaction (RT-PCR) can be used to analyze RNA samples containing mixtures of spliced and unspliced mRNA transcripts. Fluorescently tagged primers designed to span the intron are used to

amplify both spliced and unspliced targets. The resultant amplification products are then separated by gel electrophoresis and quantitated by measuring the fluorescent emission of the appropriate band(s). A comparison is made to determine the amount of spliced and unspliced transcripts present in the RNA sample.

One preferred splice donor sequence is a "consensus splice donor sequence". The nucleotide sequences surrounding intron splice sites, which sequences are evolutionarily highly conserved, are referred to as "consensus splice donor sequences". In the mRNAs of higher eukaryotes, the 5' splice site occurs within the consensus sequence AG:GUAAGU (wherein the colon denotes the site of cleavage and ligation). In the mRNAs of yeast, the 5' splice site is bounded by the consensus sequence :GUAUGU. Padgett, et al., Ann. Rev. Biochem., 55:1119 (1986).

The expression "splice acceptor site" or "SA" refers to the sequence immediately surrounding the intron-exon boundary at the 3' end of the intron, where the "exon" comprises the nucleic acid 3' to the intron. Many splice acceptor sites have been characterized and Ohshima et al., J. Mol. Biol., 195:247-259 (1987) provides a review of these. The preferred splice acceptor site is an efficient splice acceptor site which refers to a nucleic acid sequence encoding a splice acceptor site wherein the efficiency of splicing of messenger RNA precursors having the splice acceptor site is between about 80 to 99% and preferably 90 to 95% as determined by quantitative PCR. The splice acceptor site may comprise a consensus sequence. In the mRNAs of higher eukaryotes, the 3' splice acceptor site occurs within the consensus sequence (U/C)₁₁NCAG:G. In the mRNAs of yeast, the 3' acceptor splice site is bounded by the consensus sequence (C/U)AG:. Padgett, et al., *supra*.

As used herein "culturing for sufficient time to allow amplification to occur" refers to the act of physically culturing the eukaryotic host cells which have been transformed with the DNA construct in cell culture media containing the amplifying agent, until the copy number of the amplifiable gene (and preferably also the copy number of the product gene) in the host cells has increased relative to the transformed cells prior to this culturing.

The term "expression" as used herein refers to transcription or translation occurring within a host cell. The level of expression of a product gene in a host cell may be determined on the basis of either the amount of corresponding mRNA that is present in the cell or the amount of the protein encoded by the product gene that is produced by the cell. For example, mRNA transcribed from a product gene is desirably quantitated by northern hybridization. Sambrook, et al., Molecular Cloning: A Laboratory Manual, pp. 7.3-7.57 (Cold Spring Harbor Laboratory Press, 1989). Protein encoded by a product gene can be quantitated either by assaying for the biological activity of the protein or by employing assays that are independent of such activity, such as western blotting or radioimmunoassay using antibodies that are capable of reacting with the protein. Sambrook,

et al., Molecular Cloning: A Laboratory Manual, pp. 18.1-18.88 (Cold Spring Harbor Laboratory Press, 1989).

Modes for Carrying Out the Invention

Methods and compositions are provided for enhancing the stability and/or copy number of a transcribed sequence in order to allow for elevated levels of a RNA sequence of interest. In general, the methods of the present invention involve transfecting a eukaryotic host cell with an expression vector comprising both a product gene encoding a desired polypeptide and a selectable gene (preferably an amplifiable gene).

Selectable genes and product genes may be obtained from genomic DNA, cDNA transcribed from cellular RNA, or by in vitro synthesis. For example, libraries are screened with probes (such as antibodies or oligonucleotides of about 20-80 bases) designed to identify the selectable gene or the product gene (or the protein(s) encoded thereby). Screening the cDNA or genomic library with the selected probe may be conducted using standard procedures as described in chapters 10-12 of Sambrook et al., Molecular Cloning: A Laboratory Manual (New York: Cold Spring Harbor Laboratory Press, 1989). An alternative means to isolate the selectable gene or product gene is to use PCR methodology as described in section 14 of Sambrook et al., *supra*.

A preferred method of practicing this invention is to use carefully selected oligonucleotide sequences to screen cDNA libraries from various tissues known to contain the selectable gene or product gene. The oligonucleotide sequences selected as probes should be of sufficient length and sufficiently unambiguous that false positives are minimized.

The oligonucleotide generally is labeled such that it can be detected upon hybridization to DNA in the library being screened. The preferred method of labeling is to use ³²P- labeled ATP with polynucleotide kinase, as is well known in the art, to radiolabel the oligonucleotide. However, other methods may be used to label the oligonucleotide, including, but not limited to, biotinylation or enzyme labeling.

Sometimes, the DNA encoding the selectable gene and product gene is preceded by DNA encoding a signal sequence having a specific cleavage site at the N-terminus of the mature protein or polypeptide. In general, the signal sequence may be a component of the expression vector, or it may be a part of the selectable gene or product gene that is inserted into the expression vector. If a heterologous signal sequence is used, it preferably is one that is recognized and processed (i.e., cleaved by a signal peptidase) by the host cell. For yeast secretion the native signal sequence may be substituted by, e.g., the yeast invertase leader, alpha factor leader (including *Saccharomyces* and *Kluyveromyces* α -factor leaders, the latter described in U.S. Pat. No. 5,010,182 issued 23 April 1991), or acid phosphatase leader, the *C. albicans* glucoamylase leader (EP 362,179 published 4 April 1990), or the signal described in WO 90/13646 published 15 November 1990. In mammalian cell expression the native signal sequence

of the protein of interest is satisfactory, although other mammalian signal sequences may be suitable, such as signal sequences from secreted polypeptides of the same or related species, as well as viral secretory leaders, for example, the herpes simplex gD signal. The DNA for such precursor region is ligated in reading frame to the selectable gene or product gene.

As shown in Figure 1A, the selectable gene is generally provided at the 5' end of the DNA construct and this selectable gene is followed by the product gene. Therefore, the full length (non-spliced) message will contain DHFR as the first open reading frame and will therefore generate DHFR protein to allow selection of stable transfectants. The full length message is not expected to generate appreciable amounts of the protein of interest as the second AUG in a dicistronic message is an inefficient initiator of translation in mammalian cells (Kozak, J. Cell Biol., 115: 887-903 [1991]).

The selectable gene is positioned within an intron. Introns are noncoding nucleotide sequences, normally present within many eukaryotic genes, which are removed from newly transcribed mRNA precursors in a multiple-step process collectively referred to as splicing.

A single mechanism is thought to be responsible for the splicing of mRNA precursors in mammalian, plant, and yeast cells. In general, the process of splicing requires that the 5' and 3' ends of the intron be correctly cleaved and the resulting ends of the mRNA be accurately joined, such that a mature mRNA having the proper reading frame for protein synthesis is produced. Analysis of a variety of naturally occurring and synthetically constructed mutant genes has shown that nucleotide changes at many of the positions within the consensus sequences at the 5' and 3' splice sites have the effect of reducing or abolishing the synthesis of mature mRNA. Sharp, Science, 235:766 (1987); Padgett, et al., Ann. Rev. Biochem., 55:1119 (1986); Green, Ann. Rev. Genet., 20:671 (1986). Mutational studies also have shown that RNA secondary structures involving splicing sites can affect the efficiency of splicing. Solnick, Cell, 43:667 (1985); Konarska, et al., Cell, 42:165 (1985).

The length of the intron may also affect the efficiency of splicing. By making deletion mutations of different sizes within the large intron of the rabbit beta-globin gene, Wieringa, et al. determined that the minimum intron length necessary for correct splicing is about 69 nucleotides. Cell, 37:915 (1984). Similar studies of the intron of the adenovirus E1A region have shown that an intron length of about 78 nucleotides allows correct splicing to occur, but at reduced efficiency. Increasing the length of the intron to 91 nucleotides restores normal splicing efficiency, whereas truncating the intron to 63 nucleotides abolishes correct splicing. Ulfendahl, et al., Nuc. Acids Res., 13:6299 (1985).

To be useful in the invention, the intron must have a length such that splicing of the intron from the mRNA is efficient. The preparation of introns of differing lengths is a routine matter, involving methods well known in the art, such as de novo synthesis or in vitro deletion

mutagenesis of an existing intron. Typically, the intron will have a length of at least about 150 nucleotides, since introns which are shorter than this tend to be spliced less efficiently. The upper limit for the length of the intron can be up to 30 kB or more. However, as a general proposition, the intron is generally less than about 10 kB in length.

The intron is modified to contain the selectable gene not normally present within the intron using any of the various known methods for modifying a nucleic acid *in vitro*. Typically, a selectable gene will be introduced into an intron by first cleaving the intron with a restriction endonuclease, and then covalently joining the resulting restriction fragments to the selectable gene in the correct orientation for host cell expression, for example by ligation with a DNA ligase enzyme.

The DNA construct is dicistronic, i.e. the selectable gene and product gene are both under the transcriptional control of a single transcriptional regulatory region. As mentioned above, the transcriptional regulatory region comprises a promoter. Suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase (Hitzeman et al., J. Biol. Chem., 255:2073 [1980]) or other glycolytic enzymes (Hess et al., J. Adv. Enzyme Reg., 7:149 [1968]; and Holland, Biochemistry, 17:4900 [1978]), such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase.

Other yeast promoters, which are inducible promoters having the additional advantage of transcription controlled by growth conditions, are the promoter regions for alcohol dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Suitable vectors and promoters for use in yeast expression are further described in Hitzeman et al., EP 73,657A. Yeast enhancers also are advantageously used with yeast promoters.

Expression control sequences are known for eukaryotes. Virtually all eukaryotic genes have an AT-rich region located approximately 25 to 30 bases upstream from the site where transcription is initiated. Another sequence found 70 to 80 bases upstream from the start of transcription of many genes is a CXCAAT region where X may be any nucleotide.

Product gene transcription from vectors in mammalian host cells is controlled by promoters obtained from the genomes of viruses such as polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and most preferably Simian Virus 40 (SV40), from heterologous mammalian promoters, e.g. the actin promoter or an immunoglobulin promoter, from heat-shock promoters, and from the promoter normally associated with the product gene, provided such promoters are compatible with the host cell systems.

The early and late promoters of the SV40 virus are conveniently obtained as an SV40 restriction fragment that also contains the SV40 viral origin of replication. Fiers et al., Nature, 273:113 (1978); Mulligan and Berg, Science, 209:1422-1427 (1980); Pavlakis et al., Proc. Natl. Acad. Sci. USA, 78:7398-7402 (1981). The immediate early promoter of the human cytomegalovirus (CMV) is conveniently obtained as a HindIII E restriction fragment. Greenaway et al., Gene, 18:355-360 (1982). A system for expressing DNA in mammalian hosts using the bovine papilloma virus as a vector is disclosed in U.S. 4,419,446. A modification of this system is described in U.S. 4,601,978. See also Gray et al., Nature, 295:503-508 (1982) on expressing cDNA encoding immune interferon in monkey cells; , Reyes et al., Nature, 297:598-601 (1982) on expression of human β -interferon cDNA in mouse cells under the control of a thymidine kinase promoter from herpes simplex virus, Canaani and Berg, Proc. Natl. Acad. Sci. USA, 79:5166-5170 (1982) on expression of the human interferon β 1 gene in cultured mouse and rabbit cells, and Gorman et al., Proc. Natl. Acad. Sci. USA, 79:6777-6781 (1982) on expression of bacterial CAT sequences in CV-1 monkey kidney cells, chicken embryo fibroblasts, Chinese hamster ovary cells, HeLa cells, and mouse NIH-3T3 cells using the Rous sarcoma virus long terminal repeat as a promoter.

Preferably the transcriptional regulatory region in higher eukaryotes comprises an enhancer sequence. Enhancers are relatively orientation and position independent having been found 5' (Lainins et al., Proc. Natl. Acad. Sci. USA, 78:993 [1981]) and 3' (Lusky et al., Mol. Cell Bio., 3:1108 [1983]) to the transcription unit, within an intron (Banerji et al., Cell, 33:729 [1983]) as well as within the coding sequence itself (Osborne et al., Mol. Cell Bio., 4:1293 [1984]). Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, α -fetoprotein and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer (CMV), the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. See also Yaniv, Nature, 297:17-18 (1982) on enhancing elements for activation of eukaryotic promoters. The enhancer may be spliced into the vector at a position 5' or 3' to the product gene, but is preferably located at a site 5' from the promoter.

The DNA construct has a transcriptional initiation site following the transcriptional regulatory region and a transcriptional termination region following the product gene (see Figure 1A). These sequences are provided in the DNA construct using techniques which are well known in the art.

The DNA construct normally forms part of an expression vector which may have other components such as an origin of replication (i.e., a nucleic acid sequence that enables the vector to replicate in one or more selected host cells) and, if desired, one or more additional selectable gene(s). Construction of suitable vectors containing the desired coding and control sequences employs standard ligation techniques. Isolated plasmids or DNA

fragments are cleaved, tailored, and religated in the form desired to generate the plasmids required.

Generally, in cloning vectors the origin of replication is one that enables the vector to replicate independently of the host chromosomal DNA, and includes origins of replication or autonomously replicating sequences. Such sequences are well known. The 2 μ plasmid origin of replication is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells. Generally, the origin of replication component is not needed for mammalian expression vectors (the SV40 origin may typically be used only because it contains the early promoter).

Most expression vectors are "shuttle" vectors, i.e., they are capable of replication in at least one class of organisms but can be transfected into another organism for expression. For example, a vector is cloned in *E. coli* and then the same vector is transfected into yeast or mammalian cells for expression even though it is not capable of replicating independently of the host cell chromosome.

For analysis to confirm correct sequences in plasmids constructed, plasmids from the transformants are prepared, analyzed by restriction, and/or sequenced by the method of Messing et al., Nucleic Acids Res., 9:309 (1981) or by the method of Maxam et al., Methods in Enzymology, 65:499 (1980).

The expression vector having the DNA construct prepared as discussed above is transformed into a eukaryotic host cell. Suitable host cells for cloning or expressing the vectors herein are yeast or higher eukaryote cells.

Eukaryotic microbes such as filamentous fungi or yeast are suitable hosts for vectors containing the product gene. *Saccharomyces cerevisiae*, or common baker's yeast, is the most commonly used among lower eukaryotic host microorganisms. However, a number of other genera, species, and strains are commonly available and useful herein, such as *S. pombe* [Beach and Nurse, Nature, 290:140 (1981)], *Kluyveromyces lactis* [Louvencourt et al., J. Bacteriol., 737 (1983)], *Yarrowia* [EP 402,226], *Pichia pastoris* [EP 183,070], *Trichoderma reesia* [EP 244,234], *Neurospora crassa* [Case et al., Proc. Natl. Acad. Sci. USA, 76:5259-5263 (1979)], and *Aspergillus* hosts such as *A. nidulans* [Ballance et al., Biochem. Biophys. Res. Commun., 112:284-289 (1983); Tilburn et al., Gene, 26:205-221 (1983); Yelton et al., Proc. Natl. Acad. Sci. USA, 81:1470-1474 (1984)] and *A. niger* [Kelly and Hynes, EMBO J., 4:475-479 (1985)].

Suitable host cells for the expression of the product gene are derived from multicellular organisms. Such host cells are capable of complex processing and glycosylation activities. In principle, any higher eukaryotic cell culture is workable, whether from vertebrate or invertebrate culture. Examples of invertebrate cells include plant and insect cells. Numerous baculoviral strains and variants and corresponding permissive insect host cells from hosts such as *Spodoptera frugiperda*

(caterpillar), *Aedes aegypti* (mosquito), *Aedes albopictus* (mosquito), *Drosophila melanogaster* (fruitfly), and *Bombyx mori* host cells have been identified. See, e.g., Luckow et al., Bio/Technology, 6:47-55 (1988); Miller et al., in Genetic Engineering, Setlow, J.K. et al., eds., Vol. 8
5 (Plenum Publishing, 1986), pp. 277-279; and Maeda et al., Nature, 315:592-594 (1985). A variety of such viral strains are publicly available, e.g., the L-1 variant of *Autographa californica* NPV and the Bm-5 strain of *Bombyx mori* NPV, and such viruses may be used as the virus herein according to the present invention, particularly for transfection of *Spodoptera frugiperda*
10 cells.

Plant cell cultures of cotton, corn, potato, soybean, petunia, tomato, and tobacco can be utilized as hosts. Typically, plant cells are transfected by incubation with certain strains of the bacterium *Agrobacterium tumefaciens*, which has been previously manipulated to contain
15 the product gene. During incubation of the plant cell culture with *A. tumefaciens*, the product gene is transferred to the plant cell host such that it is transfected, and will, under appropriate conditions, express the product gene. In addition, regulatory and signal sequences compatible with plant cells are available, such as the nopaline synthase promoter and
20 polyadenylation signal sequences. Depicker et al., J. Mol. Appl. Gen., 1:561 (1982). In addition, DNA segments isolated from the upstream region of the T-DNA 780 gene are capable of activating or increasing transcription levels of plant-expressible genes in recombinant DNA-containing plant tissue. EP 321,196 published 21 June 1989.

25 However, interest has been greatest in vertebrate cells, and propagation of vertebrate cells in culture (tissue culture) has become a routine procedure in recent years [Tissue Culture, Academic Press, Kruse and Patterson, editors (1973)]. Examples of useful mammalian host cell lines are monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL
30 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham et al., J. Gen Virol., 36:59 [1977]); baby hamster kidney cells (BHK, ATCC CCL 10); Chinese hamster ovary cells/-DHFR (CHO, Urlaub and Chasin, Proc. Natl. Acad. Sci. USA, 77:4216 [1980]); dp12.CHO cells (EP 307,247 published 15 March 1989); mouse sertoli cells
35 (TM4, Mather, Biol. Reprod., 23:243-251 [1980]); monkey kidney cells (CV1 ATCC CCL 70); African green monkey kidney cells (VERO-76, ATCC CRL-1587); human cervical carcinoma cells (HELA, ATCC CCL 2); canine kidney cells (MDCK, ATCC CCL 34); buffalo rat liver cells (BRL 3A, ATCC CRL 1442); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); mouse
40 mammary tumor (MMT 060562, ATCC CCL51); TRI cells (Mather et al., Annals N.Y. Acad. Sci., 383:44-68 [1982]); MRC 5 cells; FS4 cells; and a human hepatoma line (Hep G2).

Host cells are transformed with the above-described expression or cloning vectors of this invention and cultured in conventional nutrient
45 media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences.

Infection with *Agrobacterium tumefaciens* is used for transformation of certain plant cells, as described by Shaw et al., Gene, 23:315 (1983) and WO 89/05859 published 29 June 1989. For mammalian cells without such cell walls, the calcium phosphate precipitation method of Graham and van der Eb, Virology, 52:456-457 (1978) may be used. General aspects of mammalian cell host system transformations have been described by Axel in U.S. 4,399,216 issued 16 August 1983. Transformations into yeast are typically carried out according to the method of Van Solingen et al., J. Bact., 130:946 (1977) and Hsiao et al., Proc. Natl. Acad. Sci. (USA), 76:3829 (1979). However, other methods for introducing DNA into cells such as by nuclear injection or by protoplast fusion may also be used.

In the preferred embodiment the DNA is introduced into the host cells using electroporation. See Andreason, J. Tiss. Cult. Meth., 15:56-62 (1993), for a review of electroporation techniques useful for practicing the instantly claimed invention. It was discovered that electroporation techniques for introducing the DNA construct into the host cells were preferable over calcium phosphate precipitation techniques insofar as the latter could cause the DNA to break up and forming concatemers.

The mammalian host cells used to express the product gene herein may be cultured in a variety of media as discussed in the definitions section above. The media contains the selection agent used for selecting transformed host cells which have taken up the DNA construct (either as an intra- or extra-chromosomal element). To achieve selection of the transformed eukaryotic cells, the host cells may be grown in cell culture plates and individual colonies expressing the selectable gene (and thus the product gene) can be isolated and grown in growth medium until the nutrients are depleted. The host cells are then analyzed for transcription and/or transformation as discussed below. The culture conditions, such as temperature, pH, and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

Gene amplification and/or expression may be measured in a sample directly, for example, by conventional Southern blotting, Northern blotting to quantitate the transcription of mRNA (Thomas, Proc. Natl. Acad. Sci. USA, 77:5201-5205 [1980]), dot blotting (DNA analysis), or in situ hybridization, using an appropriately labeled probe, based on the sequences provided herein. Various labels may be employed, most commonly radioisotopes, particularly ³²P. However, other techniques may also be employed, such as using biotin-modified nucleotides for introduction into a polynucleotide. The biotin then serves as the site for binding to avidin or antibodies, which may be labeled with a wide variety of labels, such as radionuclides, fluorescens, enzymes, or the like. Alternatively, antibodies may be employed that can recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in turn may be labeled and the assay may be carried out where the duplex is bound to a surface, so that upon the

formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Gene expression, alternatively, may be measured by immunological methods, such as immunohistochemical staining of tissue sections and assay
5 of cell culture or body fluids, to quantitate directly the expression of gene product. With immunohistochemical staining techniques, a cell sample is prepared, typically by dehydration and fixation, followed by reaction with labeled antibodies specific for the gene product coupled, where the labels are usually visually detectable, such as enzymatic labels,
10 fluorescent labels, luminescent labels, and the like. A particularly sensitive staining technique suitable for use in the present invention is described by Hsu et al., Am. J. Clin. Path., 75:734-738 (1980).

In the preferred embodiment, the mRNA is analyzed by quantitative PCR (to determine the efficiency of splicing) and protein expression is
15 measured using ELISA as described in Example 1 herein.

The product of interest preferably is recovered from the culture medium as a secreted polypeptide, although it also may be recovered from host cell lysates when directly expressed without a secretory signal. When the product gene is expressed in a recombinant cell other than one of human
20 origin, the product of interest is completely free of proteins or polypeptides of human origin. However, it is necessary to purify the product of interest from recombinant cell proteins or polypeptides to obtain preparations that are substantially homogeneous as to the product of interest. As a first step, the culture medium or lysate is centrifuged
25 to remove particulate cell debris. The product of interest thereafter is purified from contaminant soluble proteins and polypeptides, for example, by fractionation on immunoaffinity or ion-exchange columns; ethanol precipitation; reverse phase HPLC; chromatography on silica or on a cation exchange resin such as DEAE; chromatofocusing; SDS-PAGE; ammonium sulfate
30 precipitation; gel electrophoresis using, for example, Sephadex G-75; chromatography on plasminogen columns to bind the product of interest and protein A Sepharose columns to remove contaminants such as IgG.

The following examples are offered by way of illustration only and are not intended to limit the invention in any manner. All patent and
35 literature references cited herein are expressly incorporated by reference.

EXAMPLE 1

tPA production using the dicistronic expression vectors

It was sought to increase the level of homogeneity with regard to expression levels of stable clones by expressing a selectable marker (such
40 as DHFR) and the protein of interest from a single promoter. These vectors divert most of the transcript to product expression while linking it at a fixed ratio to DHFR expression via differential splicing.

Vectors were constructed which were derived from the vector pRK (Suva et al., Science, 237:893-896 [1987]) which contains an intron between the
45 cytomegalovirus immediate early promoter (CMV) and the cDNA that encodes

the polypeptide of interest. The intron of pRK is 139 nucleotides in length, has a splice donor site derived from cytomegalovirus immediate early gene (CMVIE), and a splice acceptor site from an IgG heavy chain variable region (V_H) gene (Eaton et al., Biochem., 25:8343 [1986]).

5 DHFR/intron vectors were constructed by inserting an EcoRV linker into the BSTX1 site present in the intron of pRK7. An 830 base-pair fragment containing a mouse DHFR coding fragment was inserted to obtain DHFR intron expression vectors which differ only in the sequence that
10 overlapping PCR mutagenesis to obtain sequences that match splice donor sites found between exons 3 and 4 of normal and mutant Ras genes. PCR was also used to destroy the splice donor site.

A mouse DHFR cDNA fragment (Simonsen et al., Proc. Natl. Acad. Sci. USA, 80:2495-2499 [1983]) was inserted into the intron of this vector 59
15 nucleotides downstream of the splice donor site. The splice donor site of this vector was altered by mutagenesis to change the ratio of spliced to non-spliced message in transfected cells. It has previously been shown that a single nucleotide change (G to A) converted a relatively efficient splice donor site found in the normal ras gene into an inefficient splice
20 site (Cohen et al., Nature, 334:119-124 [1988]). This effect has been demonstrated in the context of the ras gene and confirmed when these sequences were transferred to human growth hormone constructs (Cohen et al., Cell, 58:461-472 [1989]). Additionally, a non functional 5' splice site (GT to CA) was constructed as a control (Δ GT). A polylinker was
25 inserted 35 nucleotides downstream of the 3' splice site to accept the cDNA of interest. A vector containing tPA (Pennica et al., Nature, 301:214-221 [1983]) was linearized downstream of the polyadenylation site before it was introduced into CHO cells (Potter et al., Proc. Natl. Acad. Sci. USA, 81:7161 [1984]).

30 Plasmid DNA's that contained DHFR/intron, tPA and (a) wild type ras (WT ras), i.e. Figure 3 (SEQ ID NO: 1), (b) mutant ras, or (c) non-functional splice donor site (Δ GT) were introduced into CHO DHFR minus cells by electroporation. The intron vectors were each linearized downstream of the polyadenylation site by restriction endonuclease
35 treatment. The control vector was linearized downstream of the second polyadenylation site. The DNA's were ethanol precipitated after phenol/chloroform extraction and were resuspended in 20 μ l 1/10 Tris EDTA. Then, 10 μ g of DNA was incubated with 10⁷ CHO.dp12 cells (EP 307,247 published 15 March 1989) in 1 ml of PBS on ice for 10 min. before
40 electroporation at 400 volts and 330 μ f using a BRL Cell Porator.

Cells were returned to ice for 10 min. before being plated into non-selective medium. After 24 hours cells were fed nucleoside-free medium to select for stable DHFR+ clones which were pooled. The pooled DHFR+ clones were lysed and mRNA's were prepared.

45 To prepare the mRNA, RNA was extracted from 5 x 10⁷ cells which were grown from pools of more than 200 clones derived from the stable

transfection of the three vectors, the essential construction of which is shown in Figure 1B and from non-transfected CHO cells. RNA was purified over oligo-DT cellulase (Collaborative Biomedical Products). 10µg of mRNA was then subjected to Northern blotting which involved running the mRNA on
5 a 1.2% agarose, 6.6% formaldehyde gel, and transferring it to a nylon filter (Stratagene Duralon-UV membrane), prehybridized, probed and washed according to the manufacturer's instructions.

The filter was probed sequentially using probes (shown in Figure 1B) that would detect (a) the full length message, (b) both full length and
10 spliced message, or (c) beta actin. Probing with the long probe showed that the vector that contains the efficient splice donor site (i.e. WT ras) generates predominately a mRNA of the size predicted for the spliced product while the other two vectors gave rise primarily to a mRNA that corresponds in size to non-spliced message. The DHFR probe detected only
15 full length message and demonstrated that the WT ras splice donor derived vector generates very little full length message with which to confer a DHFR positive phenotype.

Figure 4 shows the number of DHFR positive colonies obtained after duplicate electroporations with the three intron vectors described above
20 and from a conventional vector that has a CMV promoter driving tPA and a SV40 promoter driving DHFR (see Figure 2). The increase in colony number parallels the increase in full length message that accumulates with the modification of the splice donor sites. The conventional vector efficiently generates colonies and does not vary significantly from the ΔGT
25 construct.

The level of tPA expression was determined by seeding cells in 1 ml of F12:DMEM (50:50, with 5% FBS) in 24 well dishes to near confluency. Growth of the cells continued until the media was exhausted. Media was then assayed by ELISA for tPA production. Briefly, anti-tPA antibody was
30 coated onto the wells of an ELISA microtiter plate, media samples were added to the wells followed by washing. Binding of the antigen (tPA) was then quantified using horse radish peroxidase (HRPO) labelled anti-tPA antibody.

Figure 5A depicts the titers of secreted tPA protein after pooling
35 the clones of each group shown in Figure 4. While the number of colonies increased with a weakening of splice donor function, the inverse was seen with respect to tPA expression. The expression levels are consistent with the RNA products that are observed; as more of the dicistronic message is spliced an increased amount of message will contain tPA as the first open
40 reading frame resulting in increased tPA expression. A mutation of GT to CA in the splice donor site results in an abundance of DHFR positive colonies which express undetectable levels of tPA, possibly resulting from inefficient utilization of the second AUG. Importantly, Figure 5A also shows that expression levels obtained from one of the dicistronic vectors
45 (with WT ras SD) was about threefold higher than that obtained with the control vector containing a CMV promoter/enhancer driving tPA, SV40

promoter/enhancer controlling DHFR and SV40 polyadenylation signals controlling the expression of tPA and DHFR.

Additionally, the homogeneity of expression in the pools was investigated. Figure 5B shows that all 20 clones generated by the WT ras splice donor site derived dicistronic vectors express detectable levels of tPA while only 4 of 20 clones generated by the control vector express tPA. None of the clones transfected with the non-splicing (Δ GT) vector expressed tPA levels detectable by ELISA. This finding is consistent with previous observations that relatively few clones generated by conventional vectors make useful levels of protein.

Expression of tPA was increased following methotrexate amplification of pools. Figure 5C shows that 2 of the dicistronic vector derived pools (i.e. with WT ras and MUTANT ras SD sites) increased in expression markedly (8.4 and 7.7 fold), while the pool generated by the conventional vector increased only slightly (2.8 fold) when each was subjected to 200 nM Mtx. An overall increase of 9 fold was obtained using the best dicistronic (WT ras SD) versus the conventional vector following amplification. Growth of the highest expressing amplified pool in nutrient rich production medium yielded titers of 4.2 μ g/ml tPA.

It was shown that manipulation of the splice donor sequence alters the ratio of spliced to full length message and the number of colonies that form in selective medium. It was also shown that dicistronic expression vectors generate clones that express high levels of recombinant proteins. Surprisingly, it was possible to isolate high expressors which had the efficient WT ras splice donor site by selection for DHFR^r cells despite the efficiency with which the DHFR gene was spliced from the RNA precursors formed in these cells.

EXAMPLE 2

TNFr-IgG production using the dicistronic expression vectors

To prove the general applicability of this approach, a second product was evaluated in the dicistronic vector system containing, as the DNA of interest, an immunoadhesin (TNFr-IgG) capable of binding tumor necrosis factor (TNF) (Ashkenazi et al., Proc. Natl. Acad. Sci. USA, 88:10535-10539 [1991]). The experiments described in Example 1 above were essentially repeated except that the product gene encoded the immunoadhesin TNFr-IgG. Plasmid DNA's that contained a TNFr-IgG cDNA and (a) WT ras, i.e. Figure 6 (SEQ ID NO: 2), (b) mutant ras or (c) nonfunctional splice donor site (Δ GT) were introduced into the dp12.CHO cells as discussed for Example 1. See Figure 1C for an illustration of the DNA constructs.

It was discovered that the number of DHFR positive colonies generated by three of these vectors was similar to that seen with the tPA constructs. Expression of TNFr-IgG also paralleled that seen with the tPA constructs (Figure 7A). Amplification of pools from two of the constructs showed a marked increase in expression of immunoadhesin (9.6 and 6.8 fold) (Figure

7B). The best of these amplified pools expressed 9.5 $\mu\text{g/ml}$ when grown in nutrient rich production medium.

Thus, it was again shown that dicistronic expression vectors generate clones that express high levels of recombinant proteins. Furthermore, contrary to expectations, it was discovered that isolation of high product expressing host DHFR⁺ cells was possible using an efficient splice donor site (i.e. the WT ras splice donor site).

EXAMPLE 3

Antibody production using a dicistronic expression vector

10 The usefulness of this system for antibody expression was evaluated by testing production of an antibody directed against IgE (Presta et al., Journal of Immunology, 151:2623-2632 [1993]). Further, the flexibility of the system with regard to transcription initiation was tested by replacing the CMV promoter/enhancer present in the previous vectors with the
15 promoter/ enhancer derived from the early region of SV40 virus (Griffin, B., Structure and Genomic Organization of SV40 and Polyoma Virus, In J. Tooze [Ed] DNA Tumor Viruses, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York). The heavy chain of the antibody was inserted downstream of DHFR as described in the earlier tPA and TNFr-IgG constructs.
20 Additionally, a new splice donor site sequence (GAC:GTAAGT) was engineered into the vector which matches the consensus splice donor site more closely than did the splice donor sites present in the vectors tested in Examples 1 and 2. The resultant expression vector is shown in Figures 1D and 9.

It was discovered that this vector produced fewer colonies than the
25 vectors previously tested, and produced predominantly a spliced RNA product. A second vector was constructed to have the light chain of the antibody under control of the SV40 promoter/enhancer and poly-A and the hygromycin B resistance gene under control of the CMV promoter/enhancer and SV40 poly-A. These vectors were linearized at unique HpaI sites downstream
30 of the poly-A signal, mixed at a ratio of light chain vector to heavy chain vector of 10:3 and electroporated into CHO cells using an optimized protocol (as discussed in Examples 1 and 2).

Figure 11 shows the levels of antibody expressed by clones and pools after selection in hygromycin B followed by selection for DHFR expression.
35 All 20 of the clones analyzed expressed high levels of antibody when grown in rich medium and varied from one another by only a factor of four. A pool of antibody producing clones was generated and assayed shortly after it was established. That pool was grown continuously for 6 weeks without a significant decrease in productivity demonstrating that its stability was
40 sufficient to generate gram quantities of protein from its large scale culture.

The pool was subjected to methotrexate amplification at 200nM and 1 μM and achieved a greater than 2 fold increase in antibody titer. The 1 μM Mtx resistant pool achieved a titer of 41 mg/L when grown under optimal
45 conditions in suspension culture.

The structure of the expressed antibody was examined. Proteins expressed by the 200nM methotrexate resistant pool and by a well characterized expression clone generated by conventional vectors (Presta et al. [1993], supra) were metabolically labeled with S³⁵ cysteine and methionine. In particular, confluent 35mm plates of cells were metabolically labeled with 50μCi each S-35 methionine and S-35 cysteine (Amersham) in serum free cysteine and methionine free F12:DMEM. After one hour, nutrient rich production media was added and labeled proteins were allowed to "chase" into the medium for six more hours. Proteins were run on a 12% SDS/PAGE gel (NOVEX) non-reduced or following reduction with B-mercaptoethanol. Dried gels were exposed to film for 16 hours. CHO control cells were also labeled.

The majority of the antibody protein is secreted with a molecular weight of about 155 kilodaltons, consistent with a properly disulfide-linked antibody molecule with 2 light and 2 heavy chains. Upon reduction the molecular weight shifts to 2 approximately equally abundant proteins of 22.5 and 55 kilodaltons. The protein generated from the pool is indistinguishable from the antibody produced by the well characterized expression clone, with no apparent increase of free heavy or light chain expressed by the pool.

CONCLUSION

The efficient expression system described herein utilizes vectors consisting of promoter/enhancer elements followed by an intron containing the selectable marker coding sequence, followed by the cDNA of interest and a polyadenylation signal.

Several splice donor site sequences were tested for their effect on colony number and expression of the cDNA of interest. A non-functional splice donor site, splice donor sites found in an intron between exons 3 and 4 of mutant (mutant ras) and normal (WT ras) forms of the Harvey Ras gene and another efficient SD site (see Example 3) were used. The vectors were designed to direct expression of dicistronic primary transcripts. Within a transfected cell some of the transcripts remain full length while the remainder are spliced to excise the DHFR coding sequence. When the splice donor site is weakened or destroyed an increase in colony number is observed.

Expression levels show the inverse pattern, with the most efficient splice donor sites generating the highest levels of tPA, TNF α immunoadhesin or anti-IgE V_H.

The homogeneity of expression of clones generated by the ras splice donor site intron DHFR vectors was compared to clones generated from a conventional vector with a separate promoter/enhancer and polyadenylation signal for each DHFR and tPA. The DHFR intron vector gives rise to colonies that are much more homogeneous with regard to expression than those generated by the conventional vector. Non-expressing clones derived from the conventional vector may be the result of breaks in the tPA or

TNFr-IgG domain of the plasmid during integration into the genome or the result of methylation of promoter elements (Busslinger et al., Cell, 34:197-206 [1983]; Watt et al., Genes and Development, 2:1136-1143 [1988]) driving tPA or TNFr-IgG expression. Promoter silencing by methylation or
5 breaks in the DHFR-intron vectors would very likely render them incapable of conferring a DHFR positive phenotype.

It was found that pools generated by the DHFR-intron vectors could be amplified in methotrexate and would increase in expression by a factor of 8.4 (tPA), or 9.8 (TNFr-IgG). Pools from conventional vectors increased
10 by only 2.8 and 3.0 fold for tPA and TNFr-IgG when amplified similarly. Amplified pools resulted in 9 fold higher tPA levels and 15 fold higher TNFr-IgG levels when compared to the conventional vector amplified pools.

Without being limited to any theory, the increase in expression of methotrexate resistant pools derived from the dicistronic vectors is likely
15 due to the transcriptional linkage of DHFR and the product; when cells are selected for increased DHFR expression they consistently over-express product. Conventional approaches lack selectable marker and cDNA expression linkage and therefore methotrexate amplification often generates DHFR overexpression without the concomitant increase in product expression.

20 A further increase of 4 and 6.3 fold in expression were obtained when amplified tPA and TNFr-IgG pools were transferred from the media used for the selections and amplifications to a nutrient rich production medium.

In Example 3, the expression vector had a splice donor site that more closely matches the consensus splice donor sequence and had the heavy chain
25 of a humanized anti-IgE antibody inserted downstream. This vector was linearized and co-electroporated with a second linearized vector that expresses the hygromycin resistance gene and the light chain of the antibody each under the control of its own promoter/enhancer and poly-A signals. An excess of light chain expression vector over the heavy chain
30 dicistronic expression vector was used to bias in favor of light chain expression. Clones and a pool were generated after hygromycin B and DHFR selections. The clones were found to express relatively consistent, high levels of antibody, as did the pool. The 1 μ M pool achieved a titer of 41mg/L when grown under optimal conditions in suspension culture.

35 The anti-IgE antibody was assessed by metabolic labeling followed by SDS/PAGE under reducing and non reducing conditions and found to be indistinguishable from the protein expressed by a highly characterized clonal cell line. Of particular importance is the finding that no free light chain is observed in the pool relative to the clone.

40 A stable expression system for CHO cells has been developed that produces high levels of recombinant proteins rapidly and with less effort than that required by other expression systems. The vector system generates stable clones that express consistently high levels thereby reducing the number of clones that must be screened to obtain a highly
45 productive clonal line. Alternatively, pools have been used to conveniently generate moderate to high levels of protein. This approach

may be particularly useful when a number of related proteins are to be expressed and compared.

Without being limited to this theory, it is possible the vectors that have very efficient splice donor sites generate very productive clones
5 because so little transcript remains non spliced that only integration events that lead to the generation of high levels of RNA produce enough DHFR protein to give rise to colonies in selective medium. The high level of spliced message from such clones is then translated into abundant amounts of the protein of interest. Pools of clones made concurrently by
10 introducing conventional vectors expressed lower levels of protein, and were unstable with regard to long term expression, and expression could not be appreciably increased when the cells were subjected to methotrexate amplification.

The system developed herein is versatile in that it allows high
15 levels of single and multiple subunit polypeptides to be rapidly generated from clones or pools of stable transfectants. This expression system combines the advantages of transient expression systems (rapid and labor non intensive generation of research amounts of protein) with the concurrent development of highly productive stable production cell lines.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- 5 (i) APPLICANT: GENENTECH, INC.
- (ii) TITLE OF INVENTION: METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLS
- 10 (iii) NUMBER OF SEQUENCES: 4
- (iv) CORRESPONDENCE ADDRESS:
- (A) ADDRESSEE: Genentech, Inc.
- (B) STREET: 460 Point San Bruno Blvd
- 15 (C) CITY: South San Francisco
- (D) STATE: California
- (E) COUNTRY: USA
- (F) ZIP: 94080
- (v) COMPUTER READABLE FORM:
- 20 (A) MEDIUM TYPE: 5.25 inch, 360 Kb floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: patin (Genentech)
- 25 (vi) CURRENT APPLICATION DATA:
- (A) APPLICATION NUMBER:
- (B) FILING DATE:
- (C) CLASSIFICATION:
- 30 (vii) PRIOR APPLICATION DATA:
- (A) APPLICATION NUMBER: 08/286740
- (B) FILING DATE: 05-AUG-1994
- (viii) ATTORNEY/AGENT INFORMATION:
- 35 (A) NAME: Lee, Wendy M.
- (B) REGISTRATION NUMBER: 00,000
- (C) REFERENCE/DOCKET NUMBER: 798PCT
- (ix) TELECOMMUNICATION INFORMATION:
- 40 (A) TELEPHONE: 415/225-1994
- (B) TELEFAX: 415/952-9881
- (C) TELEX: 910/371-7168

(2) INFORMATION FOR SEQ ID NO:1:

- 45 (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 7360 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- 50 (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:
- 55 TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50
- TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCGCG GTTACATAAC 100
- 60 TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150
- ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200
- 65 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCAC TTGGCAGTAC 250

ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300

5 AAATGGCCCG CCTGGCATTG TGCCCAGTAC ATGACCTTAT GGGACTTTCC 350

TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400

10 GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450

TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500

15 AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550

AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600

20 TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650

25 CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCCG GAACGGTGCA 700

TTGGAACGCG GATTCCCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA 750

30 GCGATAAGAG GATTTTATCC CCGCTGCCAT CATGGTTCGA CCATTGAACT 800

GCATCGTCGC CGTGTCCCAA AATATGGGGA TTGGCAAGAA CGGAGACCTA 850

35 CCCTGCCCTC CGCTCAGGAA CGCGTTCAAG TACTTCCAAA GAATGACCAC 900

40 AACCTCTTCA GTGGAAGGTA AACAGAATCT GGTGATTATG GGTAGGAAAA 950

CCTGGTTCTC CATTCCTGAG AAGAATCGAC CTTTAAAGGA CAGAATTAAT 1000

45 ATAGTTCTCA GTAGAGAACT CAAAGAACCA CCACGAGGAG CTCATTTTCT 1050

TGCCAAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTGG 1100

50 CAAGTAAAGT AGACATGGTT TGGATAGTCG GAGGCAGTTC TGTTTACCAG 1150

55 GAAGCCATGA ATCAACCAGG CCACCTTAGA CTCTTTGTGA CAAGGATCAT 1200

GCAGGAATTT GAAAGTGACA CGTTTTTCCC AGAAATTGAT TTGGGGAAAT 1250

60 ATAAACCTCT CCCAGAATAC CCAGGCGTCC TCTCTGAGGT CCAGGAGGAA 1300

AAAGGCATCA AGTATAAGTT TGAAGTCTAC GAGAAGAAAG ACTAACAGGA 1350

65 AGATGCTTTC AAGTTCTCTG CTCCCCTCCT AAAGCTATGC ATTTTTATAA 1400

GACCATGGGA CTTTGTGG CTTTAGACCC CCTTGGCTTC GTTAGAACGC 1450

5 GGCTACAATT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA 1500

CACTATAGAA TAACATCCAC TTGCGCTTTC TCTCCACAGG TGCTACTCCA 1550

10 GGTCAACTGC ACCTCGGTTC TAAGCTGGG CTGCAGGTCG CCGTGAATTT 1600

AAGGGACGCT GTGAAGCAAT CATGGATGCA ATGAAGAGAG GGCTCTGCTG 1650

15 TGTGCTGCTG CTGTGTGGAG CAGTCTTCGT TTCGCCCAGC CAGGAAATCC 1700

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20 GAAAAAACGC AGATGATATA CCAGCAACAT CAGTCATGGC TCGCCCTGT 1800

25 GCTCAGAAGC AACCGGTGG AATATTGCTG GTGCAACAGT GGCAGGGCAC 1850

AGTGCCACTC AGTGCCTGTC AAAAGTTGCA GCGAGCCAAG GTGTTTCAAC 1900

30 GGGGGCACCT GCCAGCAGGC CCTGTACTTC TCAGATTTCTG TGTGCCAGTG 1950

CCCCGAAGGA TTTGCTGGGA AGTGCTGTGA AATAGATACC AGGGCCACGT 2000

35 GCTACGAGGA CCAGGGCATC AGCTACAGGG GCACGTGGAG CACAGCGGAG 2050

40 AGTGGCGCCG AGTGCACCAA CTGGAACAGC AGCGCGTTGG CCCAGAAGCC 2100

CTACAGCGGG CGGAGGCCAG ACGCCATCAG GCTGGGCCTG GGAACCACA 2150

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AAGGCGGGGA AGTACAGCTC AGAGTTCTGC AGCACCCCTG CCTGCTCTGA 2250

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5 CCATCTTTGC CAAGCACAGG AGGTCGCCCC GAGAGCGGTT CCTGTGCGGG 2650

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15 GTCCATAAGG AATTCGATGA TGACACTTAC GACAATGACA TTGCGCTGCT 2850

GCAGCTGAAA TCGGATTCTG CCCGCTGTGC CCAGGAGAGC AGCGTGGTCC 2900

20 GCACTGTGTG CCTTCCCCCG GCGGACCTGC AGCTGCCGGA CTGGACGGAG 2950

25 TGTGAGCTCT CCGGCTACGG CAAGCATGAG GCCTTGTCTC CTTTCTATTG 3000

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30 CATCACAACA TTTACTTAAC AGAACAGTCA CCGACAACAT GCTGTGTGCT 3100

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20 CCCAGTTCCG CCCATTCTCC GCCCCATGGC TGAATAATT TTTTATTTA 4100

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25 CATTCAAATA TGTATCCGCT CATGAGACAA TAACCCTGAT AAATGCTTCA 5300

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30 TTATTCCCTT TTTGCGGCA TTTGCGCTC CTGTTTTTGC TCACCCAGAA 5400

ACGCTGGTGA AAGTAAAAGA TGCTGAAGAT CAGTTGGGTG CACGAGTGGG 5450

35 TTACATCGAA CTGGATCTCA ACAGCGGTAA GATCCTTGAG AGTTTTCGCC 5500

40 CCGAAGAACG TTTTCCAATG ATGAGCACTT TTAAAGTTCT GCTATGTGGC 5550

GCGGTATTAT CCCGTGATGA CGCCGGGCAA GAGCAACTCG GTCGCCGCAT 5600

45 ACACTATTCT CAGAATGACT TGGTTGAGTA CTCACCAGTC ACAGAAAAGC 5650

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50 ATGAGTGATA ACACTGCGGC CAACTTACTT CTGACAACGA TCGGAGGACC 5750

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60 GACACCACGA TGCCAGCAGC AATGGCAACA ACGTTGCGCA AACTATTAAC 5900

TGGCGAACTA CTTACTCTAG CTTCCCGGCA ACAATTAATA GACTGGATGG 5950

65 AGGCGGATAA AGTTGCAGGA CCACTTCTGC GCTCGGCCCT TCCGGCTGGC 6000

TGGTTTATTG CTGATAAATC TGGAGCCGGT GAGCGTGGGT CTCGCGGTAT 6050
5 CATTGCAGCA CTGGGGCCAG ATGGTAAGCC CTCCCGTATC GTAGTTATCT 6100
ACACGACGGG GAGTCAGGCA ACTATGGATG AACGAAATAG ACAGATCGCT 6150
10 GAGATAGGTG CCTCACTGAT TAAGCATTGG TAACTGTCAG ACCAAGTTTA 6200
CTCATATATA CTTTAGATTG ATTTAAAACT TCATTTTAA TTTAAAAGGA 6250
15 TCTAGGTGAA GATCCTTTTT GATAATCTCA TGACCAAAT CCCTTAACGT 6300
GAGTTTTCGT TCCACTGAGC GTCAGACCCC GTAGAAAAGA TCAAAGGATC 6350
20 TTCTTGAGAT CTTTTTTTC TCGCGTAAT CTGCTGCTTG CAAACAAAA 6400
25 AACCACCGCT ACCAGCGGTG GTTTGTTTGC CGGATCAAGA GCTACCAACT 6450
CTTTTTCCGA AGGTAAGTGG CTTCAAGCAGA GCGCAGATAC CAAATACTGT 6500
30 CCTTCTAGTG TAGCCGTAGT TAGGCCACCA CTTCAAGAAC TCTGTAGCAC 6550
CGCCTACATA CCTCGCTCTG CTAATCCTGT TACCAGTGGC TGCTGCCAGT 6600
35 GCGGATAAGT CGTGTCTTAC CGGGTTGGAC TCAAGACGAT AGTTACCGGA 6650
40 TAAGGCGCAG CGGTCGGGCT GAACGGGGG TTCGTGCACA CAGCCCAGCT 6700
TGGAGCGAAC GACCTACACC GAACTGAGAT ACCTACAGCG TGAGCATTGA 6750
45 GAAAGCGCCA CGCTTCCCGA AGGGAGAAAG GCGGACAGGT ATCCGGTAAG 6800
CGGCAGGGTC GGAACAGGAG AGCGCACGAG GGAGCTTCCA GGGGAAACG 6850
50 CCTGGTATCT TTATAGTCCT GTCGGGTTTC GCCACCTCTG ACTTGAGCGT 6900
55 CGATTTTTGT GATGCTCGTC AGGGGGGCGG AGCCTATGGA AAAACGCCAG 6950
CAACGCGGCC TTTTACGGT TCCTGGCCTT TTGCTGGCCT TTTGCTCACA 7000
60 TGTTCCTTCC TGCCTTATCC CCTGATTCTG TGGATAACCG TATTACCGCC 7050
TTTGAGTGAG CTGATACCGC TCGCCGAGC CGAACGACCG AGCGCAGCGA 7100
65 GTCAGTGAGC GAGGAAGCGG AAGAGCGCCC AATACGCAA CCGCCTCTCC 7150

CCGCGCGTTG GCCGATTCAT TAATCCAGCT GGCACGACAG GTTTCCCGAC 7200
5 TGGAAAGCGG GCAGTGAGCG CAACGCAATT AATGTGAGTT ACCTCACTCA 7250
TTAGGCACCC CAGGCTTTAC ACTTTATGCT TCCGGCTCGT ATGTTGTGTG 7300
10 GAATTGTGAG CGGATAACAA TTTCACACAG GAAACAGCTA TGACCATGAT 7350
TACGAATTAA 7360

15

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:
20 (A) LENGTH: 6889 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50
30 TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC 100
TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150
35 ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200
40 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC 250
ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300
45 AAATGGCCCG CCTGGCATTG TGCCAGTAC ATGACCTTAT GGGACTTTCC 350
TACTTGCCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400
50 GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450
55 TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500
AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550
60 AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600
TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650
65 CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCCG GAACGGTGCA 700

TTGGAACGCG GATTCCCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA 750

5 GCGATAAGAG GATTTTATCC CCGCTGCCAT CATGGTTCGA CCATTGAACT 800

GCATCGTCGC CGTGTCCCAA AATATGGGGA TTGGCAAGAA CGGAGACCTA 850

10 CCCTGCCCTC CGCTCAGGAA CGCGTTCAAG TACTTCCAAA GAATGACCAC 900

AACCTCTTCA GTGGAAGGTA AACAGAATCT GGTGATTATG GGTAGGAAAA 950

15 CCTGGTTCTC CATTCCTGAG AAGAATCGAC CTTTAAAGGA CAGAATTAAT 1000

ATAGTTCTCA GTAGAGAACT CAAAGAACCA CCACGAGGAG CTCATTTTCT 1050

20 TGCCAAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTGG 1100

25 CAAGTAAAGT AGACATGGTT TGGATAGTCG GAGGCAGTTC TGTTTACCAG 1150

GAAGCCATGA ATCAACCAGG CCACCTTAGA CTCTTTGTGA CAAGGATCAT 1200

30 GCAGGAATTT GAAAGTGACA CGTTTTTCCC AGAAATTGAT TTGGGGAAAT 1250

ATAAACCTCT CCCAGAATAC CCAGGCGTCC TCTCTGAGGT CCAGGAGGAA 1300

35 AAAGGCATCA AGTATAAGTT TGAAGTCTAC GAGAAGAAAG ACTAACAGGA 1350

40 AGATGCTTTC AAGTTCTCTG CTCCCCTCCT AAAGCTATGC ATTTTATATA 1400

GACCATGGGA CTTTGTCTGG CTTTAGACCC CTTGGCTTC GTTAGAACGC 1450

45 GGCTACAATT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA 1500

CÂCTATAGAA TAACATCCAC TTTGCCTTTC TCTCCACAGG TGTCACTCCA 1550

50 GGTCAACTGC ACCTCGGTTT TATCGATTGA ATTCCCCGGC CATAGCTGTC 1600

55 TGGCATGGGC CTCTCCACCG TGCCTGACCT GCTGCTGCCG CTGGTGCTCC 1650

TGGAGCTGTT GGTGGGAATA TACCCCTCAG GGGTTATTGG ACTGGTCCCT 1700

60 CACCTAGGGG ACAGGGAGAA GAGAGATAGT GTGTGTCCCC AAGGAAAATA 1750

TATCCACCCT CAAAATAATT CGATTGCTG TACCAAGTGC CACAAAGGAA 1800

65 CCTÂCTTGTA CAATGACTGT CCAGGCCCGG GGCAGGATAC GGAAGTGCAGG 1850

GAGTGTGAGA GCGGCTCCTT CACCGCTTCA GAAAACCACC TCAGACACTG 1900
CCTCAGCTGC TCCAAATGCC GAAAGGAAAT GGGTCAGGTG GAGATCTCTT 1950
5 CTTGCACAGT GGACCGGGAC ACCGTGTGTG GCTGCAGGAA GAACCAGTAC 2000
CGGCATTATT GGAGTGAAAA CCTTTTCCAG TGCTTCAATT GCAGCCTCTG 2050
CCTCAATGGG ACCGTGCACC TCTCCTGCCA GGAGAAACAG AACACCGTGT 2100
15 GCACCTGCCA TGCAGGTTC TTTCTAAGAG AAAACGAGTG TGTCTCCTGT 2150
AGTAACTGTA AGAAAAGCCT GGAGTGCACG AAGTTGTGCC TACCCAGAT 2200
20 TGAGAATGTT AAGGGCACTG AGGACTCAGG CACCACAGAC AAGAGAGTTG 2250
AGCTCAAAAC CCCACTTGGT GACACAATC ACACATGCCC ACGGTGCCCC 2300
GAGCCCAAAT CTTGTGACAC ACCTCCCCCG TGCCACGGT GCCCAGAGCC 2350
30 CAAATCTTGT GACACACCTC CCCCATGCCC ACGGTGCCCC GAGCCCAAAT 2400
CTTGTGACAC ACCTCCCCCA TGCCACGGT GCCCAGCACC TGAATCCTG 2450
35 GGAGGACCGT CAGTCTTCCT CTTCCCCCA AAACCCAAGG ATACCCTTAT 2500
GATTTCCCGG ACCCCTGAGG TCACGTGCGT GGTGGTGGAC GTGAGCCACG 2550
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45 AATGCCAAGA CAAAGCCGCG GGAGGAGCAG TTCAACAGCA CGTTCCGTGT 2650
GGTCAGCGTC CTCACCGTCC TGCACCAGGA CTGGCTGAAC GGCAAGGAGT 2700
50 ACAAGTGCAA GGTCTCCAAC AAAGCCCTCC CAGCCCCCAT CGAGAAAACC 2750
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60 TCAAAGGCTT CTACCCAGC GACATCGCCG TGGAGTGGGA GAGCAGCGGG 2900
CAGCCGGAGA ACAACTACAA CACCACGCCT CCCATGCTGG ACTCCGACGG 2950
65 CTCCTTCTTC CTCTACAGCA AGCTCACCGT GGACAAGAGC AGGTGGCAGC 3000

AGGGGAACAT CTTCTCATGC TCCGTGATGC ATGAGGCTCT GCACAACCGC 3050
5 TTCACGCAGA AGAGCCTCTC CCTGTCTCCG GGTAAATGAG TGCACGGCC 3100
GGGGATCCTC TAGAGTCGAC CTGCAGAAGC TTGGCCGCCA TGGCCCAACT 3150
10 TGTTTATTGC AGCTTATAAT GGTACAAAT AAAGCAATAG CATCACAAAT 3200
TTCACAAATA AAGCATTTTT TCACTGCAT TCTAGTTGTG GTTTGTCCAA 3250
15 ACTCATCAAT GTATCTTATC ATGTCTGGAT CGATCGGGAA TTAATTCGGC 3300
GCAGCACCAT GGCCTGAAAT AACCTCTGAA AGAGGAACTT GGTTAGGTAC 3350
20 CTTCTGAGGC GGAAAGAACC AGCTGTGGAA TGTGTGTCAG TTAGGGTGTG 3400
25 GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA GTATGCAAAG CATGCATCTC 3450
AATTAGTCAG CAACCAGGTG TGGAAAGTCC CCAGGCTCCC CAGCAGGCAG 3500
30 AAGTATGCAA AGCATGCATC TCAATTAGTC AGCAACCATA GTCCCGCCCC 3550
TAACTCCGCC CATCCCGCCC CTAACTCCGC CCAGTTCCGC CCATTCTCCG 3600
35 CCCCATGGCT GACTAATTTT TTTTATTTAT GCAGAGGCCG AGGCCGCCTC 3650
40 GGCCTCTGAG CTATTCCAGA AGTAGTGAGG AGGCTTTTTT GGAGGCCTAG 3700
GCTTTTGCAA AAAGCTGTTA ACAGCTTGGC ACTGGCCGTC GTTTTACAAC 3750
45 GTCGTGACTG GGAAAACCCT GCGGTTACCC AACTTAATCG CCTTGACGCA 3800
CATCCCCCCT TCGCCAGCTG GCGTAATAGC GAAGAGGCCC GCACCGATCG 3850
50 CCCTTCCCAA CAGTTGCGTA GCCTGAATGG CGAATGGCGC CTGATGCGGT 3900
55 ATTTTCTCCT TACGCATCTG TCGGTATTT CACACCGCAT ACGTCAAAGC 3950
AACCATAGTA CGCGCCCTGT AGCGGCGCAT TAAGCGCGGC GGGTGTGGTG 4000
60 GTTACGCGCA GCGTGACCGC TACACTTGCC AGCGCCCTAG CGCCCGCTCC 4050
TTTCGCTTTC TTCCCTTCCT TTCTCGCCAC GTTCGCCGGC TTTCCCGTC 4100
65 AAGCTCTAAA TCGGGGGCTC CCTTTAGGGT TCCGATTTAG TGCTTTACGG 4150

CACCTCGACC CCAAAAAACT TGATTGGGT GATGGTTCAC GTAGTGGGCC 4200
5 ATCGCCCTGA TAGACGGTTT TTCGCCCTTT GACGTTGGAG TCCACGTCT 4250
TTAATAGTGG ACTCTGTTC CAAACTGGAA CAACACTCAA CCCTATCTCG 4300
10 GGCTATTCTT TTGATTTATA AGGGATTTTG CCGATTTCGG CCTATTGGTT 4350
AAAAATGAG CTGATTTAAC AAAAATTTAA CGCGAATTTT AACAAAATAT 4400
15 TAACGTTTAC AATTTTATGG TGCACTCTCA GTACAATCTG CTCTGATGCC 4450
GCATAGTTAA GCCAACTCCG CTATCGCTAC GTGACTGGGT CATGGCTGCG 4500
20 CCCCAGACACC CGCCAACACC CGCTGACGCG CCCTGACGGG CTTGTCTGCT 4550
25 CCCGGCATCC GCTTACAGAC AAGCTGTGAC CGTCTCCGGG AGCTGCATGT 4600
GTCAGAGGTT TTCACCGTCA TCACCGAAAC GCGCGAGGCA GTATTCTTGA 4650
30 AGACGAAAGG GCCTCGTGAT ACGCCTATTT TTATAGGTTA ATGTCATGAT 4700
AATAATGGTT TCTTAGACGT CAGGTGGCAC TTTTCGGGGA AATGTGCGCG 4750
35 GAACCCCTAT TTGTTTATTT TTCTAAATAC ATTCAAATAT GTATCCGCTC 4800
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TATGAGTATT CAACATTTCC GTGTCGCCCT TATTCCTTT TTTGCGGCAT 4900
45 TTTGCCTTCC TGTTTTTGCT CACCCAGAAA CGCTGGTGAA AGTAAAAGAT 4950
GCTGAAGATC AGTTGGGTGC ACGAGTGGGT TACATCGAAC TGGATCTCAA 5000
50 CAGCGGTAAG ATCCTTGAGA GTTTTCGCCC CGAAGAACGT TTTCCAATGA 5050
TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGATGAC 5100
GCCGGGCAAG AGCAACTCGG TCGCCGCATA CACTATTCTC AGAATGACTT 5150
60 GGTGAGTAC TCACCAGTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG 5200
TAAGAGAATT ATGCAGTGCT GCCATAACCA TGAGTGATAA CACTGCGGCC 5250
65 AACTTACTTC TGACAACGAT CGGAGGACCG AAGGAGCTAA CCGCTTTTTT 5300

GCACAACATG GGGGATCATG TAACTCGCCT TGATCGTTGG GAACCGGAGC 5350

5 TGAATGAAGC CATAACAAAC GACGAGCGTG ACACCACGAT GCCAGCAGCA 5400

ATGGCAACAA CGTTGCGCAA ACTATTAACT GGCGAACTAC TTACTCTAGC 5450

10 TTCCCGGCAA CAATTAATAG ACTGGATGGA GGCGGATAAA GTTGCAGGAC 5500

CACTTCTGCG CTCGGCCCTT CCGGCTGGCT GGTATTATTC TGATAAATCT 5550

15 GGAGCCGGTG AGCGTGGGTC TCGCGGTATC ATTGCAGCAC TGGGGCCAGA 5600

TGGTAAGCCC TCCCGTATCG TAGTTATCTA CACGACGGGG AGTCAGGCAA 5650

20 CTATGGATGA ACGAAATAGA CAGATCGCTG AGATAGGTGC CTCACTGATT 5700

25 AAGCATTGGT AACTGTCAGA CCAAGTTTAC TCATATATAC TTTAGATTGA 5750

TTTAAACTT CATTTTAAAT TAAAAGGAT CTAGGTGAAG ATCCTTTTTG 5800

30 ATAATCTCAT GACCAAATC CCTTAACGTG AGTTTTCGTT CCACTGAGCG 5850

TCAGACCCCG TAGAAAAGAT CAAAGGATCT TCTTGAGATC CTTTTTTTCT 5900

35 GCGCGTAATC TGCTGCTTGC AAACAAAAAA ACCACCGCTA CCAGCGGTGG 5950

40 TTTGTTTGCC GGATCAAGAG CTACCAACTC TTTTCCGAA GGTAAGTGGC 6000

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45 AGGCCACCAC TTCAAGAACT CTGTAGCACC GCCTACATAC CTCGCTCTGC 6100

TAATCCTGTT ACCAGTGGCT GCTGCCAGTG GCGATAAGTC GTGTCTTACC 6150

50 GGGTTGGACT CAAGACGATA GTTACCGGAT AAGGCGCAGC GGTGGGGCTG 6200

55 AACGGGGGGT TCGTGACAC AGCCAGCTT GGAGCGAACG ACCTACACCG 6250

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60 GGGAGAAAGG CGGACAGGTA TCCGGTAAGC GGCAGGGTCG GAACAGGAGA 6350

GCGCACGAGG GAGCTTCCAG GGGGAAACGC CTGGTATCTT TATAGTCCTG 6400

65 TCGGGTTTCG CCACCTCTGA CTTGAGCGTC GATTTTGTG ATGCTCGTCA 6450

GGGGGGCGGA GCCTATGGAA AAACGCCAGC AACGCGGCCT TTTTACGGTT 6500
5 CCTGGCCTTT TGCTGGCCTT TTGCTCACAT GTTCTTTCCT GCGTTATCCC 6550
CTGATTCTGT GGATAACCGT ATTACCGCCT TTGAGTGAGC TGATAACCGCT 6600
10 CGCCGCAGCC GAACGACCGA GCGCAGCGAG TCAGTGAGCG AGGAAGCGGA 6650
AGAGCGCCCA ATACGCAAAC CGCCTCTCCC CGCGCGTTGG CCGATTCAAT 6700
15 AATCCAGCTG GCACGACAGG TTTCCCGACT GGAAAGCGGG CAGTGAGCGC 6750
AACGCAATTA ATGTGAGTTA CCTCACTCAT TAGGCACCCC AGGCTTTACA 6800
20 CTTTATGCTT CCGGCTCGTA TGTGTGTGG AATTGTGAGC GGATAACAAT 6850
25 TTCACACAGG AAACAGCTAT GACCATGATT ACGAATTAA 6889

(2) INFORMATION FOR SEQ ID NO:3:

30 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 6557 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
35 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

40 TTCGAGCTCG CCCGACATTG ATTATTGACT AGAGTCGATC GACAGCTGTG 50
GAATGTGTGT CAGTTAGGGT GTGGAAAGTC CCCAGGCTCC CCAGCAGGCA 100
45 GAAGTATGCA AAGCATGCAT CTCAATTAGT CAGCAACCAG GTGTGGAAAG 150
TCCCAGGCT CCCAGCAGG CAGAAGTATG CAAAGCATGC ATCTCAATTA 200
50 GTCAGCAACC ATAGTCCCGC CCCTAACTCC GCCCATCCCG CCCCTAACTC 250
55 CGCCCAGTTC CGCCCATTCT CCGCCCCATG GCTGACTAAT TTTTTTTATT 300
TATGCAGAGG CCGAGGCCGC CTCGGCCTCT GAGCTATTCC AGAAGTAGTG 350
60 AGGAGGCTTT TTTGGAGGCC TAGGCTTTTG CAAAAGCTA GCTTATCCGG 400
CCGGAACGG TGCAATGGAA CGCGGATTCC CCGTGCCAAG AGTGACGTAA 450
65 GTACCGCCTA TAGAGCGATA AGAGGATTTT ATCCCCGCTG CCATCATGGT 500

TCGACCATTG AACTGCATCG TCGCCGTGTC CCAAAATATG GGGATTGGCA 550
5 AGAACGGAGA CCTACCCTGG CCTCCGCTCA GGAACGAGTT CAAGTACTTC 600
CAAAGAATGA CCACAACCTC TTCAGTGGAA GGTAAACAGA ATCTGGTGAT 650
10 TATGGGTAGG AAAACCTGGT TCTCCATTCC TGAGAAGAAT CGACCTTTAA 700
AGGACAGAAT TAATATAGTT CTCAGTAGAG AACTCAAAGA ACCACCACGA 750
15 GGAGCTCATT TTCTTGCCAA AAGTTTGGAT GATGCCTTAA GACTTATTGA 800
ACAACCGGAA TTGGCAAGTA AAGTAGACAT GGTTTGGATA GTCGGAGGCA 850
20 GTTCTGTTTA CCAGGAAGCC ATGAATCAAC CAGGCCACCT TAGACTCTTT 900
25 GTGACAAGGA TCATGCAGGA ATTTGAAAGT GACACGTTTT TCCCAGAAAT 950
TGATTTGGGG AAATATAAAC CTCTCCCAGA ATACCCAGGC GTCCTCTCTG 1000
30 AGGTCCAGGA GGAAAAGGC ATCAAGTATA AGTTTGAAGT CTACGAGAAG 1050
AAAGACTAAC AGGAAGATGC TTTCAAGTTC TCTGCTCCCC TCCTAAAGCT 1100
35 ATGCATTTTT ATAAGACCAT GGGACTTTTG CTGGCTTTAG ATCCCCTTGG 1150
40 CTTGTTTAGA ACGCAGCTAC AATTAATACA TAACCTTATG TATCATACAC 1200
ATACGATTTA GGTGACACTA TAGATAACAT CCACTTTGCC TTTCTCTCCA 1250
45 CAGGTGTCCA CTCCCAGGTC CAACTGCACC TCGTTCTAT CGATTGAATT 1300
CCACCATGGG ATGGTCATGT ATCATCCTTT TTCTAGTAGC AACTGCAACT 1350
50 GGAGTACATT CAGAAGTTCA GCTGGTGGAG TCTGGCGGTG GCCTGGTGCA 1400
GCCAGGGGGC TCACTCCGTT TGTCCTGTGC AGTTTCTGGC TACTCCATCA 1450
CCTCCGGATA TAGCTGGAAC TGGATCCGTC AGGCCCCGGG TAAGGGCCTG 1500
60 GAATGGGTTG CATCGATTAC GTATGCCGGA TCGACTAACT ATAACCCTAG 1550
CGTCAAGGGC CGTATCACTA TAAGTCGCGA CGATTCCAAA AACACATTCT 1600
65 ACCTGCAGAT GAACAGCCTG CGTGCTGAGG AACTGCCCGT CTATTATTGT 1650

GCTCGAGGCA GCCACTATTT CGGCGCCTGG CACTTCGCCG TGTGGGGTCA 1700
5 AGGAACCCTG GTCACCGTCT CCTCGGCCTC CACCAAGGGC CCATCGGTCT 1750
TCCCCCTGGC ACCCTCCTCC AAGAGCACCT CTGGGGGCAC AGCGGCCCTG 1800
10 GGCTGCCTGG TCAAGGACTA CTTCCCCGAA CCGGTGACGG TGTCGTGGAA 1850
CTCAGGCGCC CTGACCAGCG GCGTGACAC CTTCCCGCT GTCCTACAGT 1900
15 CCTCAGGACT CTA CTCTCCCTC AGCAGCGTGG TGA CTGTGCTC CTCTAGCAGC 1950
TTGGGCACCC AGACCTACAT CTGCAACGTG AATCACAAGC CCAGCAACAC 2000
20 CAAGGTGGAC AAGAAAGTTG AGCCCAAATC TTGTGACAAA ACTCACACAT 2050
25 GCCCACCGTG CCCAGCACCT GAACTCCTGG GGGGACCGTC AGTCTTCTC 2100
TTCCCCCAA AACCCAAGGA CACCCTCATG ATCTCCCGA CCCCTGAGGT 2150
30 CACATGCGTG GTGGTGGACG TGAGCCACGA AGACCCTGAG GTCAAGTTCA 2200
ACTGGTACGT GGACGGCGTG GAGGTGCATA ATGCCAAGAC AAAGCCGCGG 2250
35 GAGGAGCAGT ACAACAGCAC GTACCGTGTG GTCAGCGTCC TCACCGTCCT 2300
40 GCACCAGGAC TGGCTGAATG GCAAGGAGTA CAAGTGCAAG GTCTCCAACA 2350
AAGCCCTCCC AGCCCCATC GAGAAAACCA TCTCCAAAGC CAAAGGGCAG 2400
45 CCCCAGAGAAC CACAGGTGTA CACCCTGCCC CCATCCCGGG AAGAGATGAC 2450
CAAGAACCAG GTCAGCCTGA CCTGCCTGGT CAAAGGCTTC TATCCCAGCG 2500
50 ACATCGCCGT GGAGTGGGAG AGCAATGGGC AGCCGAGAG CAACTACAAG 2550
55 ACCACGCCTC CCGTGCTGGA CTCCGACGGC TCCTTCTTCC TCTACAGCAA 2600
GCTCACCGTG GACAAGAGCA GGTGGCAGCA GGGGAACGTC TTCTCATGCT 2650
60 CCGTGATGCA TGAGGCTCTG CACAACCACT ACACGCAGAA GAGCCTCTCC 2700
CTGTCTCCGG GTAAATGAGT GCGACGGCCC TAGAGTCGAC CTGCAGAAGC 2750
65 TTGGCCGCCA TGGCCCAACT TGTTTATTGC AGCTTATAAT GGTTACAAAT 2800

AAAGCAATAG CATCACAAAT TTCACAAATA AAGCATTTTT TCACTGCAT 2850

5 TCTAGTTGTG GTTTGTCCAA ACTCATCAAT GTATCTTATC ATGTCTGGAT 2900

CGATCGGGAA TTAATTCGGC GCAGCACCAT GGCCTGAAAT AACCTCTGAA 2950

10 AGAGGAACTT GGTTAGGTAC CTTCTGAGGC GGAAAGAACC AGCTGTGGAA 3000

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15 GTATGCAAAG CATGCATCTC AATTAGTCAG CAACCAGGTG TGGAAAGTCC 3100

CCAGGCTCCC CAGCAGGCAG AAGTATGCAA AGCATGCATC TCAATTAGTC 3150

20 AGCAACCATA GTCCCGCCCC TAACTCCGCC CATCCCGCCC CTAACTCCGC 3200

25 CCAGTTCCGC CCATTCTCCG CCCCATGGCT GACTAATTTT TTTATTTAT 3250

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30 AGGCTTTTTT GGAGGCCTAG GCTTTTGCAA AAAGCTGTTA CCTCGAGCGG 3350

CCGCTTAATT AAGGCGCGCC ATTTAAATCC TGCAGGTAAC AGCTTGGCAC 3400

35 TGGCCGTCGT TTTACAACGT CGTGA CTGGG AAAACCCTGG CGTTACCCAA 3450

40 CTTAATCGCC TTGCAGCACA TCCCCCTTC GCCAGCTGGC GTAATAGCGA 3500

AGAGGCCCCG ACCGATCGCC CTTCCCAACA GTTGCGTAGC CTGAATGGCG 3550

45 AATGGCGCCT GATGCGGTAT TTTCTCCTTA CGCATCTGTG CGGTATTTCA 3600

CACCGCATAC GTCAAAGCAA CCATAGTACG CGCCCTGTAG CGGCGCATT 3650

50 AGCGCGGCGG GTGTGGTGGT TACGCGCAGC GTGACCGCTA CACTTGCCAG 3700

55 CGCCCTAGCG CCCGCTCCTT TCGCTTTCTT CCCTTCCTTT CTCGCCACGT 3750

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60 CGATTTAGTG CTTTACGGCA CCTCGACCCC AAAAACTTG ATTTGGGTGA 3850

TGGTTCACGT AGTGGGCCAT CGCCCTGATA GACGGTTTTT CGCCCTTGA 3900

65 CGTTGGAGTC CACGTTCTTT AATAGTGGAC TCTTGTCCA AACTGGAACA 3950

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5 GATTTTCGGCC TATTGGTTAA AAAATGAGCT GATTTAACAA AAATTTAACG 4050
CGAATTTTAA CAAAATATTA ACGTTTACAA TTTTATGGTG CACTCTCAGT 4100
10 ACAATCTGCT CTGATGCCGC ATAGTTAAGC CAACTCCGCT ATCGCTACGT 4150
GACTGGGTCA TGGCTGCGCC CCGACACCCG CCAACACCCG CTGACGCGCC 4200
15 CTGACGGGCT TGTCTGCTCC CGGCATCCGC TTACAGACAA GCTGTGACCG 4250
TCTCCGGGAG CTGCATGTGT CAGAGGTTTT CACCGTCATC ACCGAAACGC 4300
20 GCGAGGCAGT ATTCTTGAAG ACGAAAGGGC CTCGTGATAC GCCTATTTTT 4350
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TTCGGGGAAA TGTGCGCGGA ACCCCTATTT GTTTATTTTT CTAAATACAT 4450
30 TCAAATATGT ATCCGCTCAT GAGACAATAA CCCTGATAAA TGCTTCAATA 4500
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35 TTCCCTTTTT TCGCGCATTT TGCCTTCCTG TTTTGTCTCA CCCAGAAACG 4600
40 CTGGTGAAAG TAAAAGATGC TGAAGATCAG TTGGGTGCAC GAGTGGGTTA 4650
CATCGAACTG GATCTCAACA GCGGTAAGAT CCTTGAGAGT TTTCGCCCCG 4700
45 AAGAACGTTT TCCAATGATG AGCACTTTTA AAGTTCTGCT ATGTGGCGCG 4750
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50 CTATTCTCAG AATGACTTGG TTGAGTACTC ACCAGTCACA GAAAAGCATC 4850
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60 GGAGCTAACC GCTTTTTTGC ACAACATGGG GGATCATGTA ACTCGCCTTG 5000
ATCGTTGGGA ACCGGAGCTG AATGAAGCCA TACCAAACGA CGAGCGTGAC 5050
65 ACCACGATGC CAGCAGCAAT GGCAACAACG TTGCGCAAAC TATTAAGTGG 5100

CGAACTACTT ACTCTAGCTT CCCGGCAACA ATTAATAGAC TGGATGGAGG 5150

5 CGGATAAAGT TGCAGGACCA CTTCTGCGCT CGGCCCTTCC GGCTGGCTGG 5200

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10 TGCAGCACTG GGGCCAGATG GTAAGCCCTC CCGTATCGTA GTTATCTACA 5300

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15 ATAGGTGCCT CACTGATTAA GCATTGGTAA CTGTCAGACC AAGTTTACTC 5400

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20 AGGTGAAGAT CCTTTTGTAT AATCTCATGA CCAAAATCCC TTAACGTGAG 5500

25 TTTTCGTTCC ACTGAGCGTC AGACCCCGTA GAAAAGATCA AAGGATCTTC 5550

TTGAGATCCT TTTTTTCTGC GCGTAATCTG CTGCTTGCAA ACAAAAAAAC 5600

30 CACCGCTACC AGCGGTGGTT TGTTTGCCGG ATCAAGAGCT ACCAACTCTT 5650

TTTCCGAAGG TAACTGGCTT CAGCAGAGCG CAGATACCAA ATACTGTCCT 5700

35 TCTAGTGTAG CCGTAGTTAG GCCACCACTT CAAGAACTCT GTAGCACCGC 5750

40 CTACATACCT CGCTCTGCTA ATCCTGTTAC CAGTGGCTGC TGCCAGTGGC 5800

GATAAGTCGT GTCTTACCGG GTTGGACTCA AGACGATAGT TACCGGATAA 5850

45 GGCGCAGCGG TCGGGCTGAA CGGGGGGTTC GTGCACACAG CCCAGCTTGG 5900

AGCGAACGAC CTACACCGAA CTGAGATACC TACAGCGTGA GCATTGAGAA 5950

50 AGCGCCACGC TTCCCGAAGG GAGAAAGGCG GACAGGTATC CGGTAAGCGG 6000

55 CAGGGTCGGA ACAGGAGAGC GCACGAGGGA GCTTCCAGGG GGAAACGCCT 6050

GGTATCTTTA TAGTCCTGTC GGGTTTCGCC ACCTCTGACT TGAGCGTCGA 6100

60 TTTTGTGAT GCTCGTCAGG GGGGCGGAGC CTATGGAAAA ACGCCAGCAA 6150

CGCGGCCTTT TTACGGTTCC TGGCCTTTTG CTGGCCTTTT GCTCACATGT 6200

65 TCTTTCCTGC GTTATCCCCT GATTCTGTGG ATAACCGTAT TACCGCCTTT 6250

GAGTGAGCTG ATACCGCTCG CCGCAGCCGA ACGACCGAGC GCAGCGAGTC 6300
AGTGAGCGAG GAAGCGGAAG AGCGCCCAAT ACGCAAACCG CCTCTCCCCG 6350
5 CGCGTTGGCC GATTCATTAA TCCAGCTGGC ACGACAGGTT TCCCGACTGG 6400
10 AAAGCGGGCA GTGAGCGCAA CGCAATTAAT GTGAGTTACC TCACTCATTA 6450
GGCACCCAG GCTTTACACT TTATGCTTCC GGCTCGTATG TTGTGTGGAA 6500
15 TTGTGAGCGG ATAACAATT CACACAGGAA ACAGCTATGA CCATGATTAC 6550
GAATTAA 6557
20

(2) INFORMATION FOR SEQ ID NO:4:

25 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 7305 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
30

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

35 TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50
TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC 100
40 TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150
ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200
45 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC 250
ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300
50 AAATGGCCCG CCTGGCATTG TGCCAGTAC ATGACCTTAT GGGACTTTCC 350
TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400
GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450
60 TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500
AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550
65 AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600

TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650

CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCGG GAACGGTGCA 700

5 TTGGAACGCG GATTCCCCGT GCCAAGAGTG ACGTAAGTAC CGCCTATAGA 750

GTCTATAGGC CCACCCCCTT GGCTTCGTTA GAACGCGGCT ACAATTAATA 800

CATAACCTTA TGTATCATAC ACATACGATT TAGGTGACAC TATAGAATAA 850

15 CATCCACTTT GCCTTTCTCT CCACAGGTGT CCACTCCCAG GTCCAACTGC 900

ACCTCGGTTT TAAGCTTATC GATATGAAA AGCCTGAACT CACCGCGACG 950

20 TCTGTGAGA AGTTTCTGAT CGAAAAGTTC GACAGCGTCT CCGACCTGAT 1000

GCAGCTCTCG GAGGGCGAAG AATCTCGTGC TTTCAGCTTC GATGTAGGAG 1050

GGCGTGGATA TGTCTGCGG GTAAATAGCT GCGCCGATGG TTTCTACAAA 1100

30 GATCGTTATG TTTATCGGCA CTTTGCATCG GCCGCGCTCC CGATTCCGGA 1150

AGTGCTTGAC ATTGGGGAAT TCAGCGAGAG CCTGACCTAT TGCATCTCCC 1200

35 GCCGTGCACA GGGTGTACG TTGCAACACC TGCCTGAAAC CGAACTGCCC 1250

GCTGTTCTGC AGCCGGTCGC GGAGGCCATG GATGCGATCG CTGCGGCCGA 1300

TCTTAGCCAG ACGAGCGGGT TCGGCCCAT TCGGACCGCAA GGAATCGGTC 1350

45 AATACACTAC ATGGCGTGAT TTCATATGCG CGATTGCTGA TCCCCATGTG 1400

TATCACTGGC AAACGTGAT GGACGACACC GTCAGTGCGT CCGTCGCGCA 1450

50 GGCTCTCGAT GAGCTGATGC TTTGGGCCGA GGAAGTCCGGC 1500

ACCTCGTGCA CGCGGATTTC GGCTCCAACA ATGTCCTGAC GGACAATGGC 1550

CGCATAACAG CGGTCATTGA CTGGAGCGAG GCGATGTTTC GGGATTCCCA 1600

60 ATACGAGGTC GCCAACATCT TCTTCTGGAG GCCGTGGTTG GCTTGTATGG 1650

AGCAGCAGAC GACTTTCGAG CGGAGGCATC CGGAGCTTGC AGGATCGCCG 1700

65 CGGCTCCGGG CGTATATGCT CCGCATTGGT CTTGACCAAC TCTATCAGAG 1750

CTTGTTGAC GGCAATTTTCG ATGATGCAGC TTGGGCGCAG GGTTCGATGCG 1800
ACGCAATCGT CCGATCCGGA GCCGGGACTG TCGGGCGTAC ACAAATCGCC 1850
5 CGCAGAAGCG CGGCCGTCTG GACCGATGGC TGTGTAGAAG TACTCGCCGA 1900
TAGTGGAAAC CGACGCCCCA GCACTCGTCC GAGGGCAAAG GAATAGAGTA 1950
GATGCCGACC GAAGGATCCC CGGGGAATTC AATCGATGGC CGCCATGGCC 2000
15 CAACTTGTTT ATTGCAGCTT ATAATGGTTA CAAATAAAGC AATAGCATCA 2050
CAAATTTTAC AAATAAAGCA TTTTTTTTAC TGCATTCTAG TTGTGGTTTG 2100
20 TCCAAACTCA TCAATGTATC TTATCATGTC TGGATCGATC GGAATTAAT 2150
TCGGCGCAGC ACCATGGCCT GAAATAACCT CTGAAAGAGG AACTTGGTTA 2200
GGTACCTTCT GAGGCGGAAA GAACCAGCTG TGGAATGTGT GTCAGTTAGG 2250
30 GTGTGGAAAG TCCCCAGGCT CCCCAGCAGG CAGAAGTATG CAAAGCATGC 2300
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35 GGCAGAAGTA TGCAAAGCAT GCATCTCAAT TAGTCAGCAA CCATAGTCCC 2400
GCCCCTAACT CCGCCCATCC CGCCCCTAAC TCCGCCCAGT TCCGCCCATT 2450
CTCCGCCCCA TGGCTGACTA ATTTTTTTTA TTTATGCAGA GGCCGAGGCC 2500
45 GCCTCGGCCT CTGAGCTATT CCAGAAGTAG TGAGGAGGCT TTTTGGAGG 2550
CCTAGGCTTT TGCAAAAAGC TAGCTTATCC GGCCGGAAC GGTGCATTGG 2600
50 AACGCGGATT CCCCCTGCCA AGAGTCAGGT AAGTACCGCC TATAGAGTCT 2650
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60 CACAGGTGTC CACTCCCAGG TCCAACGCA CCTCGGTTTCG CGAAGCTAGC 2800
TTGGGCTGCA TCGATTGAAT TCCACCATGG GATGGTCATG TATCATCCTT 2850
65 TTTCTAGTAG CAACTGCAAC TGGAGTACAT TCAGATATCC AGCTGACCCA 2900

GTCCCCGAGC TCCCTGTCCG CCTCTGTGGG CGATAGGGTC ACCATCACCT 2950

5 GCCGTGCCAG TCAGAGCGTC GATTACGATG GTGATAGCTA CATGAACTGG 3000

TATCAACAGA AACCAGGAAA AGCTCCGAAA CTACTGATTT ACGCGGCCTC 3050

10 GTACCTGGAG TCTGGAGTCC CTTCTCGCTT CTCTGGATCC GGTTCCTGGA 3100

CGGATTTAC TCTGACCATC AGCAGTCTGC AGCCGGAAGA CTTGCAACT 3150

15 TATTACTGTC AGCAAAGTCA CGAGGATCCG TACACATTTG GACAGGGTAC 3200

CAAGGTGGAG ATCAAACGAA CTGTGGCTGC ACCATCTGTC TTCATCTTCC 3250

20 CGCCATCTGA TGAGCAGTTG AAATCTGGAA CTGCCTCTGT TGTGTGCCTG 3300

CTGAATAACT TCTATCCCAG AGAGGCCAAA GTACAGTGGA AGGTGGATAA 3350

CGCCCTCCAA TCGGGTAACT CCCAGGAGAG TGTCACAGAG CAGGACAGCA 3400

30 AGGACAGCAC CTACAGCCTC AGCAGCACCC TGACGCTGAG CAAAGCAGAC 3450

TACGAGAAAC ACAAAGTCTA CGCCTGCGAA GTCACCCATC AGGGCCTGAG 3500

35 CTCGCCCCGTC ACAAAGAGCT TCAACAGGGG AGAGTGTTAA GCTTCGATGG 3550

CCGCCATGGC CCAACTTGTT TATTGCAGCT TATAATGGTT ACAAATAAAG 3600

CAATAGCATC ACAAATTTCA CAAATAAAGC ATTTTTTTTCA CTGCATTCTA 3650

45 GTTGTGGTTT GTCCAACTC ATCAATGTAT CTTATCATGT CTGGATCGAT 3700

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50 GAACTTGGTT AGGTACCTTC TGAGGCGGAA AGAACCAGCT GTGGAATGTG 3800

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5 GCCGTCGTTT TACAACGTCG TGA CTGGGAA AACCTGGCG TTACCCAACT 4200

10 TAATCGCCTT GCAGCACATC CCCCCTTCGC CAGCTGGCGT AATAGCGAAG 4250

AGGCCCGCAC CGATCGCCCT TCCCAACAGT TCGTAGCCT GAATGGCGAA 4300

15 TGGCGCCTGA TGCGGTATTT TCTCCTTACG CATCTGTGCG GTATTTTACA 4350

CCGCATACGT CAAAGCAACC ATAGTACGCG CCCTGTAGCG GCGCATTAAAG 4400

20 CGCGGCGGGT GTGGTGGTTA CGCGCAGCGT GACCGCTACA CTTGCCAGCG 4450

25 CCCTAGCGCC CGCTCCTTTC GCTTTCTTCC CTTCTTTCT CGCCACGTTT 4500

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50 CTGGGTCATG GCTGCGCCCC GACACCCGCC AACACCCGCT GACGCGCCCT 4950

55 GACGGGCTTG TCTGCTCCCG GCATCCGCTT ACAGACAAGC TGTGACCGTC 5000

TCCGGGAGCT GCATGTGTCA GAGGTTTTCA CCGTCATCAC CGAAACGCGC 5050

60 GAGGCAGTAT TCTTGAAGAC GAAAGGGCCT CGTGATACGC CTATTTTTTAT 5100

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65 CGGGGAAATG TCGCGGGAAC CCCTATTTGT TTATTTTTTCT AAATACATTC 5200

AAATATGTAT CCGCTCATGA GACAATAACC CTGATAAATG CTTCAATAAT 5250

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5 CCCTTTTTTG CGGCATTTTG CCTTCTGTT TTTGCTCACC CAGAAACGCT 5350

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15 GAACGTTTTTC CAATGATGAG CACTTTTAAA GTTCTGCTAT GTGGCGCGGT 5500

ATTATCCCGT GATGACGCCG GGCAAGAGCA ACTCGGTTCG CGCATACT 5550

20 ATTCTCAGAA TGAATTGGT GAGTACTCAC CAGTCACAGA AAAGCATCTT 5600

ACGGATGGCA TGACAGTAAG AGAATTATGC AGTGCTGCCA TAACCATGAG 5650

25 TGATAAAGT GCGGCCAAT TACTTCTGAC AACGATCGGA GGACCGAAGG 5700

30 AGCTAACCGC TTTTTTGCAC AACATGGGGG ATCATGTAAC TCGCCTTGAT 5750

CGTTGGGAAC CGGAGCTGAA TGAAGCCATA CCAAACGAG AGCGTGACAC 5800

35 CACGATGCCA GCAGCAATGG CAACAACGTT GCGCAAACTA TTAAGTGGCG 5850

AACTACTTAC TCTAGCTTCC CGGCAACAAT TAATAGACTG GATGGAGGCG 5900

40 GATAAAGTTG CAGGACCACT TCTGCGCTCG GCCCTTCCGG CTGGCTGGTT 5950

45 TATTGCTGAT AAATCTGGAG CCGGTGAGCG TGGGTCTCGC GGTATCATTG 6000

CAGCACTGGG GCCAGATGGT AAGCCCTCCC GTATCGTAGT TATCTACACG 6050

50 ACGGGGAGTC AGGCAACTAT GGATGAACGA AATAGACAGA TCGCTGAGAT 6100

AGGTGCCTCA CTGATTAAGC ATTGGTAACT GTCAGACCAA GTTTACTCAT 6150

ATATACTTTA GATTGATTTA AAATTCATT TTTAATTAA AAGGATCTAG 6200

60 GTGAAGATCC TTTTGTGATA TCTCATGACC AAAATCCCTT AACGTGAGTT 6250

TTCGTTCCAC TGAGCGTCAG ACCCGTAGA AAAGATCAAA GGATCTTCTT 6300

65 GAGATCCTTT TTTTCTGCGC GTAATCTGCT GCTTGCAAC AAAAAACCA 6350

CCGCTACCAG CGGTGGTTTG TTTGCCGGAT CAAGAGCTAC CAACTCTTTT 6400

5 TCCGAAGGTA ACTGGCTTCA GCAGAGCGCA GATACCAAAT ACTGTCCTTC 6450

TAGTGTAGCC GTAGTTAGGC CACCACTTCA AGAACTCTGT AGCACCGCCT 6500

10 ACATACCTCG CTCTGCTAAT CCTGTTACCA GTGGCTGCTG CCAGTGGCGA 6550

TAAGTCGTGT CTTACCGGGT TGGACTCAAG ACGATAGTTA CCGGATAAGG 6600

15 CGCAGCGGTC GGGCTGAACG GGGGGTTCGT GCACACAGCC CAGCTTGGAG 6650

CGAACGACCT ACACCGAACT GAGATACCTA CAGCGTGAGC ATTGAGAAAG 6700

20 CGCCACGCTT CCCGAAGGGA GAAAGGCGGA CAGGTATCCG GTAAGCGGCA 6750

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TATCTTTATA GTCCTGTCGG GTTTCGCCAC CTCTGACTTG AGCGTCGATT 6850

30 TTTGTGATGC TCGTCAGGGG GGCGGAGCCT ATGGAAAAAC GCCAGCAACG 6900

CGGCCTTTTT ACGGTTCCCTG GCCTTTTGCT GGCCTTTTGC TCACATGTTC 6950

35 TTTCTGCGT TATCCCTGA TTCTGTGGAT AACCGTATTA CCGCCTTTGA 7000

40 GTGAGCTGAT ACCGCTCGCC GCAGCCGAAC GACCGAGCGC AGCGAGTCAG 7050

TGAGCGAGGA AGCGGAAGAG CGCCAATAC GCAAACCGCC TCTCCCCGCG 7100

45 CGTTGGCCGA TTCATTAATC CAGCTGGCAC GACAGGTTTC CCGACTGGAA 7150

AGCGGGCAGT GAGCGCAACG CAATTAATGT GAGTTACCTC ACTCATTAGG 7200

50 CACCCAGGC TTTACACTTT ATGCTTCCGG CTCGTATGTT GTGTGGAATT 7250

55 GTGAGCGGAT AACAAATTCA CACAGGAAAC AGCTATGACC ATGATTACGA 7300

ATTAA 7305

60

CLAIMS

1. A DNA construct comprising a transcriptional initiation site, a transcriptional termination site, a selectable gene, a product gene
5 provided 3' to the selectable gene, a transcriptional regulatory region regulating transcription of both the selectable gene and the product gene, the selectable gene being positioned within an intron having a splice donor site 5' of the intron, which splice donor site regulates expression of the product gene using the transcriptional
10 regulatory region.
2. The DNA construct of claim 1 wherein the splice donor site comprises an efficient splice donor sequence.
- 15 3. The DNA construct of claim 2 wherein the splice donor site comprises a consensus splice donor sequence.
4. The DNA construct of claim 2 wherein the splice donor site comprises the sequence GACGTAAGT.
20
5. The DNA construct of claim 1 wherein the selectable gene is an amplifiable gene.
6. The DNA construct of claim 5 wherein the amplifiable gene is DHFR.
25
7. The DNA construct of claim 1 wherein the transcriptional regulatory region comprises a promoter and an enhancer.
8. A vector comprising the DNA construct of claim 1.
30
9. The vector of claim 8 wherein the selectable gene of the DNA construct is an amplifiable gene.
10. The vector of claim 8 that is capable of replication in a eukaryotic
35 host.
11. A eukaryotic host cell comprising the vector of claim 10.
12. A eukaryotic host cell comprising the DNA construct of claim 5.
40
13. The host cell of claim 11 wherein the vector is introduced into the host cell by electroporation.
14. A eukaryotic host cell comprising the DNA construct of claim 1
45 integrated into a chromosome of the host cell.

15. The host cell of claim 14 that is a mammalian cell.
16. A method for producing a product of interest comprising culturing the host cell of claim 11 so as to express the product gene and recovering the product from the host cell culture.
17. The method of claim 16 further comprising recovering the product from the culture medium.
18. The method of claim 16 wherein the selectable gene is an amplifiable gene and the splice donor site comprises an efficient splice donor sequence.
19. A method for producing a product of interest comprising culturing the host cell of claim 12 so as to express the product gene in a selective medium comprising an amplifying agent for sufficient time to allow amplification to occur, and recovering the product.
20. A method for producing eukaryotic cells having multiple copies of a product gene comprising transforming eukaryotic cells with the DNA construct of claim 5, growing the cells in a selective medium comprising an amplifying agent for a sufficient time for amplification to occur, and selecting cells having multiple copies of the product gene.
21. The method of claim 20 further comprising recovering from the selected cells the product of interest.

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FIG. 1A

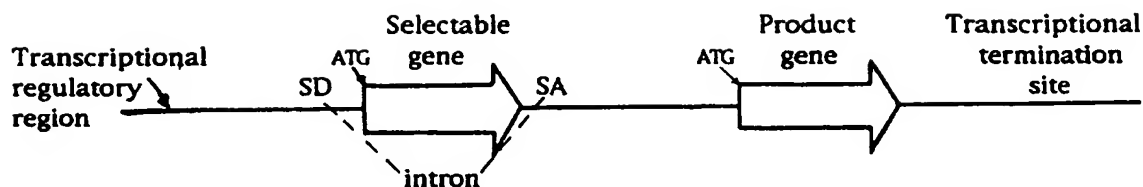


FIG. 1B

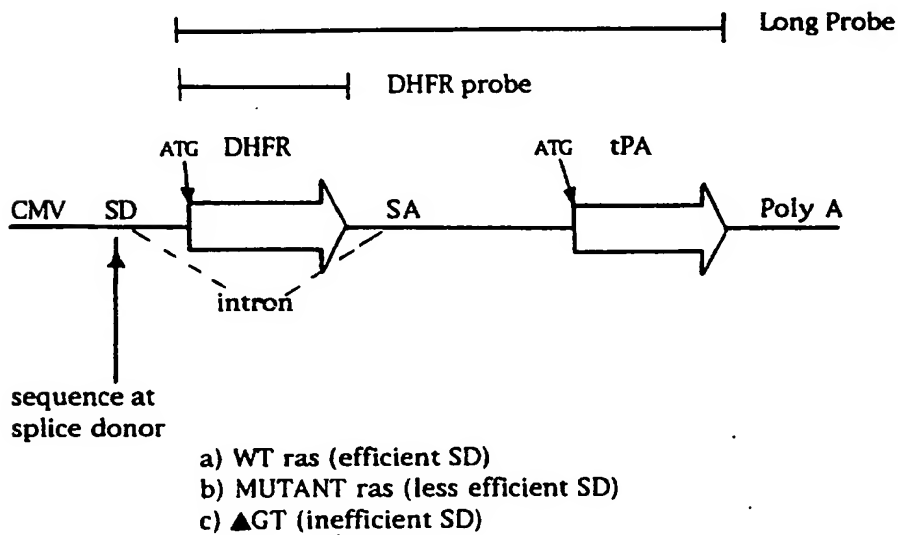
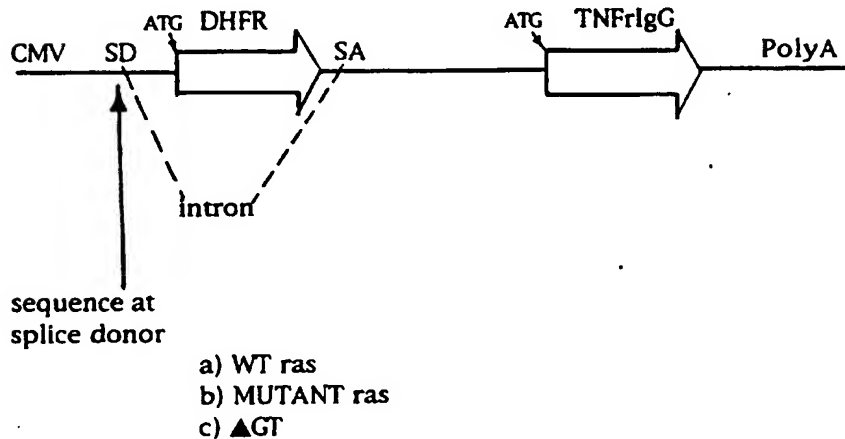


FIG. 1C



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FIG. 1D

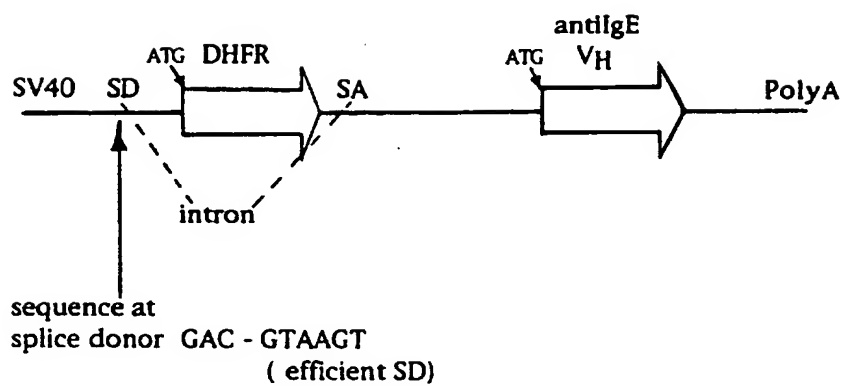
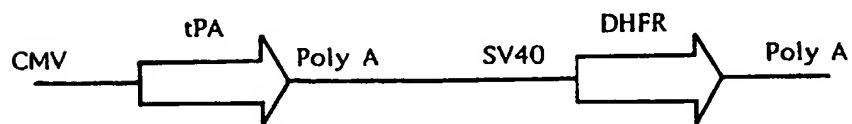


FIG. 2



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FIG. 3A

```

alul
sstI
sacI
hgiJII
hgiAI/aspHI
ecI136II
bspI286
bsiHKA1
bmyI
baniI
taqI
1 TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAT AGTAATCAAT TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC
AAGCTCGAGC GGGCTGTAC TAATAACTGA TCAATAATTA TCATTAGTTA ATGCCCCAGT AATCAAGTAT CGGGTATATA CCTCAAGGCG CAATGTATTG

          xmaI   tru9I
          maeI   mseI
          speI   aseI/asnI/vspI
          bslI
          aciI maeIII
          scrFI
          mvaI
          ecorII
          dsav
          aciI
          bglI bstNI
          sau96I
          haeIII/palI aciI
          asuI apyI(dcm+)
          maeII
          hinII/acyI
          ahaII/bsaHI
          aatII
          maeIII
101 TTACGGTAA TGGCCCGCCT GGCTGACCG CCAACGACCC CGGCCCATG ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA
AATGCCATT ACCGGCGGA CCGACTGGC GGTGCTGGG GCGGGGTAAC TGCAGTTATT ACTGCATACA AGGTATCAT TCGGTATTC CCTGAAAGGT

          maeII
          hinII/acyI
          ahaII/bsaHI
          aatII
          rsaI
          csp6I
          maeII
          nlaIII
          styI
          ncoI
          dsal hphI aciI
          bsaJI sfaNI
201 TTGACGTCAA TGGGTGGAGT ATTACGGTA AACTGCCCC TGGCAGTAC ATCAAGTGA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT
AACTGCAGT ACCCACCTCA TAAATGCCAT TTGACGGGTG AACCGTCATG TAGTTCACAT AGTATACGGT TCATCGGGGG GATAACTGCA GTTACTGCCA

          scrFI
          mvaI
          ecorII
          aciI
          bglI dsav
          sau96I bstNI
          haeIII/palI
          asuI apyI(dcm+)
          bsrI nlaIII
          rsaI
          csp6I
          maeII
          snaBI
          bsaAI
          csp6I
          rsaI
          maeII
          ncoI
          dsal hphI aciI
          bsaJI sfaNI
301 AAATGGCCG CCTGGCATT TGGCCAGTAC ATGACCTTAT GGGACTTTCC TACTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC
TTTACCGGCG GGACCGTAAT ACGGTCTATG ACGGTGGAATA CCTGGAAGG ATGAACGTC ATGATAGTGC ATAATCAGTA CGGATAATGG TACCACTACG

```

FIG. 3B

401 GGTTCGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA TTTCCAAGTC TCCACCCCAT TGCACGTCAAT GGGAGTTTGT TTTCGACCA
CCAAAACCGT CATGTAGTTA CCGGCACCTA TCGCCAACT GAGTGCCCT AAAGGTTTCA AGGTGGGTA ACTGCAGTTA CCTCAACA AAACCGTGGT

501 AAATCAACGG GACTTTCCAA AATGTCGTAA CAATCGGCC CCATGACGC AAATGGCGG TAGGCGTGA CGTGGGAGG TCTATATAAG CAGAGCTCGT
TTTAGTTGCC CTGAAAGGTT TTACAGCATT GTTACGCGG GGTAACTGG GTTACCCGCC ATCCGCACAT GCCACCCCTC AGATATATTC GTTCGAGCA

601 TTAGTGAACC GTCAGATCG CTGGAGACGC CATCCAGCT GTTTGACCT CCATAGAAGA CACCGGACC GGTATCTTCT GTGGCCCTGG CATAGTCGGA GGCSCGGCC CTGCCCAGT
AATCACTTGG CAGTCTAGCG GACCTCTGCG GTAGTGGCA CAAAACCTGGA GGTATCTTCT GTGGCCCTGG GTAGTGGCA GGCSCGGCC CTGCCCAGT

FIG. 3C

701 TTGGAACGCG GATTCCCGGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA GCGATAAGAG GATTTTATCC CATGTTTCGA CCATTGAACT
 AACCTTGGCG CTAAGGGGCA CGGTTCTCAC GACATTTCATG CCGGATATCT CGCTATTCTC CTAATAATAGG GCGACGGTA GTACCAAGCT GGTAACTTGA

801 GCATCGTCG CGTGTCGCCAA AATATGGGA TTGGCAAGAA CGGAGACCTA CCTGCCCTC CGCTCAGGAA CGGTTCAAG TACTTCCAA GAATGACCAC
 CGTAGCAGCG GCACAGGTT TTATACCCCT AACCGTTCTT GCCTCTGGAT GGGACGGGAG GCGAGTCCTT GCGCAAGTTC ATGAAGGTTT CTACTGGTG

901 AACCTCTTCA GTGGAAGGTA AACAGAACTT GGTGATTATG GTTAGGAAA CCTGGTTCTC CATTCCTGAG AAGAATCGAC CTTTAAAGGA CAGAATTAT
 TTGGAGAAGT CACCTTCCAT TTGTCTTAGA CCACTAATAC CCATCCTTTT GGACCAAGAG GTAAGGACTC TTCTTAGCTG GAAATTTCTT GTCTTAATTA

1001 ATAGTTCTCA GTAGAGAACT CAAAGAACCA CCACGAGGAG CTCATTTTCT TGCCAAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTGG
 TATCAAGAGT CATCTCTTGA GTTCTTGGT GGTGCTCCTC GAGTAAAGA ACGGTTTTCA AACCTACTAC GGAATTCTGA ATAACCTGTT GGCCTTAACC

tfil
 aciI
 thaI hinfI
 fnuDII/mvnI
 bstUI
 bsh1236I
 fnu4HI
 bbvI
 nspBII
 aciI
 nlaIII
 taqI
 rsaI
 csp6I
 scfI
 mnlI
 rsaI
 bstUI
 bsh1236I
 mluI
 bsrBI
 aciI
 xmnI
 rsaI
 csp6I
 scaI
 mnlI
 ddeI
 asp700
 bsmAI
 bsaI
 pflMI
 bslI
 sfaNI
 eco57I
 mboII
 earI/ksp632I
 mnlI
 tfil
 hinfI
 alwNI
 hphI
 scrFI
 mvaI
 ecoRII
 dsaV
 bstNI
 apyI[dcM+]
 sexAI
 mboII
 taqI
 mseI
 ahaIII/draI
 aseI/asnI/vspI
 tru9I
 mseI
 aluI
 sstI
 sacI
 hgiJII
 hgiAI/aspHI
 ecl136II
 bsp1286
 bsiHKAI
 bmyI
 banII
 bslI
 mnlI
 bstXI
 foki
 sfanI
 mseI
 afaIII/bfrI
 bsaWI
 mspI
 hpaII

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FIG. 3D

1101 CAAGTAAAGT AGACATGGTT TGGATAGTCG GAGGCAGTTC TGTTACCAG GAAGCCATGA ATCAACCAGG CCACCTTAGA CTCTTGTGA CAAGGATCAT
 GTTCATTCA TCTGTACCAA ACCTATCAGC CTCCTGTCAG ACAATGGTC CTTCGGTACT TAGTGGTCC GGTGGAATCT GAGAAACACT GTTCCTAGTA
 accI nlaIII mnlI apyI(dcm+) hinfI bstNI ddeI maeIII alwI(dam-) dpnII(dam-) mboI/ndeII(dam-) dpnI(dam+) sau3AI
 scrFI mvaI ecorII dsav tfiI nlaIII bstNI pleI haeIII/palI haeI nlaIII
 1201 GCAGGAATTT GAAAGTGACA CGTTTTTCCC AGAAATTGAT TTGGGGAAT ATAAACCTCT mnlI bsaJI hgaI ddeI apyI(dcm+) mnlI bstNI
 CGTCCTTAA CTTTCACTGT GCAAAAGGG TCTTTAACTA AACCCCTTTA TATTGGAGA GGTCTTTATG GGTCCGACG AGAGACTCCA GGTCTCTCTT
 maeII aflIII apyI(dcm+) mnlI bsaJI hgaI ddeI apyI(dcm+) mnlI bstNI
 1301 AAAGGCATCA AGTATAAGTT TGAAGTCTAC GAGAAGAAAG ACTAACAGGA AGATGCTTTC AAGTCTCTCG CTCCTCTCTT AAAGCTATGC ATTTTATATA
 TTTCCGTAGT TCATATTCAA ACTTCAGATG CTCTTCTTTC TGATTGCTCT TCTACGAAAG TTCAGAGAC GAGGGGAGGA TTTCGATACG TAAAAATATT
 sfaNI accI mboII mnlI aluI ppulOI
 1401 GACCATGGGA CTTTGTCTGG CTTAGACCC CTTGGCTTC GTTAGAACGC GGCTACAAAT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA
 CTGGTACCCT GAAAACGACC GAAATCTGGG GGAACCGAAG CAATCTTGG CCGATGTAA TTATGTATTG GAATACATAG TATGTATC TAAATCCACT
 nlaIII styI ncoI dsal bsaJI maeIII hphI
 fnu4HI aciI thalI fnuDII/mvnI tru9I bstUI mseI bsh1236I aseI/asnI/vspI
 maeIII hphI

[illegible]

FIG. 3F

bspMI sau96I haeIII/pali
 nlaIV haeIII/pali
 hgiCI asuI rsal
 bsaI ecoO109I/draII
 bsp1286 alwNI csp6I ddeI
 bmyI bsrI bcgI
 1901 GGGGGCACCT GCCAGCAGGC CCTGTACTTC TCAGATTTCG CCCGAAGGA TTGCTGGGA AGTCTGTGA AATAGATACC AGGCCACGT
 CCCCCGTGA CGGTCTGTCG GGACATGAAG AGTCTAAGC ACACGTAC GGGCTTCCT AACGACCT TCACGACACT TTATCTATGG TCCCGGTGCA

 scrFI hhaI/cfoI
 mvaI nlaIV hgiAI/aspHI
 ecorII nari bsp1286
 dsav kasI bsiHKA
 bstNI hinII/acyI
 bsaJI bmyI
 sau96I hgiCI bmyI
 avaII haeII apaII/snoI
 asuI sfanI scfI bsp1286 bmyI aciI banI alw44I/snoI fnu4HI asuI bslI
 mnli apyI[dcM+] alul bmyI maeII nspBII ahaiI/bsaHI bsrI bbvI bsh1236I bslI
 2001 GCTACGAGGA CCAGGGCATC AGCTACAGG GCACGTGGAG CACAGCGAG AGTGCGCCG AGTGACCAA CTGGAACAGC AGCGGTGG CCCAGAAGCC
 CGATGCTCTT GGTCCCGTAG TCGATGTCCC CGTGACCTC GTGTCCCTC TCACCGCGC TCACGTGGT GACCTGTG TCGGCAACC GGGTCTTCG

 scrFI pleI bsmaI scrFI
 mvaI taqI[dam-] mvaI
 ecorII sau3AI hinfi ecorII
 dsav mboI/ndeII[dam-] bstNI
 bstNI dpmI[dam+] apyI[dcM+] tru9I
 bsaJI dpmII[dam-] bsaJI
 2101 CTACAGCGG CGGAGGCCAG ACGCCATCAG GCTGGGCTG GGAACACA ACTACTGCAG AACCCAGAT CGAGACTCAA AGCCCTGGTG CTAGTCTTT
 GATGTGCCC GCCTCCGGT TCGGTAGT GATGACGTC TTGGGTCTA GCTCTGAGT TCGGACCAAC GATGCAGAA

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FIG. 3G

fnu4HI
bbvI
scfI
rsal
csp6I ddeI
aciI AAGCGGGGA AGTACAGTC AGAGTTCTGC AGCACCCCTG CCTGTCTGA GGGAAACAGT GACTGCTACT TTGGGAATGG GTCAGCTAC CGTGCAACGC
TTCCGCCCTC TCATGTCGAG TCTCAAGACG TCGTGGGAC GGACGAGACT CCCTTTGTCA CTGACGATGA AACCTTACC CAGTCGGATG GCACCGTGCG
scrFI
pfIMI
mvaI
ecorII
dsav
bstNI
bslI
apyI[dcm+] haeIII/pali
bsp1286 sau96I
bmyI alwNI asuI
bsrI bsaJI bsrI
2301 ACAGCCTCAC CGAGTCGGGT GCCTCCTGCC TCCCCTGGAA TTCATGATC CTGATAGGCA AGGTTTACAC AGCACAGAAC CCCAGTGCCC AGGCACTGGG
TGTCGGAGTG GCTCAGCCCC CGGAGGACCG AGGGCACTT AAGGTACTAG GACTATCCGT TCCAATGTC TCGTGTCTTG GGGTCACGGG TCCGTGACCC
nlaIV
hgiCI
banI maeII
scrFI
mval pmlI
ecorII
dsav
bstNI
pfIMI
eco72I
bsaAI
tflI hinfi
mspi
hpaII
apyl[dcm+]
CATAATTACT GCCGGATCC TGATGGGAT GCCAAGCCCT GTGTCCACCT GCTGAAGAAC CGCAGGCTGA CGTGGGAGTA CTGTGATGTG
2401 CGACCCGTTT GTATTAATGA CGGCCTTAGG ACTACCCCTA CGGTTCGGGA CCACGGTGCA CGACTTCTTG GCGTCCGACT GCACCTCAT GACACTACAC
scrFI
mval
ecorII
dsav
bstNI
apyI[dcm+]
fnu4HI bsmAI
aciI ddeI rsal
bspMI haeIII/pali csp6I
mnII CCCTCCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCG CATCAAAGGA GGGCTCTTCG CCGACATCGC CTCACACCCC TGGCAGGCTG
GGGAGGACGA GGTGGACGCC GGACTCTGTC ATGTGGGTG GAGTCAAAGC GTAGTTTCT CCCGAGAAGC GGCTGTAGCG GAGGTGGG ACCGTCCGAC
mnII
bslI bsaJI fnu4HI
bslI bslI bbvI

FIG. 3

[illegible]

FIG. 31

FIG. 3I

2901

3001

3101

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FIG. 3J

[illegible]

SUBSTITUTE SHEET (RULE 26)

[illegible]

[illegible]

[illegible]

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FIG. 30

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nlaIV
aciI
thaI
fnuDII/mvnI
bstUI
bsh1236I
hinPI
hhaI/cfoI
5201 CTTTTCGGG AAATGTGGC GGAACCCCTA TTTGTTTATT TTTCTAATA CATTCAATA TGTATCGCT CATGACAA TAACCTGTAT AAATGCTTCA
GAAAGCCCC TTACACGCG CTTGGGGAT AAACAAATAA AAAGATTAT GTAGTTTAT ACATAGCGA GTACTCTGTT ATTGGGACTA TTACGAAGT

rcaI
bspHI
bsrBI bsmAI
aciI nlaIII
fnu4HI
mboII
sspI earI/ksp632I
5301 ATAATATTGA AAAAGGAAGA GTATGAGTAT TCAACATTTC CGTGTGCGC TTATTCCCTT TTTTGGGCA TTTTGCTTC CTGTTTTC TCACCCAGAA
TATTATAACT TTTTCTTCT CATACTCATA AGTTGTAAAG GCACAGCGG AATAAGGGA AAAACGCCGT AAACCGAAG GACAAAACG AGTGGGTCTT

hgiAI/aspHI
bsp1286
sau3AI bsiHKAI
mboI/ndeII(dam-)
dpmI(dam+) bmyI
dpmII(dam-)
eco57I apaII/snoI
hphI sfaNI mboII(dam-) alw4I/snoI maeIII taqI alwI(dam-) aciI bstYI/xhoII
5401 ACGCTGGTGA AAGTAAAGA TGCTGAAGAT CAGTTGGTG CACGAGTGG TTACATCGAA CTGGATTCA ACAGCGGTAA GATCCTTGAG AGTTTTCGCC
TGGACCACT TTCATTTTCT ACGACTTCTA GTCAACCCAC GTGCTCACCC AATGTAGCTT GACCTAGAGT TGTGCGCATT CTAGGAACTC TCAAAAGCGG

scrFI
nciI
mspI
hpaII
dsav
hinII/acyI
hgaI caulI
ahaII/bsaHI
bcgI mcrI fnu4HI
5501 CCGAAGAAGC TTTTCCAATG ATGAGCACTT TTAAGTTCT GCTATGTGC GCGGTATTAT CCGGTATGA CGCGGGCAA GAGCAACTCG GTGCGCGCAT
GGCTTCTTGC AAAAGGTTAC TACTCGTGAA AATTCAAGA CGATACACC CGCCATAATA GGGCACTACT GCGGCCGTT CTCGTTGAGC CAGCGCGGTA

rsal
csp6I bsrI
scaI hphI maeIII sfaNI foki nlaIII
5601 ACACATTCT CAGAATGACT TGGTTGAGTA CTCACCACTC ACAGAAAAGC ATCTTACGGA TGGCATGACA GTAAGAGAAT TATGAGTGC TGCCATAACC
TGTGATAAGA GTCTTACTGA ACCAACTCAT GAGTGGTTCAG TGCTTTTTCG TAGAATGCTT ACCGACTGCT CATCTCTTCA ATACGTCACG ACGGTATTGG
nlaIII

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FIG. 3P

```

sau96I      nlaIII
avaII
sau3AI asuI      nlaIII
mboI/ndeII{dam-} sau3AI maeIII
dpnI{dam+}      mboI/ndeII{dam-}
dpnII{dam-}     dpnI{dam+}
pvuI/bspCI     dpnII{dam-}
mcrI mnlI      nlaIII aluI aciI
CTGACAACTGATA ACAGTCGGC CAACCTACTT CTGACAACGA TCGGAGGACC GAAGAGCTA ACCGCTTTT TGCACAACAT GGGGATCAT GTAACTCGCC
TACTCACTAT TGTGACGCCG GTTGAATGAA GACTGTGCT AGCCTCTCGG CTTCCTCGAT TGGCGAAAAA ACGTCTTGT CCCCCTAGTA CATTGAGCGG

5701

haeIII/palI
eaeI
cfrI
fnu4HI
aciI
sau3AI nlaIV
mboI/ndeII{dam-} aluI
dpnI{dam+} hpaII
dpnII{dam-} bsaII
TTGATCGTTG GGAACCGGAG CTGAATGAAG CCATACCAAA CGACGAGCGT GACACCACGA TGCACGACG AATGGCAACA ACGTTGCGCA AACTATTAAAC
AACTAGCAAC CCTTGGCCTC GACTTACTTC GGTAATGCTT GCTGTGCGCA CTGTGTGCT ACGTCTGCTG TACCGTGTGT TGCACGCGT TTGATAATTG

5801

mspI      hpaII
scrFI
aluI nciI      foki
rmaI dsav      mseI bsrI      fnu4HI
maeI cauI      aseI/asnI/vspI mnlI      fnu4HI bsvI      maeII      psp1406I      bsrI
CTTACTCTAG CTTCCCGGCA ACAATTATA GACTGGATGG AGCGGATAA AGTTGCAGGA CCACCTTCTGC GCTCGGCCCT TCCGGCTGGC
ACCGCTTGAT GAATGAGATG GAAGGCGCGT TGTTAATTAT CTGACCTACC TCCGCTATT TCAACGCTCT GGTGAAGACG CGAGCCGGA AGGCCGACCG

5901

mspI      hpaII      fnuDII/mvni      haeIII/palI
cfr10I      bstUI      sau96I
nlaIV hphI      bsmAI aciI      fnu4HI nlaIV
gsuI/bpmI      bsaI bsh1236I      bsvI bsrI asuI      mnlI
CTGATAAATC TGGAGCCGGT GAGCGTGGT CTCGCGGTAT CATTGCAGCA CTGGGCCAG ATGTAAGCC CTCCGTATC GTAGTTATCT
ACCAATAAC GACTATTAG ACCTCGGCA CTCGCACCCA GAGCGCCATA GTAACGTCGT GACCCCGTC TACCATTGG GAGGSCATAG CATCAATAGA

6001

ddeI
sau3AI nlaIV
mboI/ndeII{dam-}
dpnI{dam+} hgiCI      tru9I
dpnII{dam-}      banI mnlI      mseI      maeIII
eaml105I      foki
ACTATGGATG AACGAAATAG ACAGATCGCT GAGATAGTG CCTCACTGAT TAAGCATTTG TAACGTCTAG ACCAAGTTTA
6101 ACACGACGGG GAGTCAGGCA
TGTGCTGCC CTCAGTCCGT TGATACCTAC TTGCTTTATC TGTCTAGCA CTCTATCCAC GGAGTGACTA ATTCGTAACC ATTGACAGTC TGGTTCAAAT

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[illegible]

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FIG. 3R

6601 GCGGATAAGT CGTGCTTTAC CGGGTTGGAC TCAAGACGAT AGTTACCGGA TAAGCGCGC GGTGCGGCT GAACGGGGG TTCTGTCACA CAGCCAGCT
 CCGCTATTCA GCACAGAATG GCCCAACCTG AGTTCGTCTA TCAATGGCT ATTCGGCTC GCCAGCCGA CTTCGCCCCC AAGCAGGTGT GTCCGGTCCA
 6701 TGGAGCGMAC GACCTACACC AACCTGAGT ACCTACAGCG TGAGCATTGA GAAAGCGCCA CGCTTCCGA AGGAGAAAG CGGACAGGT ATCCGGTAAG
 ACCTCGCTTG CTGGATGTGG CTGACTCTA TGGATGTCG ACTCGTAAC TTTTCGGCT GCGAAGGCT TCCCTCTTC CGCTGTCCA TAGGCCATTC
 6801 CGGCAGGGT GGAACAGGAG AGCGACGAG GGAGCTTCCA GGGGAAACG CTTGATATCT TTATAGTCT GTCCGGTTTC GCCACCTCTG ACTTGAGCT
 GCCGTCCAG CCTGTCTC TCGGTGCTC CTTGAAGGT CCCCCTTTC GGACCATAGA AATATCAGA CAGCCCAAAG CGGTGGAGAC TGAATCGCA
 6901 CGATTTTGT GATGCTCGT AGGGGGCGG AGCCTATGGA AAAACGCCAG CAACGGGCGC TTTTACGCT TCCTGGCCTT TTGCTGGCCT TTGCTCACA
 GCTAAACA CTACGAGCAG TCCCCCGCC TCGGATACCT TTTGCGGTC GTTGGCGG AAAAATGCCA AGGACCGGA AACGAGGT
 7001 TGTCTTTCC TCGGTTATCC CTTGATTCTG TGGATAACCG TATTACGCC CTTGAGTGAG CTGATACCG TCCTGGCGC CGAAGCAGC AGCGAGCA
 ACAAGAAAG ACGCATAGG ACCTATTGGC ATAATGGCG ATAATGGCG GACTATGCG AGCGCGTGC GCTTGTGCTG TCGGTGCT

Restriction Enzymes and Sites:
 6601: *scriFI*, *ncii*, *mspi*, *hpaII*, *dsav*, *cauII*, *pleI*, *hinfI*, *mspi*, *hpaII*, *bsaWI*, *maeIII*, *hinPI*, *mcrl*, *hhaI/cfoI*, *bbvI*, *fnu4HI*, *nspBII*, *aciI*, *hgiAI/aspHI*, *bsp1286*, *bsiHKA1*, *bmyI*, *apaLI/snoI*, *alw44I/snoI*, *alul*
 6701: *scriFI*, *mvalI*, *ecorII*, *dsav*, *bstNI*, *bsaJI*, *alul*, *apyI[dcM+]*, *hhaI/cfoI*, *hinPI*, *mnlI*, *bsaJI*, *dsav*, *ecorII*, *mvalI*, *scriFI*, *hpaII*, *hhaI/cfoI*, *hinPI*, *mspi*, *hpaII*, *bsaWI*, *maeIII*, *hinPI*, *mcrl*, *hhaI/cfoI*, *bbvI*, *fnu4HI*, *nspBII*, *aciI*, *hgiAI/aspHI*, *bsp1286*, *bsiHKA1*, *bmyI*, *apaLI/snoI*, *alw44I/snoI*, *alul*
 6801: *scriFI*, *mvalI*, *ecorII*, *dsav*, *bstNI*, *bsaJI*, *alul*, *apyI[dcM+]*, *hhaI/cfoI*, *hinPI*, *mnlI*, *bsaJI*, *dsav*, *ecorII*, *mvalI*, *scriFI*, *hpaII*, *hhaI/cfoI*, *hinPI*, *mspi*, *hpaII*, *bsaWI*, *maeIII*, *hinPI*, *mcrl*, *hhaI/cfoI*, *bbvI*, *fnu4HI*, *nspBII*, *aciI*, *hgiAI/aspHI*, *bsp1286*, *bsiHKA1*, *bmyI*, *apaLI/snoI*, *alw44I/snoI*, *alul*
 6901: *scriFI*, *mvalI*, *ecorII*, *dsav*, *bstNI*, *bsaJI*, *alul*, *apyI[dcM+]*, *hhaI/cfoI*, *hinPI*, *mnlI*, *bsaJI*, *dsav*, *ecorII*, *mvalI*, *scriFI*, *hpaII*, *hhaI/cfoI*, *hinPI*, *mspi*, *hpaII*, *bsaWI*, *maeIII*, *hinPI*, *mcrl*, *hhaI/cfoI*, *bbvI*, *fnu4HI*, *nspBII*, *aciI*, *hgiAI/aspHI*, *bsp1286*, *bsiHKA1*, *bmyI*, *apaLI/snoI*, *alw44I/snoI*, *alul*
 7001: *scriFI*, *mvalI*, *ecorII*, *dsav*, *bstNI*, *bsaJI*, *alul*, *apyI[dcM+]*, *hhaI/cfoI*, *hinPI*, *mnlI*, *bsaJI*, *dsav*, *ecorII*, *mvalI*, *scriFI*, *hpaII*, *hhaI/cfoI*, *hinPI*, *mspi*, *hpaII*, *bsaWI*, *maeIII*, *hinPI*, *mcrl*, *hhaI/cfoI*, *bbvI*, *fnu4HI*, *nspBII*, *aciI*, *hgiAI/aspHI*, *bsp1286*, *bsiHKA1*, *bmyI*, *apaLI/snoI*, *alw44I/snoI*, *alul*

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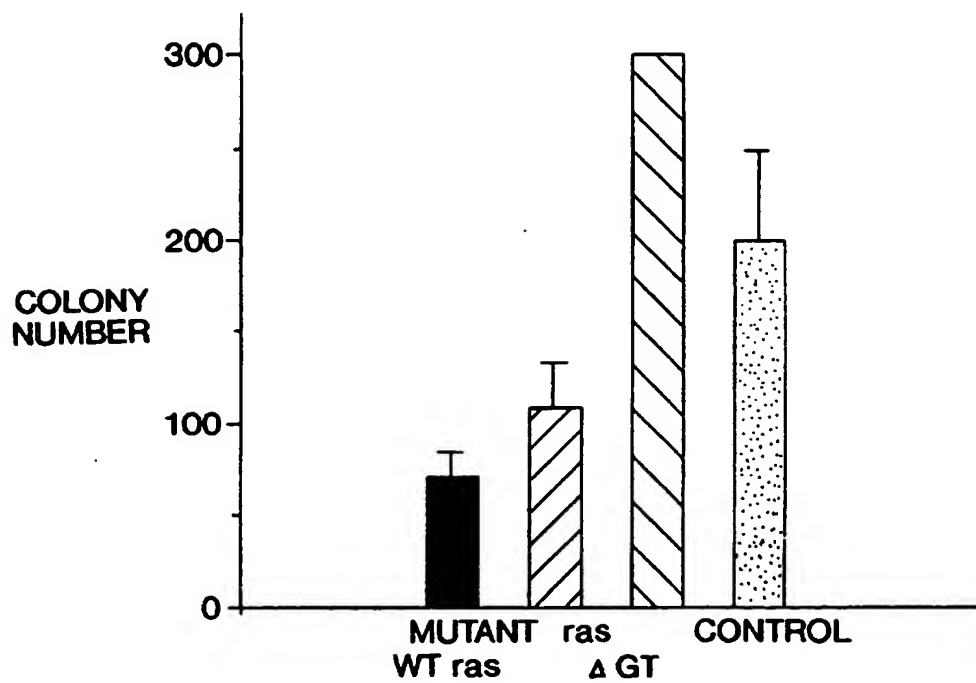


FIG. 4

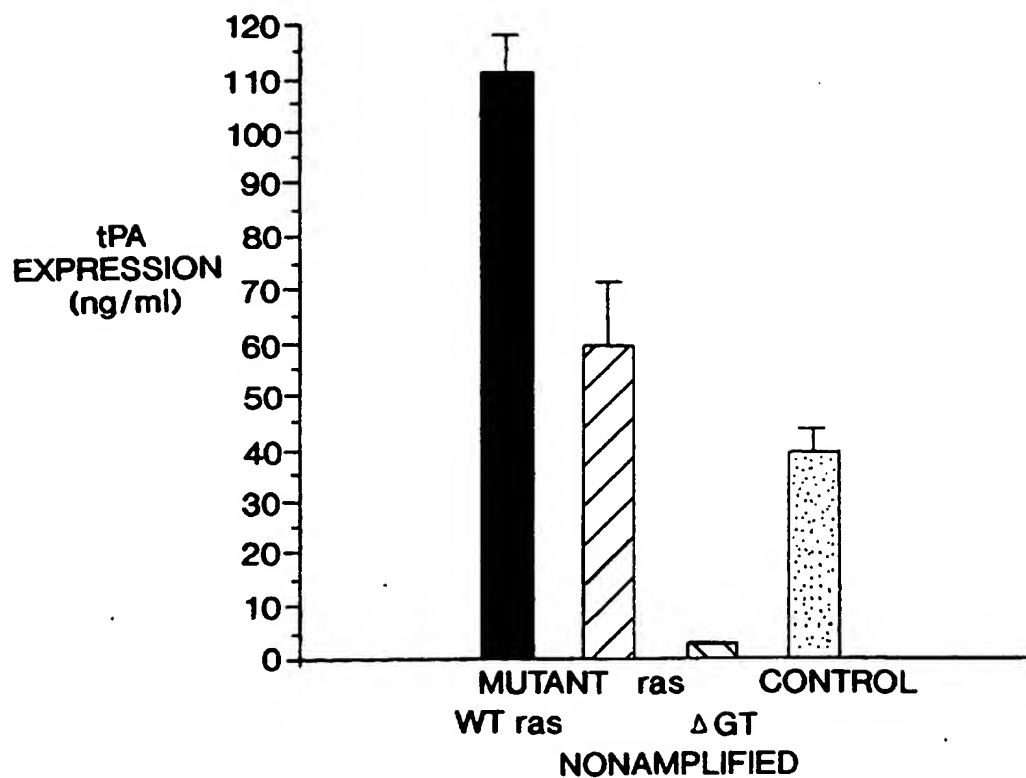


FIG. 5A

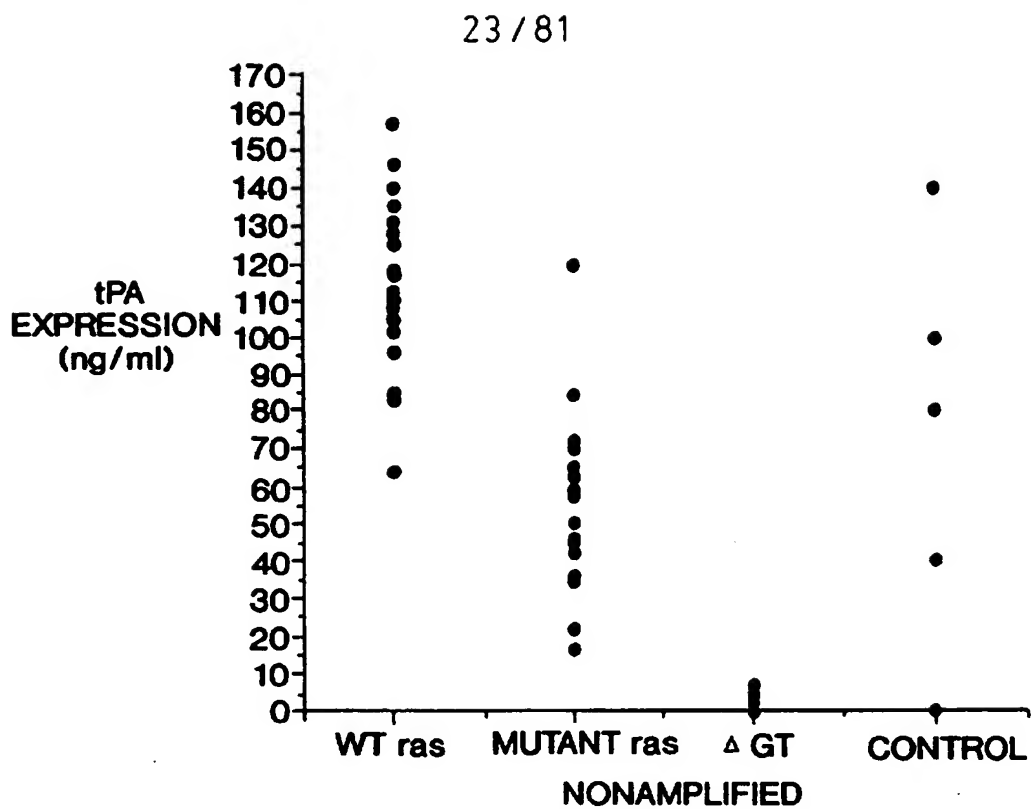


FIG. 5B

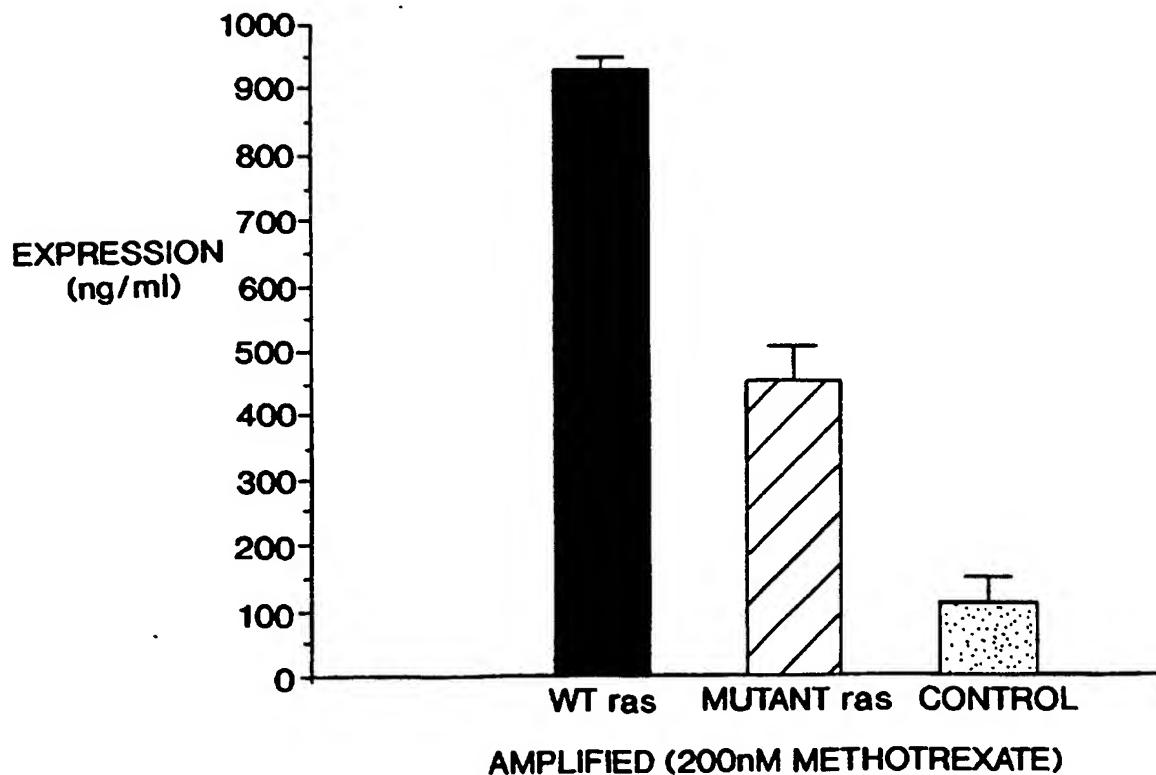


FIG. 5C

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FIG. 6A

```

alul
sstI
sacI
hgiJII
hgiAI/asphi
ecll36II
bsp1286
bsiHKA1
bmyI
banII
taqI
1 TTCGAGCTCG CCGGACATG ATTATTGACT AGTTATTAAT AGTAATCAAT TACGGGGTCA TTAGTTTCATA GCCCATATAT GGAGTTCGC GTTACATAAC
AAGCTCGAGC GGGCTGTAAC TAATAACTGA TCAATAATTA TCATTAGTTA ATGCCCCAGT AATCAAGTAT CGGGTATATA CCTCAAGGCG CAATGTATTG
thaI
fnuDII/mvnI
bstUI
bsh1236I
aciI maeIII
bslI
rmaI tru9I
maeI mseI
speI aseI/asnI/vspI
maeII
hinII/acyI
ahaII/bsaHI
aciI
aciI apyI[dcM+]
haeIII/palI
sau96I
101 TTACGGTAA TGGCCCGCT GGCTGACCG CCAAGGACCC CGGCCCATG ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCTA
AATGCCATT ACCGGGCGA CCGACTGGC GGTTCCTGG GCGGGTAACT TGCAGTTATT ACTGCATACA AGGGTATCAT TCGGGTTATC CCTGAAAGGT
maeII
hinII/acyI
ahaII/bsaHI
aatII
maeII
rsal
rsal
ndeI
csp6I
201 TTGACGTCAA TGGGTGGAGT ATTTACGTA AACTGCCAC TTGGCAGTAC ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT
AAGTGCAGTT ACCACCTCA TAAATGCCAT TTGACGGGTG AACCGTCATG TAGTTCACAT AGTATACGAT TCATGCGGGG GATTAAGTCA GTTACTGCCA
maeII
scrFI
mvaI
ecorII
dsav
aciI
bglI bstNI
sau96I
haeIII/palI
aciI
aciI
bglI
rsal
csp6I
301 AAATGGCCCG CCTGGCATT TGCCCGATT TGCACGATAC ATGACCTTAT GGGACTTTCC TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC
TTTACCGGGC GGACCGTAAT ACGGTTCATG TACTTGAATA CCTGAAAGG ATGAACCGTC ATGTAGATGC ATAATCAGTA GCGATAATGG TACCACTACG
nlaIII
styI
ncol
dsal hphI aciI
bsaJI sfaNI

```

FIG. 6B

[illegible]

FIG. 6C

tfiI
 aciI
 thaI hinfI
 fnuDII/mvnI
 bstUI
 bsh1236I
 701 TTGMAAGCG GATTCGCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA GCGATAAGAG GATTTTATCC CCGCTGCCAT CATGTTTGA CCATTGAAC
 AACCTTGGC CTAAGGGCA CGGTCTCAC GACATTCTAC GCGATATCT CCGTATTCTC CTAAATAGG GCGACGGTA GTACCAAGCT GGTAACTTGA
 fnu4HI
 bbvI
 nspBII
 aciI
 nlaIII
 taqI
 thaI
 fnuDII/mvnI
 bstUI
 bsh1236I
 mluI
 barBI
 aflIII
 rsaI
 aciI
 xmnI
 csp6I
 mnlI
 ddel asp700
 scaI
 801 GCATCGTGC CGTGTCCCA AATATGGGA TTGGCAAGAA CGGAGACCTA CCTGCCCCC CGCTCAGGAA CGCGTTCAAG TACTTCCAA GAATGACCAC
 CGTAGCAGC GCACAGGGT TTATACCCCT AACCGTTCTT GCCTCTGGAT GCGACGGAG GCGAGTCTT GCGCAAGTTC ATGAAGGTTT CTACTGGTG
 pflMI
 bslI
 sfaNI
 eco57I
 mboII
 earI/ksp632I
 mnlI
 901 AACCTCTTCA GTGGAAGGA AACAGAATCT GGTGATTATG GGTAGGAAGA CCTGGTTCTC CATTCCTGAG AAGAATCGAC CTTTAAAGGA CAGAATTAAT
 TTGGAGAAGT CACCTTCCAT TTGTCTTAGA CCACTAATAC CCATCCTTTT GGACCAAGAG GTAAGGACTC TTCTTAGCTG GAATTTCTT CTCTTAATTA
 tfiI
 hinfI
 alwNI
 hphI
 mboII
 taqI
 mseI
 tru9I
 mseI
 ahaIII/draI
 aseI/asnI/vspI
 aluI
 sstI
 sacI
 hgiIII
 hgiAI/asphi
 ecl136II
 bsp1286
 bsiHKA
 bmyI
 banII
 mnlI
 bslI
 bstXI
 foki
 sfanI
 mseI
 afluI/bfrI
 bsaWI
 mspI
 hpaII
 1001 ATAGTTCTCA GTAGAGAAGT CAAAGAACA CCACGAGGAG CTCATTTTCT TGCCAAAAGT TTGGATGATG CCTTAAGACT TATGAACAA CCGGAATTGG
 TATCAAGAGT CATCTCTTGA GTTCTTGGT GGTGCTCCTC GAGTAAAGA ACGTCTTCA AACCTACTAC AACCTACTAC GGAATTTCTG ATACTTGT GGCCTTAACC

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FIG. 6D

```

accI nlaIII          haeIII/palI
scrFI          scrFI          haeI
mvaI          mvaI          mboI/ndeII(dam-)
ecoRII          ecoRII          dpnI(dam+)
dsav          tfiI          pleI          dpnII(dam-)
bstNI          nlaIII          bstNI          ddeI          hinfI          maeIII          alwI(dam-)
apyl(dcm+)          hinfI          apyl(dcm+)          hinfI          maeIII          alwI(dam-)
TGTATTACCAAG GAGGCAATTC TGGATAGTCG GAGGCAATTC ATCAACCATGA CCACCTTAGA CTCTTGTA CAAGGATCAT
GTTCCATTCA TCTGTACCAA ACCTATCAGC CTCGTCAGC ACAAATGGTC CTTCGTACT TAGTTGGTCC GGTGGAATCT GAGAAACACT GTTCCTAGTA

1101 CAAGTAAAGT AGACATGGTT TGGATAGTCG GAGGCAATTC TGGGGAAT ATAACTCTC CCCAGAATAC CCAGGCGTCC TCTCTGAGGT CCAGGAGGAA
CGTCCCTTAA CTTTCACTGT GCAAAAAGGG TCTTTAACTA AACCCCTTA TATTGGAGA GGGTCTTATG GGTCCGAGG AGAGACTCCA GGTCTCTCCTT

apoI          maeII          mnlI          hnlI/acyI          scrFI          hnlI          scrFI
          aflIII          ahaII/bsaHI          mvaI          mvaI          ecorII          dsav          bstNI          bslI          asuI          mnlI
          maeIII          scrFI          mvaI          ecoNI          sau96I          dsav          apyl(dcm+)          mnlI          bstNI
          apyl(dcm+)          mnlI          bsaJI          hgaI          ddeI          apyl(dcm+)
1201 GCAGGAATTT GAAAGTGACA CGTTTTTCCC AGAATTGAT TTGGGGAAT ATAACTCTC CCCAGAATAC CCAGGCGTCC TCTCTGAGGT CCAGGAGGAA
CGTCCCTTAA CTTTCACTGT GCAAAAAGGG TCTTTAACTA AACCCCTTA TATTGGAGA GGGTCTTATG GGTCCGAGG AGAGACTCCA GGTCTCTCCTT

sfaNI          accI          mboII          mboII          mnlI          aluI          ppulOI          nsII/avaIII
1301 AAAGCATCA ACTATAAGTT TGAAGTCTAC GAGAAGAAG ACTAACAGGA AGATGCTTTC AAGTTCTCTG CTCCCCTCCT AAAGCTATGC ATTTTATAA
TTTCCGTAGT TCATATTCAA ACTTCAGATG CTCTTCTTTC TGATTGTCCT TCTACGAAAG TTCAAGAGAC GAGGGGAGGA TTTCGATACG TAAAAATATT

nlaIII          styI          ncoI          dsaI          bsaJI          styI          bsaJI          styI          mnlI          hphI          maeIII
          fnu4HI          aciI          thalI          fnuDII/mvnlI          tru9I          bstUI          msel          bsh1236I          aseI/asnI/vspI
1401 GACCATGGGA CTTTGTCTGG CTTTAGACCC CCTTGGCTTC GTTAGAACGC GGCTACAATT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA
CTGTACCCT GAAAACGACC GAAATCTGGG GGAACCGAAG CAATCTTGG CCGATGTAA TTATGTATTG GAATACATAG TATGTATC TAAATCCACT

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FIG. 6F

```

scrFI      nciI      mspI      hpaII      dsav      cauII      xmaI/pspAI
smal      scrFI      nciI      dsav      cauII      bslI      sau96I      haeIII/palI
asuI      scrFI      mvaI bsaJI      ecorII      dsav      bstNI bsaJI      bslI avai      apyI[dcn+]
mval bsaJI      ecorII      dsav      bstNI bsaJI      bslI avai      apyI[dcn+]      rsal      csp6I      bsp1407I
1801 CCTACTTGTA CAATGACTGT CCAGGCCCGG GGCAGGATAC GGACTGCAGG GAGTCGAGA GCGGCTCCTT CACCGCTTCA GAAAACCACC TCAGACACTG
GGATGAACAT GTTACTGACA GGTCGGGGCC CCGTCTATG CCTGACGTCC CTCACACTCT CCCCAGGAGAA GTGGCGAAGT CTTTGGTGG AGTCTGTGAC

nlaIV      fnu4HI      aciI      hphI      eco57I      mnlI      alwNI      ddel
scrFI      nciI      mspI      hpaII      dsav      cauII      sau96I      aval      asuI      draIII      bbvI      mboII      bsrI cfr10I
1901 CCTCAGCTGC TCCAAATGCC GAAAGGAAAT GGTTCAGGTG GAGATCTCTT CTTCACACAGT GGACCGGGAC ACCGTGTGTG GCTGCAGGAA GAACCACTAC
GGAGTCGACG AGGTTTACG CTTTCTTTTA CCCAGTCCAC CTCTAGAGAA GAACGTGTCA CCTGGCCCTG TGGCACACAC CGACGTCCCTT CTTGGTCTAG

```

[illegible]

2101 GCACCTGCCA TGCAGGTTTC TTTCTAAGAG AAAACGAGTG TGTCCTCTGT AGTAACTGTA AGAAAAGCCT GGAGTGCACG AAGTTGTGCC TACCCACAGAT
CGTGGACGCT ACGTCCAAAG AAGATTCTC TTTTGCTCAC ACAGAGGACA TCATTGACAT TCCTTTTCGGA CTCACGTGC TTCAACACGG ATGGGGTCTA

[illegible]

rsal
csp61
bspl4071 bsll
TTGT ACACCCTGCC
CACA TGTGGGACGG

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[illegible]

FIG. 6L

[illegible]

[illegible]

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[illegible]

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FIG. 6P

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maeIII          rmaI      sau3AI
sau3AI hphI mboI/ndeII(dam-)
mboI/ndeII(dam-)
dpmI(dam+) dpmI(dam-)
dpmII(dam-) dpmII(dam-)
          ahaIII/draI maeI      alwI(dam-)
tru9I      tru9I bstYI/xhoII bstYI/xhoII
mseI          mseI      mseI mboII(dam-)
ahaIII/draI          mboII(dam-)
5701 AAGCATTGGT AACTGTCAGA CCAAGTTTAC TCATATATAC TTTAGATTGA TTTAAACCTT CATTTTAAAT TTTAAAGGAT CTAGGTGAAG ATCCTTTTGG
TTCGTAACCA TTGACAGTCT GGTTCAAATG AGTATATATG AAATCTAACT AAATTTTGAA GTAAAAATTA AATTTTCTA GATCCACTTC TAGGAAAAAC

          sau3AI
          mboI/ndeII(dam-)
          dpmI(dam+) sau3AI
          dpmII(dam-) mboI/ndeII(dam-)
          bstYI/xhoII dpmI(dam+)
          sau3AI alwI(dam-) dpmII(dam-)
          mboI/ndeII(dam-) alwI(dam-)
          dpmI(dam+) mboII(dam-)
          dpmII(dam-) bstYI/xhoII
5801 ATAATCTCAT GACCAAAATC CCTTAACGTG AGTTTTCGTT CCACTGAGCG TCAGACCCCG TAGAAAAGAT CAAAGGATCT TCTTGAGATC CTTTTTCT
TATTAGAGTA CTGGTTTGTAG GGAATTGCAC TCAAAAGCAA GGTGACTCGC AGTCTGGGCG ATCTTTTCTA GTTTCCTAGA AGAACTCTAG GAAAAAAGA

          sau3AI
          mboI/ndeII(dam-)
          dpmI(dam+)
          dpmII(dam-)
          alwI(dam-)
          mspI
          hpaII      aluI
          mspI      acII      nspBII
          acII      nspBII
          acII      ACCACCGCTA CCAGCGGTGG TTTGTTTGGC GGATCAAGAG CTACCAACTC TTTTCCGAA GGTAACCTGC
5901 CGCGCATTAG ACGACGAACG TTTGTTTTTT TGGTGGCGAT GGTGCGCACC AACAAAACGG CCTAGTTCTC GATGTTGAG AAAAAGGCTT CCATTGACCG
          bsrI      maeIII      eco57I

          haeIII/palI
          haeI
          bslI      bslI
          rmaI      maeI
          hinPI      hhaI/cfoI
          hhaI/cfoI
6001 TTCAGCAGAG CGCAGATACC AAATACTGTC CTTCTAGTGT AGCGTAGTT AGGCCACCAC TTCAAGAACT CTGTAGCACC GCCTACATAC CTCGCTCTGC
AAGTCGTCTC GCGTCTATGG TTTATGACAG GAAGATCACA TCGGCATCAA TCCGGTGGTG AAGTCTTGA GACATCGTGG CGGATGTATG GAGCGAGACG

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>length: 6889
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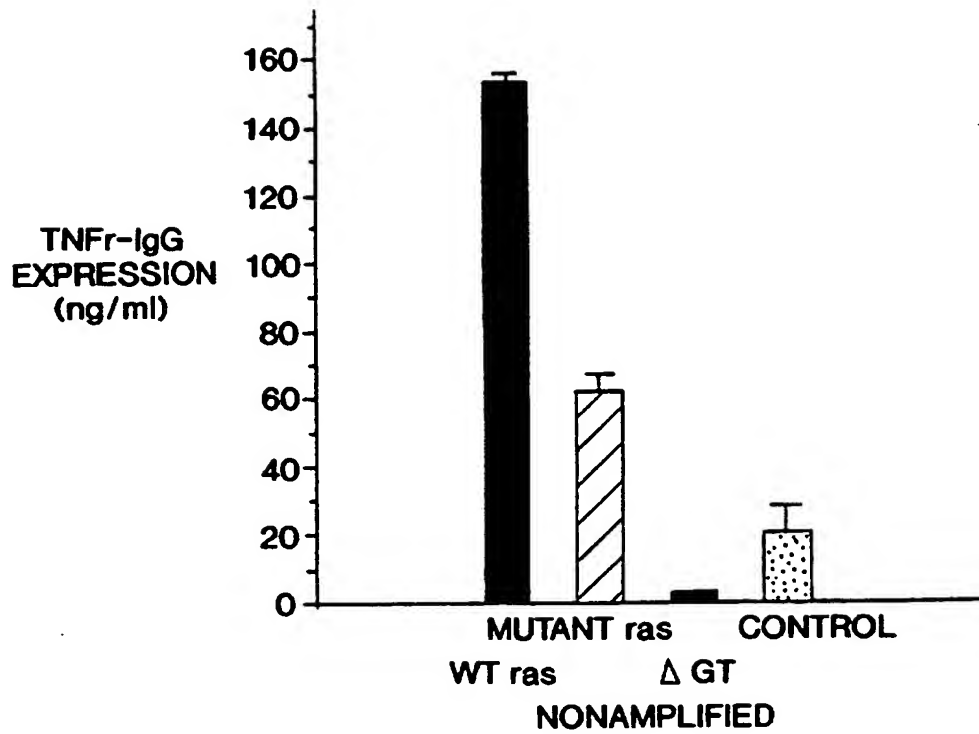


FIG. 7A

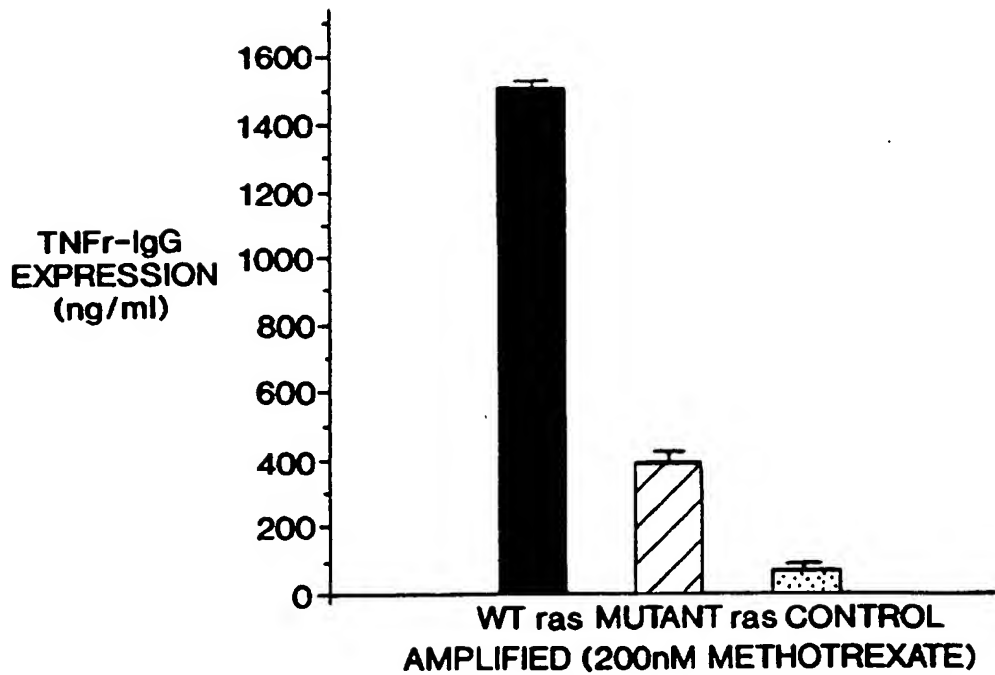


FIG. 7B

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FIG. 8

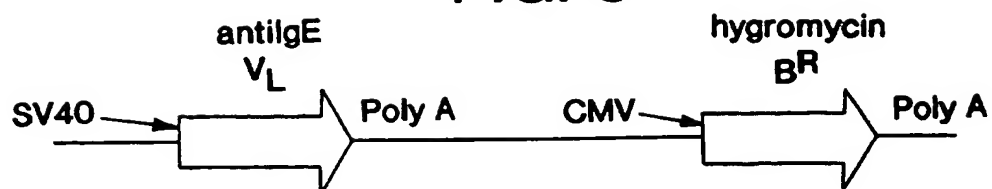


FIG. 11

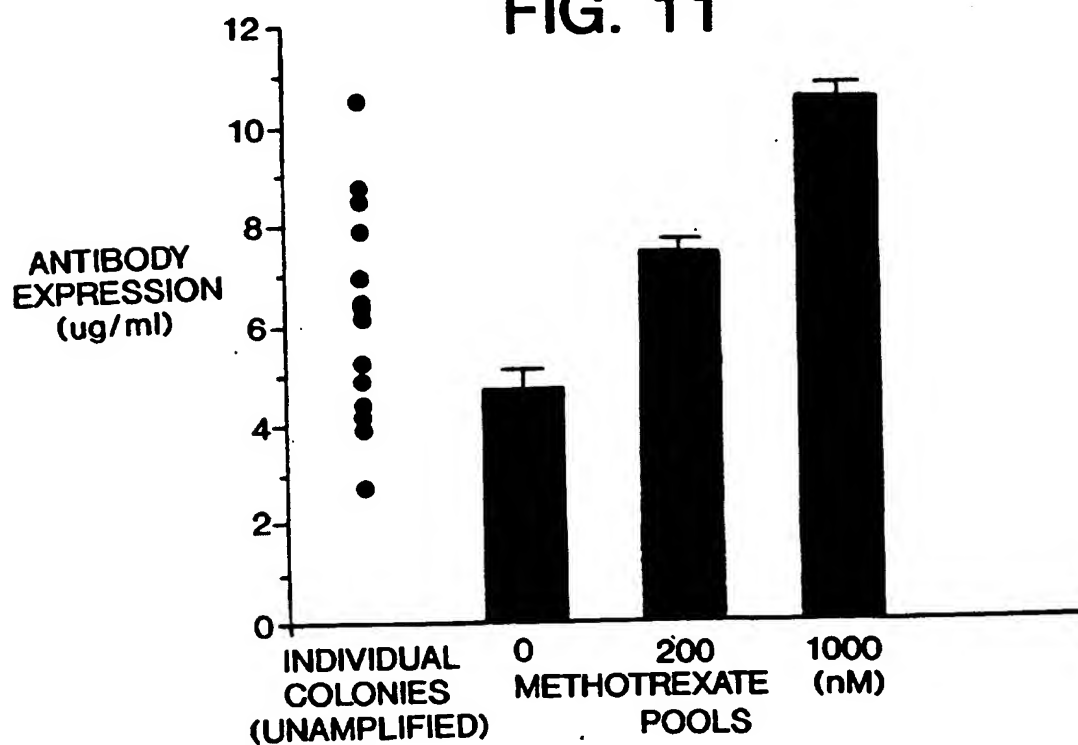


FIG. 9A

[illegible]

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FIG. 9B

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401 CCGGGAACGG TGCAATTGGAA CGCGGATTCC CCGTGCAAG AGTGACGTAA GTACCGCTA TAGACGATA AGAGGATTTT ATCCCGCTG CCATCATGGT
      scrFI      tfII      hinFI      aciI      fnu4HI      bbvI      nspBII      aciI      nlaIII
      nciI      mspI      hpaII      dsav      cauII      fnuDII/mvni      bstUI      bsh1236I      maeIII      rsaI      csp6I      scfI      mmlI
      dsav      cauII      fnuDII/mvni      bstUI      bsh1236I      maeIII      rsaI      csp6I      scfI      mmlI      aciI      nlaIII
      GGGCCCTGCC ACGTAACCTT GCGCCTAAGG GGCACGGTTC TCACTGCATT CATGGCGGAT ATCTGGCTAT TCTCTAANA TAGGGGCGAC GGTAATACCA
      haeIII/palI
      haeI
      scrFI      mvaI      bsrBI      ecorII      dsav      bstNI      aciI      xmmI      rsaI      csp6I
      taqI      sfaNI      pflMI      bsmAI      apyI(dcm+)      xmmI      asp700      scaI
501 TCGACCATTC AACTGCATCG TCGCCGTGTC CCAAAATATG GGGATTGGCA AGAAGCGAGA CCTACCTGG CCTCGCTCA GGAACGAGTT CAAGTACTTC
      taqI      sfaNI      pflMI      bsmAI      apyI(dcm+)      xmmI      asp700      scaI
      AGCTGGTAAC TTGACGTAGC AGCGGCACAG GGTTTTATAC CCTAACCGT TCTTGCTCT GGATGGACC GGAGCGAGT CCTTGCTCAA GTTCATGAAG
      scrFI      mvaI      ecorII      dsav      bstNI      apyI(dcm+)      xmmI      asp700      scaI
      taqI      sfaNI      pflMI      bsmAI      apyI(dcm+)      xmmI      asp700      scaI
601 CAAAGAATGA CCACAACCTC TTCAGTGGAA GGTAAACAGA ATCTGGTGAT TATGGTAGG AAAACCTGGT TCTCCATTC TGAGAGAGAT CGACCTTAA
      eco57I      mboII      earI/ksp632I      mnlI      tfII      hinFI      alwNI      hphI      ddeI      mboII      taqI      ahaII/draI
      GTTCTTACT GGTGTTGGAG AGTCACCTT CCAATTGTCT TAGACCACCTA ATACCATCC TTTTGACCA AGAGGTAAG ACTCTTCTTA GCTGGAATTT
      sstI      sacI      hgiIII      hgiAI/aspHI      ecl136II      bsp1286      bsiHFAI      bmyI      banII      bali      mmlI      aluI      bstXI      foki      sfaNI      mseI
      tru9I      mseI      aseI/asnI/vspI      ddeI      ahlII/bfrI
701 AGGACAGAAT TAATATAGTT CTCAGTAGAG AACTCAAGA ACCACCACGA GAGGCTCATTT TCTTGCCAA AAGTTGGAT GATGCCCTAA GACTTATTGA
      tru9I      mseI      aseI/asnI/vspI      ddeI      ahlII/bfrI
      TCTGTCTTA ATTATATCAA GACTCATCTC TTGAGTTCT TGGTGGTCT CCTCGAGTAA AAGAAGGTT TTCAACCTA CTACGGAAAT CTGAATAACT

```

FIG. 9C

801 ACAACCGGAA TTGGCAAGTA AAGTAGACAT GGTGGGATA GTGGAGGCA GTTCTGTTTA CCAGGAGCC ATGAATCAAC CAGGCCACCT TAGACTCTTT
 TGTGGCCTT AACCGTTTCAT TTCATCTGTA CCAACCTAT CAGCCTCGT CAAGACAAAT GGTCTTCGG TACTTAGTTG GTCCGTGGA ATCTGAGAAA
 mspI hpaII bsaBI
 accI nlaIII mnlI
 scrFI mvaI ecorII dsav bstNI nlaIII bstNI ddeI pleI
 scrFI mvaI ecorII dsav bstNI nlaIII bstNI ddeI pleI
 haeIII/palI haeI
 901 GTGACAAGGA TCATGCAGGA ATTTGAAAGT GACAGTTTT TCCAGAAAT TGATTTGGG AAATATAAC CTCTCCAGA ATACCCAGGC GTCTCTCTG
 CACTGTTCT AGTAGCTCT TAACTTTCA CTGTGCAAA AGGTCTTTA ACTAAACCC TTTATATTG GAGAGGTCT TATGGGTCCG CAGGAGAGAC
 nlaIII
 sau3AI mboI/ndeII[dam-] maeII
 dpnI[dam+] afIII
 dpnII[dam-] maeIII
 maeIII alwI[dam-] apoI
 1001 AGGTCCAGGA GGAAAAGGC ATCAAGTATA AGTTGAAGT CTACGAGAAG AAAGACTAAC AGGAAGATGC TTTCAGATTC TCTGCTCCC TCCTAAAGCT
 TCCAGGTCCT CCTTTTCCG TAGTTCATAT TCAAACTTCA GATGCTCTTC TTTCTGATTG TCCTTCTACG AAAGTTCAAG AGACGAGGG AGGATTTGGA
 scrFI mvaI ecorII dsav bstNI apyI[dcm+] sau96I
 asuI mnlI sfaNI accI mboII mboII mnlI aluI
 1101 ATGCATTTT ATAAGACCAT GGGACTTTTG CTGGCTTTAG ATCCCTTTG CTTCGTTAGA AGCAGCTAC AATTAAATACA TAACCTTATG TATCATACAC
 TACGTAAAA TATTCTGGTA CCTGTGGAAC GACCGAAATC TAGGGGAACC GAAGCAATCT TGCCTCATG TTAATTATGT ATTGGAATAC ATAGTATGTG
 ppu10I nsiI/avaII bsaJI
 nlaIII
 styI ncoI dsal
 sau3AI mboI/ndeII[dam-] dpnI[dam+] dpnII[dam-] alwI[dam-] bstVI/xhoII
 aluI fnu4HI mseI
 bstNI asnI/vspI

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FIG. 9E

FIG. 9E

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FIG. 9F

```

scrFI      scrFI      scrFI      scrFI      scrFI      scrFI      scrFI      scrFI      scrFI      scrFI
mvaI       mvaI       mvaI       mvaI       mvaI       mvaI       mvaI       mvaI       mvaI       mvaI
ecorII     ecorII     ecorII     ecorII     ecorII     ecorII     ecorII     ecorII     ecorII     ecorII
dsav       dsav       dsav       dsav       dsav       dsav       dsav       dsav       dsav       dsav
bstNI      bstNI      bstNI      bstNI      bstNI      bstNI      bstNI      bstNI      bstNI      bstNI
apyI[dcn+] apyI[dcn+] apyI[dcn+] apyI[dcn+] apyI[dcn+] apyI[dcn+] apyI[dcn+] apyI[dcn+] apyI[dcn+] apyI[dcn+]
hinPI      hinPI      hinPI      hinPI      hinPI      hinPI      hinPI      hinPI      hinPI      hinPI
hhaI/cfoI  hhaI/cfoI  hhaI/cfoI  hhaI/cfoI  hhaI/cfoI  hhaI/cfoI  hhaI/cfoI  hhaI/cfoI  hhaI/cfoI  hhaI/cfoI
nlaIV      nlaIV      nlaIV      nlaIV      nlaIV      nlaIV      nlaIV      nlaIV      nlaIV      nlaIV
nari       nari       nari       nari       nari       nari       nari       nari       nari       nari
kasi       kasi       kasi       kasi       kasi       kasi       kasi       kasi       kasi       kasi
hinII/acyI hinII/acyI hinII/acyI hinII/acyI hinII/acyI hinII/acyI hinII/acyI hinII/acyI hinII/acyI hinII/acyI
hgiCI      hgiCI      hgiCI      hgiCI      hgiCI      hgiCI      hgiCI      hgiCI      hgiCI      hgiCI
haeII      haeII      haeII      haeII      haeII      haeII      haeII      haeII      haeII      haeII
bani       bani       bani       bani       bani       bani       bani       bani       bani       bani
ahaII/bsaH ahaII/bsaH ahaII/bsaH ahaII/bsaH ahaII/bsaH ahaII/bsaH ahaII/bsaH ahaII/bsaH ahaII/bsaH ahaII/bsaH
1601 ACTGTCAGAT GAACAGCCTG CGTGTCTGAGG AACTGTGCGT CTATTATTGT GCCACTATTG CCGCGCTGG CACTTGGCCG TGTGGGGTCA
TGGACGTCTA CTTGTGGGAC GCACGACTCC TGTGACGGCA GATAATAACA CGAGCTCCGT CGTGATATAA GCCGCGGACC GTGAAGCGGC ACACCCAGT

mmlI       mmlI       mmlI       mmlI       mmlI       mmlI       mmlI       mmlI       mmlI       mmlI
xhoI       xhoI       xhoI       xhoI       xhoI       xhoI       xhoI       xhoI       xhoI       xhoI
paer7I     paer7I     paer7I     paer7I     paer7I     paer7I     paer7I     paer7I     paer7I     paer7I
avaI       avaI       avaI       avaI       avaI       avaI       avaI       avaI       avaI       avaI
hgiAI/aspH hgiAI/aspH hgiAI/aspH hgiAI/aspH hgiAI/aspH hgiAI/aspH hgiAI/aspH hgiAI/aspH hgiAI/aspH hgiAI/aspH
bsp1286    bsp1286    bsp1286    bsp1286    bsp1286    bsp1286    bsp1286    bsp1286    bsp1286    bsp1286
bsiHKAI    bsiHKAI    bsiHKAI    bsiHKAI    bsiHKAI    bsiHKAI    bsiHKAI    bsiHKAI    bsiHKAI    bsiHKAI
bmyI taqI bbvI bmyI taqI bbvI bmyI taqI bbvI bmyI taqI bbvI bmyI taqI bbvI bmyI taqI bbvI bmyI taqI bbvI bmyI taqI bbvI
sau96I     sau96I     sau96I     sau96I     sau96I     sau96I     sau96I     sau96I     sau96I     sau96I
haeIII/paI haeIII/paI haeIII/paI haeIII/paI haeIII/paI haeIII/paI haeIII/paI haeIII/paI haeIII/paI haeIII/paI
nlaIV      nlaIV      nlaIV      nlaIV      nlaIV      nlaIV      nlaIV      nlaIV      nlaIV      nlaIV
hgiJII     hgiJII     hgiJII     hgiJII     hgiJII     hgiJII     hgiJII     hgiJII     hgiJII     hgiJII
bsp1286    bsp1286    bsp1286    bsp1286    bsp1286    bsp1286    bsp1286    bsp1286    bsp1286    bsp1286
bani       bani       bani       bani       bani       bani       bani       bani       bani       bani
scrFI      scrFI      scrFI      scrFI      scrFI      scrFI      scrFI      scrFI      scrFI      scrFI
mvaI       mvaI       mvaI       mvaI       mvaI       mvaI       mvaI       mvaI       mvaI       mvaI
ecorII     ecorII     ecorII     ecorII     ecorII     ecorII     ecorII     ecorII     ecorII     ecorII
dsav       dsav       dsav       dsav       dsav       dsav       dsav       dsav       dsav       dsav
bstNI      bstNI      bstNI      bstNI      bstNI      bstNI      bstNI      bstNI      bstNI      bstNI
hphI       hphI       hphI       hphI       hphI       hphI       hphI       hphI       hphI       hphI
apyI[dcn+] apyI[dcn+] apyI[dcn+] apyI[dcn+] apyI[dcn+] apyI[dcn+] apyI[dcn+] apyI[dcn+] apyI[dcn+] apyI[dcn+]
bsaJI maeIII bsaJI maeIII bsaJI maeIII bsaJI maeIII bsaJI maeIII bsaJI maeIII bsaJI maeIII bsaJI maeIII bsaJI maeIII bsaJI maeIII
nlaIV bsteII esp3I bsaJI mmlI bsaJI mmlI bsaJI mmlI bsaJI mmlI bsaJI mmlI bsaJI mmlI bsaJI mmlI bsaJI mmlI bsaJI mmlI bsaJI mmlI
1701 AGGAACCCCTG GTACCGTCT CCTCGGCCTC CACCAAGGCG CCATCGGTCT TCCCTGGC ACCCTCTCC AGAGCACCT CTGGGGGAC AGCGCCCTG
TCCTTGGGAC CAGTGGCAGA GGAGCCGAG GTGTTTCCG GTAGCCAGA AGGGGACCG TGGGAGGAGG TTCTGTGGA GACCCCTCG TCCTGGGAC

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FIG. 9G

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scrFI      hinPI      hgiAI/asphi      hpaII      hpaII      hpaII
mvaI      nlaIV      bsp1286      bsiHKAI      bmyI      scrPI
ecoRII     nari      hpaII      hgiCI      haeII      bani
dsav      hinII/acyI      hgiCI      haeII      bani      fnu4HI
bstNI      hphI      mspi      hpaII      cfr10I      bsaWI
bsII      tth111I/aspi      bsaWI      tth111I/aspi      bsaWI
apyI[dcM+]      bsaWI      tth111I/aspi      bsaWI      tth111I/aspi
fnu4HI      bsaWI      tth111I/aspi      bsaWI      tth111I/aspi
bbvI      bsaWI      tth111I/aspi      bsaWI      tth111I/aspi
1801 GGCTGCCTCG TCAAGGACTA CTTCCTCCGAA CCGGTGACGG TGTGCTGGAA CTCAGGCGCC CTGACGACGG GCGTGCACAC CTTCCTCCGCT GTCTTACAGT
CGGACGGACC AGTTCTCTGAT GAAGGGGCTT GGCCACTGCC ACAGCACCTT GAGTCGGCGG GACTGTGCG CCGACGTGTG GAAGGGCCGA CAGGATGTCA

ddeI pleI      nlaIV      hgiCI      bniI      bniI      bniI
mnlI      hinfI      fnu4HI      hgiCI      bniI      bniI
eco81I      mnlI      fnu4HI      hgiCI      bniI      bniI
bsu36I/mstII/sauI ddeI bbvI bsp1286 rmaI bmyI maeI aluI bmyI
1901 CCTCAGGACT CTACTCCCTC AGCAGCGTGG TGACTGTGCC CTCTAGCAGC TTGGGACACC AGACCTACAT CTGCAAGTG ANTCAACAGC CCAGCAACAC
GGAGTCCTGA GATGAGGGAG TGTGCGACC ACTGACACGG GAGATCGTGG AACCGTGGG TGTGATGTA GACGTGCAC TTAGTGTGG GGTCTGTGG

eam11051
sau96I      scrFI      mvaI      avaiI      dsav      bstNI      bsaJI      nlaIV      bmyI[dcM+]      bbsI      mmlI
hgiJII      bsp1286      hgiJII      bsp1286      hgiJII      bsp1286      hgiJII      bsp1286      hgiJII      bsp1286      hgiJII
bmyI      bniI      maeIII      nspHI      nspHI      nspHI      nspHI      nspHI      nspHI      nspHI
2001 CAAGGTGGAC AAGAAGTTG AGCCCAATC TTGTGACAA ACTCACAT GCCCAGCTG CCCAGCACCT GAATCTCTGG GGGACCGTC AGTCTTCTC
GTTCACCTG TTCTTTCAAC TCGGTTTAG AACACTGTTT TGAGTGTGA CGGGGGCAC GGGTGGTGA CCGTGGACC CCGTGGACC TCAGAGGAG

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FIG. 9H

```

sau96I
nlaIV
avaII
mspI
scrFI
ncII
sau3AI hpaII
mboI/ndeII[dam-]
dpnI[dam+]
nlaIII dsav
rcal caulI
bspHI[dam-] asuI eco8II nspHI
nlaIII dsav mnlI mspI nlaIII
rcal caulI ddeI maeII mboII ddeI
bspHI[dam-] asuI eco8II nspHI bpuAI eco8II
mnlI dpnII[dam-] bsu36I/mstII/sauI maeII bbsI bsu36I/mstII/sauI
styI bsaJI
2101 TTCCCCCAA AACCCAGGA CACCTCATG ATCTCCCGA CCGCTGAGGT CACATGCGTG GTGGTGACG TGAGCCACGA AGACCTGAG GTCAAGTTCA
AAGGGGGT TTGGGTTCT GTGGAGTAC TAGAGGGCT GGGGACTCCA GTGTACGCAC CACCACCTGC ACTCGGTCT TCTGGGACTC CAGTTCAAGT
aciI
thai
fnuDI/mvnl
bstUI
bsh1236I
sacII/estII
nspBII
kspI
dsaI
bsaJI
maeII
rsaI
csp6I
bsrI bsaI
mnlI
bsrI bsaI
2201 ACTGTACGT GGACGGCGTG GAGTGCATA ATGCCAGAC AAGCCCGCG GAGGAGCAGT ACACAGCAC GTACCGTGTG GTACGGTCC TCACCGTCT
TGACCATGCA CTGCCGCAC CTCCACGTAT TACGGTTCTG TTTCCGGCC CTCTCTCA TGTGTCTG CATGGCAC CAGTGCAGG AGTGGCAGGA
maeII
rsaI
csp6I
bsrI bsaI
mnlI
bsrI bsaI
2201 ACTGTACGT GGACGGCGTG GAGTGCATA ATGCCAGAC AAGCCCGCG GAGGAGCAGT ACACAGCAC GTACCGTGTG GTACGGTCC TCACCGTCT
TGACCATGCA CTGCCGCAC CTCCACGTAT TACGGTTCTG TTTCCGGCC CTCTCTCA TGTGTCTG CATGGCAC CAGTGCAGG AGTGGCAGGA
scrFI
mvaI bsrI
ecoRII
dsav
bstNI
apyI[dcn+]
2301 GCACCCAGGAC TGGCTGAATG GCAAGGAGTA CAAGTGAAG GTCTCCAACA AAGCCCTCCC AGCCCCCATC GAGAAACCA TCTCCAAAGC CAAAGGCAG
CGTGGTCTCTG ACCGACTTAC CGTTCCTCAT GTTCACGTTT CAGAGTTGT TCGGGGAGG TCGGGGAGT CTCTTTTGT AGAGGTTCTG GTTCCCGTC
fnu4HI
bbvI

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taqI[dam-]
clai/bsp106[dam-]
sau3AI
mboI/ndeII[dam-]
dpmI[dam+]
dpmII[dam-]
nlaIII alwI[dam-]
ATC ATGTCGAT
GAG TACAGACCTA

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Fig. 9L

[illegible]

[illegible]

[illegible]

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FIG. 90

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mspI      hinPI      bsrI
hpaII     mstI       avII/fspI   tru9I
bsaWI     maeII     hhaI/cfoI   mseI
nlaIV     psp1406I
aluI      fnu4HI     bbvI      GGCACAACG TTGCGCAAC TATTAACCTGG
5001 ATCGTTGGG ACCGGAGCTG AATGAAGCCA TACCAACGA CGAGCTGAC ACCAGATGC CAGCACAAT GGCACAACG TTGCGCAAC TATTAACCTGG
TAGCAACCT TGGCCTCGAC TTACTTCGGT ATGTTTGTCT GCTCGACTG TGGTGCTACG GTGCTGCTTA CCGTTGTTGC AACGCGTTG ATAATTGACC

mspI      bglI
hpaII     sau96I
scrFI     haeIII/palI
ncII      hinPI     asuI     mspI
dsav      hhaI/cfoI   hpaII
maeI      fnu4HI     bbvI      GGCACAACG TTGCGCAAC TATTAACCTGG
5101 CGAACTACTT ACTTAGCTT CCCGGCAACA ATTAATAGAC TGGATGAGG CGGATAAAGT TGCAGACCA CTCTGCGCT CGGCCCTTCC GCGTGGCTGG
GCTTGATGAA TGAGATCGAA GGGCCGTTGT TAATTATCTG ACCTACTTCC GCTTATTCCA ACCTCTGCTG GAAGACGCGA GCCGGGAAGG CCGACCGACC

mspI      bglI
hpaII     sau96I
scrFI     haeIII/palI
ncII      hinPI     asuI     mspI
dsav      hhaI/cfoI   hpaII
maeI      fnu4HI     bbvI      GGCACAACG TTGCGCAAC TATTAACCTGG
5201 TTTATTGCTG ATAAATCTGG AGCGGTGAG CGTGGGTCTC GCGGTATCAT TGCAGCACTG GGGCCAGATG GTAAGCCCTC CCGTATCGTA GTTATCTACA
AAATAACGAC TATTAGACC TCGGCCACTC GCACCCAGAG CGCCATAGTA ACCTCGTGAC CCGGCTCTAC CATTCGGGAG GGCATAGCAT CAATAGATGT

pleI      ddeI
hinFI     sau3AI     nlaIV
eam1105I  mboI/ndeII[dam-] mnlI
fokI      dpnI[dam+] hgiCI     tru9I
          dpnII[dam-] baniI     mseI
          maeIII
5301 CGACGGGAG TCAGGCACT ATGGCACTG GAAATAGACA GATCGCTGAG ATAGGTGCTT CACTGATTA GCATTGGTAA CTGTCAGACC AAGTTTACTC
GCTGCCCTC AGTCCGTTGA TACCTACTTG CTTTATCTGT CTAGCGACTC TATCCACGGA GTGACTAATT CGTAACCAATT GACAGTCTGG TTCAATGAG

          rmaI     sau3AI
          sau3AI hphI mboI/ndeII[dam-]
          mboI/ndeII[dam-]
          dpnI[dam+] dpnII[dam+]
          tru9I dpnII[dam-] dpnII[dam-]
          ahaII/draI maeI alwI[dam-]
          tru9I bspVI/xhoII bspVI/xhoII
          mseI mseI alwI[dam-] mboI[dam-]
          ahaII/draI mseI mseI alwI[dam-] mboI[dam-]
          nlaIII rcaI bspHI
          maeII tru9I mseI
5401 ATATATACTT TAGATTGATT TAAACTTCA TTTTAAATTT AAAGGATCT AGGTGAAGAT CCTTTTTCAT AATCTCATGA CCAAAATCCC TTAACGCTGAG
TATATATGAA ATCTAACTAA ATTTGAACT AAAAATTA TTTTCTTACA TCCACTTCTA GGAAAAACTA TTAGAGTACT GGTTTTAGGG AATTGCACCTC

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FIG. 9P

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sau3AI
mboI/ndeII[dam-]
dpmI[dam+] sau3AI          thal
dpmII[dam-] mboI/ndeII[dam-]
bstYI/xhoII dpmI[dam+] fnuDII/mvnI
sau3AI alwI[dam-] dpmII[dam-] bstUI
mboI/ndeII[dam-] alwI[dam-] bsh1236I
dpmI[dam+] mboII[dam-] hinPI fnu4HI
dpmII[dam-] bstYI/xhoII hhaI/cfoI bbvI
hgaI
ddeI
5501 TTTTCGTTCC ACTGAGCGTC AGACCCCGTA GAAAGATCTTC TTGAGATCCT TTTTTCCTGC GCGTAATCTG CTGCTTGCAA ACAAAAAAAC
AAAAGCAAGG TCACTCCAG TCTGGGGCAT CTTTCTAGT TTCCTAGAAG AACTTAGGA AAAAAGACG CGCATTAGAC GACGAACGTT TGTTCCTTTCG

sau3AI
mboI/ndeII[dam-]
dpmI[dam+]
dpmII[dam-]
alwI[dam-]
mspi
acin
nspBII
hpaII
aluI
5601 CACCGCTACC AGCGGTGTT TGTTCGCGG ATCAAGAGCT ACCAACTCTT TTTCCGAAGG TAACTGGCTT CAGCAGAGCG CAGATACCAA ATACTGTCTT
GTGGCGATGG TCGCCACCAA ACAACGGCC TAGTTCTCGA TGCTTGAGAA AAAGGCTTCC ATGACCGAA GTCGTCTCGC GTCTATGCTT TATGACAGGA

xmaI      haeIII/palI
maeI      bslI      haeI      scfI      acII      mnlI      maeIII      bbvI      bsrI      fnu4HI
AGATCACATC GGCATCAATC CGGTGGTGAA GTTCTTGAGA CATCGTGGCG GATGTATGGA GCGAGACGAT TAGGACAATG GTCACCGACG ACGGTCAACG

scrFI      ncII      mspi      hpaII      dsav      cauII      hinfI      pleI      bsaWI      hinPI      mcrI      aciI      nspBII      fnu4HI      bbvI
5701 TCTAGTGTAG CCGTAGTTAG GCCACCACTT CAAGAACTCT GTAGCACCGC CTACATACCT CGTCTGTGTA ATCTGTGTAC CAGTGGCTGC TGCCAGTGGC
AGATCACATC GGCATCAATC CGGTGGTGAA GTTCTTGAGA CATCGTGGCG GATGTATGGA GCGAGACGAT TAGGACAATG GTCACCGACG ACGGTCAACG

hgiAI/aspHI
bsp1286
bsiHKA1
bmyI
apaLI/snoI
alw44I/snoI
aluI
5801 GATAAGTCGT GTCTTACCG GTTGACTCA AGACGATAGT TACCGATAA GCGGAGCGG TCGGGGTGAA CGGGGGGTTT GTGCACACAG CCCAGCTTGG
CTATTCAGCA CAGATGGCC CAACCTGAGT TCTGTATCA ATGCCCTATT CCGCGTCGCC AGCCCGACTT GCCCCCCAAG CACGTGTGTC GGGTCGAACC

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FIG. 9Q

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5901 AGCGAAGAC CTACACCGAA CTGAGATACC TACAGCGTGA GCATTGAGAA AGCGCCACGC TTCCCGAAGG GAGAAAGCGG GACAGGTATC CGGTAAGCGG
TCGCTTGCTG GATGTGGCTT GACTCTATGG ATGTGCACT CGTAACCTTT TCGCGGTGG AAGGCTTCC CTCTTCCG CTTGCCATAG GCCATTCCGC
                                     hinPI          hhaI/cfoI          haeII          aciI          bsaWI          fnu4HI
                                     mspI          hpaII          bslI          bsaWI          aciI
                                     scrFI          mvaI          ecorII          dsav          bstNI          bsaJI          apyI[dcM+]          mmlI          drdI          hgaI          taqI
6001 CAGGTCGGA ACAGGAGAGC GCACGAGGGA GCTTCCAGG GGAAGCCCT GTATCTTTA TAGTCCTGTC GGGTTTCGCC ACCTCTGACT TGAGCGTCGA
GTCCAGCCT TGTCTCTCG CGTCTCTCCT CGAAGGTCCC CTTTCCGGA CCATAGAAAT ATCAGGACAG CCCAAAGCGG TGGAGACTGA ACTCGCAGCT
                                     hinPI          mmlI          hhaI/cfoI          aluI          apyI[dcM+]          dsav          bstNI          bsaJI          apyI[dcM+]          haeIII/palI
                                     scrFI          mvaI          ecorII          dsav          bstNI          bsaJI          apyI[dcM+]          haeIII/palI
                                     fnu4HI          mvaI          ecorII          dsav          bstNI          bslI          apyI[dcM+]          haeI          haeIII/palI          nspI          nlaIII
                                     nlaIV          aciI          bsh1236I          nlaIV          haeI          haeI          haeI          haeI          haeI          haeI          haeI          haeI
6101 TTTTGTGAT GTCGTGTCAG GGGCGGAGC CTATGGAAA ACGCCAGCAA CGCGGCTTT TTACGGTTC TGGCCTTTTG CTGGCTTTT GCTCACAATG
AAAAACACTA CGAGCAGTCC CCCCCTCTCG GATACCTTTT TCGGTCGTT TCGGTCGTT TCGGTCGTT TCGGTCGTT TCGGTCGTT TCGGTCGTT TCGGTCGTT TCGGTCGTT
                                     sfaNI          nlaIV          aciI          bsh1236I          nlaIV          haeI          haeI          haeI          haeI          haeI          haeI          haeI
                                     tfiI          hinFI          aciI          aluI          bsrBI          aciI          fnu4HI          mcrI          hhaI/cfoI          fnu4HI          bbvI          pleI          hinPI          hinFI
6201 TCTTTCCTGC GTTATCCCCT GATTCGTGG ATAACCGTAT TACCGCTTT TACCGCTTT TACCGCTTT TACCGCTTT TACCGCTTT TACCGCTTT TACCGCTTT TACCGCTTT TACCGCTTT
AGAAAGGAGC CAATAGGGA CTAAGACACC TATTGGCATA ATGGCGAAA CTCACCTCGAC TATGGCGAGC GCGTCGCT TCGTGGCTCG GTCGCTCAG

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FIG. 9R

```

thai
fndI11/mvni
bstUI
bsh1236I
hinPI
hhaI/cfoI
thai
fndI11/mvni
bstUI
bsh1236I haeI11/paI1 tru9I aluI
mnlI bslI eaeI tfil aseI/asnI/vspi pvuI1
aciI cfrI hinfi msei nsbI1
6301 AGTGAGCGAG GAAGCGGAG AGGCCCAAT AGGCAACCG CCTCTCCCC CGCGTTGGCC GATTCAATTA TCCAGCTGGC ACGACAGGTT TCCCGACTGG
TCACTCGCTC CTTCGCCTTC TCGCGGTTA TCGGTTTGGC GGAGAGGGG GCGCAACCG CTAAGTAATT AGTCCGACC TGCTGTCCAA AGGGCTGACC
scrFI
mvaI
ecorII
dsav
nlaIV bstNI
hgiCI apyI(dcm+) mspi
banI bsaJI hpaII
6401 AAAGCGGGCA GTGAGCGCAA CGCAATTAAT GTGAGTTACC TCACTCATTG GGCACCCAG GCTTTACACT TTATGCTTCC GGCTCGTATG TTGTGTGGAA
TTTCGCCCGT CACTCGCGTT GCGTTAATTA CACTCAATGG AGTGAGTAAT CCGTGGGCTC CGAATGTGA AATACGAAG CCGAGCATAC AACACACCTT
tru9I
msei
aseI/asnI/vspi
xmuI
6501 TTGTGAGCGG ATAACAATTT CACACAGGAA ACAGCTATGA CCATGATTAC GAATTAA
AACACTCGCC TATTGTTAA GTGTGTCCTT TGTGATACT GGTACTAATG CTTAATT
>length: 6557

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FIG. 10A

```

aluI          thuI          fnuDII/mvnI
sstI          bstUI
sacI          bsh1236I
hgiJII
hgiAI/aspHI
ec1136II
bsp1286
bsiHKA1
bmyI
banII
taqI          acII maeIII
1  TTGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAAT AGTAATCAAT TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCGCG GTTACATAAC
AAGCTCGAGC GGGCTGTAAC TAATAACTGA TCAATAATTA TCATTAGTTA ATGCCCCAGT AATCAAGTAT CGGTATATA CCTCAAGGCG CAATGTATTG

          rmaI          tru9I          maeI          bslI
          speI          aseI/asnI/vspI
          scrFI          mvaI          ecorII          dsav
          aciI          bgli          bstNI          sau96I          haeIII/palI          aciI          asuI          apyI(dcm+)
          maeII          hinII/acyI          ahaII/bsaHI          aatII          maeIII
101 TTACGGTAA TGGCGGCCT GCGTACCGC CCAACGACCC CCGCCCATTT AGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA
AATGCCATTT ACCGGCGGA CCGACTGGC GGTTCCTGGG GCGGGGTAAC TGCAGTTATT ACTGCATACA AGGTATCAT TCGGGTTATC CCTGAAAGGT

          maeII          hinII/acyI          ahaII/bsaHI          aatII          maeII          maeIII
          rsaI          rsaI          csp6I          csp6I          csp6I
201 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCAC TTGGCAGTAC ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT
AAGTGCAGTT ACCCACTCA TAAATGCCAT TTGACGGGTG AACCGTCATG TAGTTCACAT AGTATACGGT TCATGCGGGG GATAACTGCA GTTACTGCCA

          scrFI          mvaI          ecorII          aciI          bgli          dsav          sau96I          haeIII/palI          asuI          apyI(dcm+)          bsrI          nlaIII
          rsaI          csp6I          rsaI          snaBI          maeII          styI          nlaIII
301 AAATGGCCG CCTGGCATT TGGCCAGTAC ATGACCTTAT GGGACTTTCC TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC
TTTACCAGGC GGACCGTAAT ACGGTTCATG TACTGGAATA CCTGAAAGG ATGAACCGTC ATAGTAGTC ATAATCAGTA GCGATAATGG TACCACCTAGC

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FIG. 10B

401 GGTGTTGGCA GTACATCAAT GCGCGTGGAT ACGGGTTTGA CTCACGGGGA TTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA
 CCAAAACCGT CATGTAGTTA CCGCACCTA TCGCCAAACT GAGTGCCCT AAGGTTTCA AGGTGGGTA ACTGCAGTTA CCTCAACA AAACCGTGTG

501 AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGAGC AATGGGCGG TAGCGGTGTA CCGTGGGAGG TCTATATAAG CAGAGCTCGT
 TTTAGTTGCC CTGAAGGTT TTACAGCATT GTTGAGGCGG GGTAACTGGG TTTACCGGCC ATCCGCACAT GCCACCTCC AGATATATTC GTCTCGAGCA

601 TTAGTGAACC GTCAGATCC CTGGAGACG CATCCACGT GTTTGACCT CCATAGAAGA CACCGGACC GATCCAGCT CCGCGCGCGG GAACGGTGCA
 AATCATTGG CAGTCTAGG GACCTCTCG GTAGTGCGA CAAAACTGGA GGTATCTTCT GTGCGCCTGG CTAGTCTGGA GGCGCGCGCC CTGCCCAGT

Restriction Enzymes:

- rsal csp61 maeIII acil hgaI hinfI pleI acil hinfI
- hinII/acyI nlaIV hgiCI bni
- ahaII/bsaHI aatII bsmAI bsp1286 bsiHKA bmyI banII
- aluI sstI sacI hgiJII hgiAI/aspHI ecl136II
- xbaI mcrI eaeI cfrI fnu4HI aciI
- haeIII/palI mndII/mvnI
- sau96I sacII/sstII nspBII kspI scrFI dsaI nciI bglI bslI mspI
- asuI avaII hpaII mspI mboI/ndeII(dam-) hpaII dpaI(dam+) bsaI dsaV
- nlaIV nciI sau3AI mnlI bstUI mboI/ndeII(dam-) hpaII dpaI(dam+) bsaI dsaV
- scrFI nciI mspI mboI hpaII dpaI dsaV cauII aluI(dam-) aciI cauII
- bstNI hinII/acyI apyI(dcm+) sau3AI gsuI/bpmI mboI/ndeII(dam-) dpaI(dam+) hgaI foki
- dpaII(dam-) ahaII/bsaHI

FIG. 10C

[illegible]

FIG. 10D

hinPI mspI
 hhaI/cfoI hpaII
 thal mroI
 acII bspMII
 haeIII/palI
 mcrI fnuDII/mvnl bspEI
 eagI/xmaIII/ecI XI bsaWI
 eaeI bstUI tfil
 cfrI bsh1236I hinfI
 sfanI fnu4HI bslI accIII
 CTTTGCATCG GCCGCGCTCC CGATTCCGGA AGTGCCTTAC ATTGGGGAAT TCAGCGAGAG CCTGACCTAT TGCATCTCCC
 1101 GATCGTTATG TTTATCGGCA CTTTGCATCG GCCGCGCTCC CGATTCCGGA AGTGCCTTAC ATTGGGGAAT TCAGCGAGAG CCTGACCTAT TGCATCTCCC
 CTAGCAATAC AAATAGCCGT GAAACGTAGC CGGCGCGAGG GCTAAGGCT TCACGAACTG TAACCCCTTA AGTCGCTCTC GGACTGGATA ACGTAGAGGG
 sau3AI
 mboI/ndeII(dam-) sfanI acII
 dpnII(dam+) apoI
 dpnII(dam-) ecorI
 hgiAI/asphi
 bsp1286
 bsiHKA1
 bmyI
 apaLI/snoI
 alw44I/snoI maeII
 bslI draIII maeIII
 1201 GCCGTGCACA GGGTGTACG TTGCAACACC TGCCTGAAC CGAACTGCC CTTGTCTTGC AGCCGCTGC GGAGGCCATG GATCGATCG CTGCGGCCGA
 CGGCACGTGT CCCACAGTGC AACGTTGTGG ACGGACTTTG CTTTGACGGG CGACAAGACG TCGGCCAGCG CTCCGGTAC CTACGCTAGC GACGCCGGCT
 sau96I
 avall
 asuI
 sau96I rsrII/cspI
 haeIII/palI acII tfil
 asuI cpoI hinfI
 ddeI acII bsrBI
 1301 TCCTAGCCAG ACGAGCGGGT TCGGCCCATT CGGACCCGAA GGAATCGGTC AATACACTAC ATGGCGTGT TTTATATGCG CGATTGCTGA TCCCATGTG
 AGAATCGGTC TGCTCGCCCA AGCCGGGTAA GCCTGGCGTT CCTTAGCCAG TTATGTGATG TACCGCACTA AAGTATACG GCTAAGGCT AGGGGTACAC
 hinPI
 hhaI/cfoI
 thal
 fnuDII/mvnl
 bstUI
 bsh1236I
 taqI
 aluI
 sfanI
 sau96I
 haeIII/palI
 bsaJI
 dralII
 nlaIV
 hgiCI
 bani
 mspI
 bslI
 hpaII
 1401 TATCACTGGC AAACGTGTAT GGACGACACC GTCAGTGGT CCGTCTCGAT GAGCTGATGC GAGCTGCCGA TTTGGGCCGA GGACTGCCCC GAAGTCCGCG
 ATAGTGACCG TTTGACACTA CCTGCTGTGG CAGTCACGCA GGCAGCGCGT CCGAGAGCTA CTCGACTACG AAACCCGGCT CTGACGGGG CTTCAGGCCG

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FIG. 10F

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nlaIV
mspI      scrFI
hpaII     nciI
bslI      mspI
mroI      hpaII
bspMI     hpaII
bspEI[dam-]
bsaWI     dsav
accIII[dam-]
sau3AI    cauII
mboI/ndeII[dam-]
dpmI[dam+]
dpmII[dam-]
alwI[dam-]

1801  ACGCAATCGT  CCGATCCGGA  GCCGGGACTG  TCGGGCGTAC  ACAAATCGCC  CGCAGAGCG  CGGCCGTCTG  GACCGATGGC  TGTGTAGAAG  TACTCGCCGA
TGC GTTAGCA  GGCTAGGCT  CGCCCTGAC  AGCCCGCATG  TGTTAGCGG  GCGTCTTCG  GCCGCAGAC  CTGGCTACCG  ACACATCTTC  ATGAGCGGCT

rsal      csp6I
hpaII     nciI
hpaII     dsav
xmaI/pspAI
smaI
scrFI
nciI
dsav
cauII
bsaJI
avaI
bsaJI
sau3AI
mboI/ndeII[dam-]
dpmI[dam+]
dpmII[dam-]
alwI[dam-]
nlaIV cauII
bstYI/xhoII
bamHI bsaJI ecoRI
alwI[dam-] apoI
ciaoI/bspl06 bsaJI
taqI haeIII/palI
acil      fnu4HI asuI
bglI      nlaIII
sfii      styI
eaeI      ncoI
cfrI      dsal
sau96I

hinli/acyI
hgaI
ahaII/bsaHI
CGACGCCCA  GCACTCGTCC  GAGGGCAAG  GAATAGACTA  GATCCGACC  GAAGGATCCC  CGGGGATTC  AATCGATGGC  CGCCATGGCC
ATCACCTTTG  GCTCGGGGT  CGTGAGCAGG  CTCCCGTTTC  CTATCTCAT  CTACGGCTGG  CTTCTAGCG  GCCCTTAAG  TTAGCTACCG  GCGGTACCGG

1901  TAGTGAAAC  CGACGCCCA  GCACTCGTCC  GAGGGCAAG  GAATAGACTA  GATCCGACC  GAAGGATCCC  CGGGGATTC  AATCGATGGC  CGCCATGGCC
ATCACCTTTG  GCTCGGGGT  CGTGAGCAGG  CTCCCGTTTC  CTATCTCAT  CTACGGCTGG  CTTCTAGCG  GCCCTTAAG  TTAGCTACCG  GCGGTACCGG

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FIG. 10G

2001 CAACCTGTTT ATTCGAGCTT ATAATGGTTA CAAATTAAGC AATAGCATCA CAAATTTCAC AAATAAGCA TTTTTCAC TGCATCTAG TTGTCGTTG
 GTTGAACAA TAACGTCGAA TATTACCAAT GTTTATTTCG TTATCGTAGT GTTAAAGTG TTTATTTCGT AAAAAAGTG AGTAAGATC AACACCAAC
 aluI fnu4HI bsvI maeIII sfaNI apoI rnaI bsmI maeI
 sau3AI mboI/ndeII(dam-) dpnI(dam+) dpnII(dam-) pvuI/bspCI mcrI
 taqI(dam-) tru9I clai/bsp106(dam-) sau3AI mseI fnu4HI styI haeIII/palI haeI
 mboI/ndeII(dam-) dpnI(dam+) xmnI hinPI dsai bsvI ncoI
 dpnII(dam-) aseI/asnI/vspI bsaJI
 nlaIII alwI(dam-) asp700 hhaI/cfoI nlaIII mnlI
 2101 TCCAAACTCA TCAATGTATC TTATCATGTC TGGATCGATC GGAATTAAT TCGCGCAGC ACCATGCGCT GAATAACCT CTGAAAGAGG AACTTGTTA
 AGGTTTGAGT AGTTACATAG AATAGTACAG ACCTAGCTAG CCCTTAATTA AGCGCGTGC TGGTACCGA CTTTATTGGA GACTTCTCC TTGAACCAAT
 rsaI csp6I nlaIV kpnI hgiCI bsaJI aluI pvuII nspBII
 asp718 acc65I ddeI aciI
 2201 GGTAACCTTCT GAGGCGGAAA GAACCACTG TGGATGTGT GTCAGTTAGG GTGTGGAAG TCCCAGGCT CCCACGAGG CAGAAGTATG CAAAGCATGC
 CCATGGAAGA CTCCGCTTT CTGGTCGAC ACCTTACACA CAGTCAATCC CACACCTTC AGGGTCCGA GGGTCTGTC GTCTTCATAC GTTTCGTACG
 scrFI mvaI ecorII dsav bstNI apyI(dcm+) bsaJI nlaIV
 scrFI mvaI ecorII dsav bstNI apyI(dcm+) bsaJI nlaIV
 2301 ATCTCAATTA GTCAGCAACC AGGTGTGGA AGTCCCCAGG CTCCCCAGCA GGCAGAAGTA TGCAAGCAT GCATCTCAAT TAGTCAGCAA CCATAGTCCC
 TAGAGTTAAT CAGTCGTTGG TCCACACCTT TCAGGGGTCC GAGGGTCTC CCGTCTTCAT AGTTTCGTA CGTAGAGTA ATCAGTCGTT GGTATCAGG
 scrFI mvaI ecorII dsav bstNI apyI(dcm+) bsaJI nspHI aciI
 scrFI mvaI ecorII dsav bstNI apyI(dcm+) bsaJI nspHI nspI sfaNI nspHI

FIG. 10H

[illegible]

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FIG. 10I

sau3AI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 alwI(dam-)
 taqI(dam-)
 claiI/bsp106(dam-)
 sau3AI
 mboI/ndeII(dam-)
 dpnI(dam+)
 dpnII(dam-)
 alwI(dam-)
 foki
 2701 ACCTTTTGA TCGATCCTAC TGACACTGAC ATCCACTTTT TCTTTTCTC CACAGGTGTC TCCAACTGCA CCTCGGTTTC CGAAGCTAGC
 TGGAAAACCT AGCTAGGATG ACTGTGACTG TAGGTGAAAA AGAAAAAGAG GTGTCCACAG GTGAGGGTTC AGGTTGACGT GGAGCCAAGC GCTTCGATCG

sau96I
 avalI
 asuI
 scrFI
 mvaI
 ecoRII
 dsav
 bstNI
 apyI(dcm+)
 bsII bsaJI
 mnII bsh1236I aluI
 bsaJI nruI aluI
 mnII bsh1236I aluI
 bstUI nheI
 fnuDII/mvni
 thalI maeI
 zmaI

nlaIII
 styI
 pfIM
 ncoI
 dsal
 bsII foki
 bsajI
 fnu4HI taqI apoI
 bbvI claiI/bsp106
 sfaNI ecoRI
 fnu4HI taqI apoI
 bbvI claiI/bsp106
 TCCACCATGG GATGGTCATG TATCATCCTT TTTCTAGTAG CAACTGCAAC TGGAGTACAT TCAGATATCC AGCTGACCCA
 AACCCGACGT AGCTAACTTA AGTGGTACC CTACCAGTAC ATAGTAGGAA AAAGATCATC GTTGACGTTG ACCTCATGTA AGTCTATAGG TCGACTGGGT

aluI
 sstI
 sacI
 hgiJII
 hgiAI/aspHI
 ecl136II
 bsp1286
 bsiHKA
 bmyI
 banII
 aciI
 mnII
 hphI
 maeIII
 bstEII
 hphI
 bspMI
 hphI
 bsrI
 hgaI
 taqI
 bsrI
 nlaiII

2801 TTGGGCTGCA TCGATTGAAT TCCACCATGG GATGGTCATG TATCATCCTT TTTCTAGTAG CAACTGCAAC TGGAGTACAT TCAGATATCC AGCTGACCCA
 AACCCGACGT AGCTAACTTA AGTGGTACC CTACCAGTAC ATAGTAGGAA AAAGATCATC GTTGACGTTG ACCTCATGTA AGTCTATAGG TCGACTGGGT

2901 GTCCCGGAGC TCCTGTGTCG CCTCTGTGCG CGATAGGGTC ACCATCACCT GCCGTGCCAG TCAGAGCGTC GATTACGATG GTGATAGCTA CATGAACTGG
 CAGGGGCTCG AGGACACAGC GGAGACACCC GCTATCCAG GCTATAGTGA TGGTAGTGA CGGCACGGTC AGTCTGCGAG CTAATGCTAC CACTATCGAT GTACTTGACC

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FIG. 10J

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mspI      gsuI/bpmI      mboI/ndeII[dam-]
hpall     scrFI      dpnI[dam+]
bsII      mvaI
bsaWI
sau3AI
mboI/ndeII[dam-]
dpnI[dam+]
alwI[dam-]
nlaIV
bstYI/xhoII
bamHI
alwI[dam-]
CTTCTCGCTT CTCTGGATCC GGTTCGGGA
3001 TATCAACAGA AACCAGGAAA AGCTCGGAAA CTACTGATTT AGCGGCCTC GTACCTGGAG TCGGAGTCC CTTCTCGCTT CTCTGGATCC GGTTCGGGA
ATAAGTTGCT TTGGTCTTT TCGAGGCTTT GATGACTAAA TCGCGCGGAG CATGGACCTC AGACCTCAGG GAAGAGCGAA GAGACCTAGG CCAAGACCTT
scrFI      fnu4HI      haeIII/palI      gsuI/bpmI      mboI/ndeII[dam-]
mvaI      ecorII      dsav      dpnI[dam+]
alwI[dam-]
thaI mnlI bstNI
fnu4II/mvni apyI[dcM+] pleI
bstUI rsaI pleI gsuI/bpmI
bsh1236I csp6I hinfI hinfI
3001 TATCAACAGA AACCAGGAAA AGCTCGGAAA CTACTGATTT AGCGGCCTC GTACCTGGAG TCGGAGTCC CTTCTCGCTT CTCTGGATCC GGTTCGGGA
ATAAGTTGCT TTGGTCTTT TCGAGGCTTT GATGACTAAA TCGCGCGGAG CATGGACCTC AGACCTCAGG GAAGAGCGAA GAGACCTAGG CCAAGACCTT
fnu4HI mboII
bbvI bpuAI
scfI bbsI
pstI mspI
bsgI hpall
3101 CGGATTTTAC TCTGACCATC AGCAGTCTGC AGCGGGAAGA CTTCGCAACT TATTACTGTC AGCAAAGTCA CGAGGATCCG TACACATTG GACAGGGTAC
GCCTAAAGTG AGACTGGTAG TCGTCAGAG TCGGCCTTCT GAAGGTTGA ATAATGACAG TCGTTTCAGT GCTCCTAGG ATGTGTAAC CTGTCCCATG
sau3AI      mboI/ndeII[dam-]      bsaJI
dpnI[dam+]      fnu4HI      bpuAI      acII
dpnII[dam-]      bbvI      bbsI      mboII
3201 CAAGGTGGAG ATCAACGAA CTGTGGCTGC ACCATCTGTC TTCACTTCC CGCCATCTGA TGAGCAGTTG AAATCTGGA CTGCTCTGT TGTGTGCTG
GTTCCACCTC TAGTTTGCTT GACACCGAG TGGTAGACAG AAGTAGAAG GGGTAGACT ACTCGTCAAC TTTAGACCTT GACGGAGACA ACACAGGAC
haeIII/palI      mnlI      apyI[dcM+]      maeIII
hael      rsaI      bsaJI      maeIII
mnlI      csp6I
3301 CTGAATAACT TCTATCCAG AGAGCCAAA GTACAGTGA AGGTGATAA CGCCTCCAA TCGGTTACT CCCAGGAG TGTACAGAG CAGGACAGCA
GACTTATTGA AGATAGGTC TCTCGGTTT CATGTCACCT TCCACCTATT TCCACCTATT GCGGAGGTT AGCCCAATGA GGTCTCTCTC ACAGTCTC GTCTGTGCT

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FIG. 10K

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sstI      hgiIII
sacI      hgiIII
hgiIII    hgiAI/aspHI
ecII36II  bspI286
bsiHKAI   bmyI
           haeIII/palI
           sau96I aluI
           asuI banII
           hphI   eco0109I/draII
           maeIII alwNI ddeI
3401 AGGACAGCAC CTACAGCCTC AGCAGCACCC TGACGCTGAG CAAAGCAGAC TAGAGAAAC ACAAGTCTA CGCCTGGAA GTCACCCATC AGGCGCTGAG
TCCTGTCTG GATGTGGG GATGTGGG ACTGCGACTC GTTCTGCTG ATGCTCTTG TGTTCAGAT GCGGACGCTT CAGTGGGTAG TCCGGGACTC
           ddeI
           celII/espI
           bpulI02I
           hgaI
           ddeI fnu4HI
           mnlI bbvI
           scfI
           3401 AGGACAGCAC CTACAGCCTC AGCAGCACCC TGACGCTGAG CAAAGCAGAC TAGAGAAAC ACAAGTCTA CGCCTGGAA GTCACCCATC AGGCGCTGAG
TCCTGTCTG GATGTGGG GATGTGGG ACTGCGACTC GTTCTGCTG ATGCTCTTG TGTTCAGAT GCGGACGCTT CAGTGGGTAG TCCGGGACTC
           sau96I
           nlaIII
           aciI haeIII/palI
           fnu4HI asuI
           bglI styI
           aluI sfiI ncoI
           hindIII eaeI dsal
           tru9I cfrI bsaJI
           mseI taqI haeIII/palI
           maeIII aluI
           3501 CTCGCCCGTC ACAAGAGCT TCAACAGGGG AGAGTGTAA GCTTCGATGG CCGCCATGGC CCAACTTGT TATTGCAGCT TATAATGTT ACAATAAAG
GAGCGGGCAG TGTTCCTGA AGTTGTCCCC TGTACAAAT CGAAGCTACC GCGGTACCG GGTGAACAA ATAACGTGA ATATTACCA TGTATTTC
           sau3AI
           mboI/ndeII(dam-)
           dpnI(dam+)
           dpnII(dam-)
           pvuI/bspCI
           mcrI
           taqI(dam-)
           claI/bspl06(dam-)
           sau3AI
           mboI/ndeII(dam-)
           dpnI(dam+)
           dpnII(dam-)
           nlaIII alwI(dam-)
           3601 CAATAGCATC ACAATTTCA CAAATAAGC ATTTTTTCA CTGCATTCTA GTTGTGTTT GTCCAACTC ATCAATGTAT CTTATCATGT CTGCATCGAT
GTTATCGTAG TGTATAAGT GTTTATTTCG TAAAAAAGT GACGTAAGT CAACACCACA CAGTTTGG TAGTTACATA GAATAGTACA GACCTAGCTA
           xmaI
           bsmI maeI
           sfaNI apoI
           3601 CAATAGCATC ACAATTTCA CAAATAAGC ATTTTTTCA CTGCATTCTA GTTGTGTTT GTCCAACTC ATCAATGTAT CTTATCATGT CTGCATCGAT
GTTATCGTAG TGTATAAGT GTTTATTTCG TAAAAAAGT GACGTAAGT CAACACCACA CAGTTTGG TAGTTACATA GAATAGTACA GACCTAGCTA

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FIG. 10L

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          haeIII/palI
          haeI
          fnu4HI styI
          mseI   ncoI
          aseI/asnI/vspI   dsaI
          xmnI   hinPI   bsaJI
          asp700   hhaI/cfoI   nlaIII
          3701 CGGAATTAA TTCGGCGCAG CACCATGGCC TGAATAAACC TCTGAAAGAG GAACTTGGTT AGGTACCTTC TGAGGCGGAA AGAACCACT GTGAATGTG
              GCCTTAATT AAGCCGCGTC GTGTACCGG ACTTTATTGG AGACTTTCTC CTGGAACCAA TCCATGGAAG ACTCGCCTT TCTTGGTGA CACTTACAC
              rsal
              csp6I
              nlaIV
              kpnI
              hgiCI
              banI
              asp718   mnli
              acc65I   ddeI   aciI
              nspBII
              aluI
              pvuII
              nspBII
              3801 TGTCAGTTAG GGTGTGGA   GTCCCGAGC   TCCCGAGC   GCAGAAGTAT   GCAAGCATG   CATCTCAATT   AGTCAGCAAC   CAGGTGTGGA   AAGTCCCGCAG
              ACAGTCAATC   CCACACCTT   CAGGGGTCCG   AGGGGTGCTC   CGTCTTCATA   CGTTTCGTAC   GTAGAGTTAA   TCAGTCGTTG   GTCCACACCT   TTCAGGGGTC
              nlaIV
              scrFI
              mvaI
              ecorII
              dsaV
              bstNI
              apyI[dcM+]
              bsaJI
              ppulOI
              nsII/avaIII
              nlaIII
              sphI
              nspI
              nspHI
              3901 GCTCCCCAGC AGGCAGAGT ATGCAAGCA TGCACTCTCA   TGATCAGCA ACCATAGTCC CGCCCTTAAC TCCGCCATC CGCCCTTAA CTCGCCCGCAG
              CGAGGGGTG   TCCGTCTTCA   TACGTTCTG   ACGTAGATT   AATCAGTCT   TGGTATCAG   GCGGGGATT   AGCGGGTAG   GCGGGGATT   GAGCGGGTC
              aciI
              fokuI   aciI
              bsrI
              fnu4HI
              bglI
              sfiI
              haeIII/palI
              mnli
              ddeI
              haeIII/palI   bsaJI   mnli   aluI
              mnli   bsaJI   aciI   haeIII/palI
              4001 TTCCGCCCAT TCTCCGCC   ATGGCTGACT   AATTTTTTTT   ATTATGCA   AGCCGAGC   CGCTCGGC   TCTGAGCTAT   TCCAGAAGTA   GTGAGGAGGC
              AAGCGGGTA   AGAGCGGG   TACCGACTGA   TTAATAAATA   TAAATACGTC   TCCGGTCCG   GCGGAGCCG   AGACTCGATA   AGTCTTCAT   CACTCTCCG
              aciI
              bslI   dsaI
              aciI   bsaJI
              mnli
              mnli

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FIG. 10M

[illegible]

ml

SUBSTITUTE SHEET (RULE 26)

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FIG. 10P

sau3AI
 mboI/ndeII(dam-)
 dpnI(1am+)
 dpnII(dam-)
 lstyI/xhoII
 bsrI
 tagI alwI(dam-) acII nspBII
 taqI alwI(dam-) acII bstYI/xhoII
 5401 TCGAACTGGA TCTCAACAGC GGTAAAGATCC TTGAGAGTTT TCGCCCCGAA GAACGTTTT CAATGATGAG CACTTTTAA GTTCTGCTAT GTGGCGCGGT
 AGCTTGACCT AGAGTTGTCG CCATTCTAGG AACTCTCAA AGCGGGGCTT CTGCAAAAG GTTACTACTC GTCAAAATTT CAAGACGATA CACCGGCCA
 scrFI
 nciI
 mspI
 hpaII
 dsav
 caulI
 hinII/acyI
 hgaI
 ahaII/bseHI
 5501 ATTATCCGT GATCAGCCG GGCAAGAGCA ACTCGTGC ACTGCATACACT ATTCTCAGAA TGACTTGGTT GACTACTCAC CAGTCACAGA AAGCATCTT
 TAATAGGCA CTACTGCGC CGTTCTCGT TGAGCCAGCG GCGTATGTGA TAAGAGTCTT ACTGAACCAA CTCATGAGTG GTCAGTGCT TTTCGTAGAA
 rsaI
 csp6I bsrI
 scaI hphI maeIII
 5601 ACGGATGCA TGACAGTAAG AGAATTATGC AGTGCTGCCA TAACCATGAG TGATAACACT GCGGCCAACT TACTTCTGAC AAGCATCGA GGACCGAAGG
 TGCCTACCGT ACTGTCAATC TCTTAATAG TCACGACCGT ATTGCTACTC ACTATTGTA CGCGGTTGA ATGAAGACTG TTGCTAGCCT CCTGCTTCC
 foki nlaIII
 fnu4HI
 bbvI
 nlaIII
 sau3AI maeIII
 mboI/ndeII(dam-)
 dpnI(1am+)
 alwI(dam-)
 nlaIII dpnII(dam-)
 5701 AGCTAACCGC TTTTTCAC AACATGGGG ATCATGTAAC TCGCTTGTAT CGTTGGGAA CCGAGCTGAA TGAAGCCATA CCAACGAGC AGCTGACAC
 TCGATTGGCG AAAAACGCTG TTGTACCCCT TAGTACATTT AGCGGAACTA GCACCCCTTG GCCTGACTT ACTTCGGTAT GCTTCTGCTC TCGCACTGTC
 aluI acII
 maeIII
 sau3AI
 mboI/ndeII(dam-)
 dpnI(1am+)
 alwI(dam-)
 nlaIII dpnII(dam-)
 bstYI/xhoII
 dpnII(dam-)
 asp700
 xmnI
 psp1406I
 hgaI/aspHI
 bsp1286 tru9I
 bsiHKA mseI
 bmyI ahaIII/draI
 hbaI/cfoI
 hinPI
 bsh1236I
 bstUI
 fnuDII/mvnI
 thaI
 acII

[illegible]

[illegible]

FIG. 10S

[illegible]

```

scrFI
mvaI
ecorII
dsav
bstNI
apyI{dcm+}
bsaJI
CACCCCAGGC TTTC
GTGGGGTCCG AAAC

tru9I
mseI
aseI/asnI/vspI
ATTA
TAATT

```

	apyIIdcm+]	mspI	aciI	bsrBI	alul	nlaIII	xmnI
7201	CACCCCAGGC TTTACACTTT ATGCTTCCGG CTCGTATGTT GTGTGGAATT GTGAGCGGAT AACAAATTCA CACAGAAAC AGCATGACC ATGATTACGA	bsaJI	hpaII				asp700
	GTGGGGTCCG AAATGCGAAA TACGAAGGCC GAGCATACAA CACACCTTAA CACTGCCTTA TTGTTAAAGT GTGTCCTTTG TCATACTGCG TACTAAATGT						

>length: 7305

INTERNATIONAL SEARCH REPORT

International Application No

PC1/US 95/09576

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/64 C12N15/67 C12N15/85 C12N9/72 C12N5/10

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DNA CLONING, VOLUME III, EDITED BY D.M. GLOVER, 1987 IRL PRESS, OXFORD, GB;; pages 189-212, A.M.C. BROWN AND M.R.D. SCOTT 'Retroviral vectors'	1-3,7,8
Y	see page 192, line 7 - page 196, line 5; figures 2,3 --- -/--	5,6, 9-12, 16-21

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

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Date of the actual completion of the international search

23 November 1995

Date of mailing of the international search report

08.12.95

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Authorized officer

Hornig, H

INTERNATIONAL SEARCH REPORT

Int. Application No
PCT/US 95/09576

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	CELL, vol. 37, no. 3, July 1984 CELL PRESS, CAMBRIDGE, MA, US;, pages 1053-1062, C.L. CEPKO ET AL. 'Construction and applications of a highly transmissible murine retrovirus shuttle vector' cited in the application	1-3,7,8
Y	pZIP-Neo SV(B)1 see figure 1	5,6, 9-12, 16-21
Y	--- MOL. CELL. BIOL., vol. 5, no. 3, March 1985 ASM WASHINGTON, DC, US, pages 431-437, A.D. MILLER ET AL. 'Generation of helper-free amphotrophic retroviruses that transduce a dominant-acting, methotrexate-resistant dihydrofolate reductase gene' see page 432, right column, line 5 - page 436, right column, line 7; figure 1	5,6, 9-12, 16-21
Y	--- WO,A,94 05784 (US) 17 March 1994 see the whole document	5,6, 9-12, 16-21
Y	--- EP,A,0 215 548 (ZYMOGENETICS INC ;UNIV WASHINGTON (US)) 25 March 1987 see the whole document	5,6, 9-12, 16-21
A	--- WO,A,92 17566 (GENENTECH INC) 15 October 1992 cited in the application see the whole document	1-21
A	--- WO,A,90 12025 (UNIV LELAND STANFORD JUNIOR) 18 October 1990 cited in the application see the whole document	1-21
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A	--- EP,A,0 160 457 (GENENTECH INC) 6 November 1985 cited in the application see the whole document	1-21
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INTERNATIONAL SEARCH REPORT

Int ional Application No
PCT/US 95/09576

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>PROC. NATL.ACAD SCI., vol. 86, February 1989 NATL. ACAD SCI., WASHINGTON, DC, US;, pages 1041-1045, M. VIVAUD ET AL. 'A 5' splice-region G-C mutation in exon 1 of the human beta-globin gene inhibits pre-mRNA splicing: A mechanism for beta+-thalassemia' see the whole document -----</p>	1-4

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No
PCT/US 95/09576

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
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		DE-A- 3688900	23-09-93
		DE-T- 3688900	09-12-93
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EP-A-0260148	16-03-88	AU-B- 613316	01-08-91
		AU-B- 7831787	19-05-88
		DE-A- 3730599	07-07-88
		FR-A- 2603899	18-03-88
		GB-A, B 2197321	18-05-88
		US-A- 5024939	18-06-91
		JP-A- 63152986	25-06-88
EP-A-0160457	06-11-85	AU-B- 601358	13-09-90
		AU-B- 4134585	24-10-85
		AU-B- 5295890	30-08-90
		EP-A- 0385558	05-09-90
		HK-A- 8395	27-01-95
		JP-A- 60243023	03-12-85
		JP-A- 6040942	15-02-94
		NO-B- 174934	25-04-94
		SG-A- 3994	10-06-94
		US-A- 4965199	23-10-90